

# Functionally Defective High-Density Lipoprotein: A New Therapeutic Target at the Crossroads of Dyslipidemia, Inflammation, and Atherosclerosis

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**Abstract**—High-density lipoproteins (HDL) possess key atheroprotective biological properties, including cellular cholesterol efflux capacity, and anti-oxidative and anti-inflammatory activities. Plasma HDL particles are highly heterogeneous in physico-chemical properties, metabolism, and biological activity. Within the circulating HDL particle population, small, dense HDL particles display elevated cellular cholesterol efflux capacity, afford potent protection of atherogenic low-density lipoprotein against oxidative stress and attenuate inflammation. The antiatherogenic properties of HDL can, however be compromised in metabolic diseases associated with accelerated atherosclerosis. Indeed, metabolic syndrome and type 2 diabetes are characterized not only by elevated cardiovascular risk and by low HDL-cholesterol (HDL-C) levels but also by defective HDL function. Functional HDL deficiency is intimately associated with alterations in intravascular HDL metabolism and structure. Indeed, formation of HDL particles with attenuated antiatherogenic activity is mechanistically

related to core lipid enrichment in triglycerides and cholesteryl ester depletion, altered apolipoprotein A-I (apoA-I) conformation, replacement of apoA-I by serum amyloid A, and covalent modification of HDL protein components by oxidation and glycation. Deficient HDL function and subnormal HDL-C levels may act synergistically to accelerate atherosclerosis in metabolic disease. Therapeutic normalization of attenuated antiatherogenic HDL function in terms of both particle number and quality of HDL particles is the target of innovative pharmacological approaches to HDL raising, including inhibition of cholesteryl ester transfer protein, enhanced lipidation of apoA-I with nicotinic acid and infusion of reconstituted HDL or apoA-I mimetics. A preferential increase in circulating concentrations of HDL particles possessing normalized antiatherogenic activity is therefore a promising therapeutic strategy for the treatment of common metabolic diseases featuring dyslipidemia, inflammation, and premature atherosclerosis.

## I. Introduction

According to the recent estimates of the World Health Organization, approximately one-third of all deaths (16.7 million people) around the globe resulted from cardiovascular (CV<sup>1</sup>) disease in 2002 (World Health Organization, 2004). As shown in the recent INTERHEART study, which enrolled 29,972 subjects in 52 countries worldwide, the most strongly predictive CV risk factors for myocardial infarction were dyslipidemia, smoking, hypertension, diabetes, abdominal obesity, psychosocial factors, consumption of fruits, vegetables, and alcohol, and lack of regular physical activity (Yusuf et al., 2004). Collectively, these factors accounted for

most ( $\geq 90\%$ ) of the risk of myocardial infarction in both sexes and at all ages in all regions.

Atherosclerosis represents the pathological process that typically underlies CV morbidity and mortality, formation of plaques in the intima and media of the arterial wall. Such atherosclerotic plaques result from the progressive accumulation of cholesterol and diverse lipids in native and oxidized forms, extracellular matrix material, and inflammatory cells. Atherogenic dyslipidemia, a highly prominent CV risk factor, is intimately associated with premature atherosclerosis and corresponds to an imbalance between excess circulating levels of cholesterol in the form of pro-atherogenic apolipoprotein (apo) B-containing lipoproteins compared with subnormal levels of antiatherogenic apoA-I-containing lipoproteins (Fig. 1). Indeed, apoB is the predominant protein component of proatherogenic, cholesterol-rich low-density lipoprotein (LDL), triglyceride (TG)-rich very-low density lipoproteins (VLDL), VLDL remnants

<sup>1</sup> Abbreviations: CV, cardiovascular; apo, apolipoprotein; LDL, low-density lipoprotein(s); TG, triglyceride(s); VLDL, very low-density lipoprotein(s); IDL, intermediate-density lipoprotein(s); HDL, high-density lipoprotein(s); LDL-C, LDL-cholesterol; HDL-C, HDL-cholesterol; CHD, coronary heart disease; MetS, metabolic syndrome; CAD, coronary artery disease; LOOH, lipid hydroperoxide(s); PL, phospholipid(s); NOS, nitric oxide synthase; oxLDL, oxidized LDL; CRP, C-reactive protein; RCT, reverse cholesterol transport; CE, cholesteryl ester; LCAT, lecithin/cholesterol acyltransferase; PAF-AH, platelet-activating factor-acetyl hydrolase; PON 1, paraoxonase 1; GSPx, glutathione selenoperoxidase; SAA, serum amyloid A; LpA-I, lipoprotein particles containing only apoA-I; LpA-I/A-II, lipoprotein particles containing both apoA-I and apoA-II; ABC, ATP-binding cassette transporter; PLTP, phospholipid transfer protein; CETP, cholesteryl ester transfer protein; SR-BI, scavenger receptor type BI; HL, hepatic lipase; ROS, reactive oxygen species; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; IL, interleukin; rHDL, reconstituted HDL; S1P, sphingosine-1-phosphate; hs, high sensitivity; FH, familial hypercholesterolemia; NEFA, nonesterified fatty acid(s); IMT, intima-media thickness; VA-HIT, Veterans Affairs High-Density Lipoprotein Cholesterol Intervention Trial; AFREGS, Armed Forces Regression Study; BIP, Bezafibrate Infarction Prevention Trial; HATS, HDL-Atherosclerosis Treatment Study; ARBITER, Arterial Biology for the Investigation of the Treatment Effects of Reducing Cholesterol; JTT-705, S-[2-([1-(2-ethylbutyl)cyclohexyl]carbonyl)-amino]phenyl]2-methylpropanoic acid; FIELD, Fenofibrate Intervention and Event Lowering in Diabetes; REVERSAL, Reversal of Atherosclerosis with Aggressive Lipid Lowering.

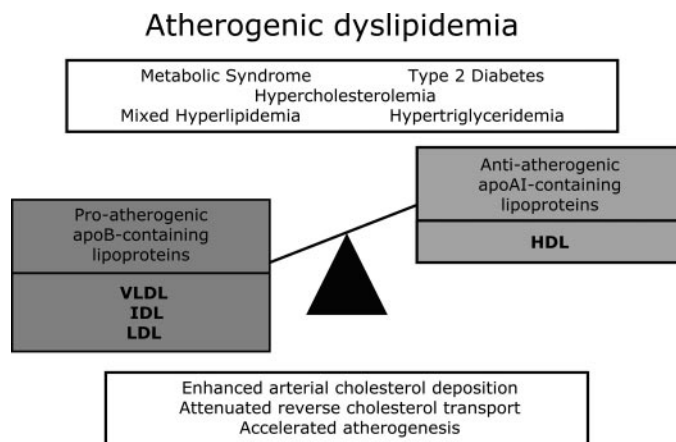


FIG. 1. Atherogenic dyslipidemia as an imbalance between circulating levels of proatherogenic apoB-containing lipoproteins and antiatherogenic apoA-I-containing lipoproteins.

and intermediate-density lipoprotein (IDL), whereas apoA-I is the major protein component of antiatherogenic high-density lipoprotein (HDL). In the INTERHEART study, dyslipidemia was assessed as an elevated ratio of plasma levels of proatherogenic apoB to antiatherogenic apoA-I ( $\geq 5:1$ ) (Yusuf et al., 2004), and as such, represented a direct estimate of atherogenic potential in any individual.

Elevated circulating concentrations of LDL-cholesterol (LDL-C) occur frequently as hypercholesterolemia, a common form of atherogenic dyslipidemia (Wilson, 1990). LDL is the major vehicle for transport of cholesterol not only to peripheral tissues but also to the arterial wall (Lusis, 2000); indeed, ionic interaction of positively charged domains of apoB with negatively charged proteins of the extracellular matrix, including proteoglycans, collagen, and fibronectin, leads to intimal retention of apoB-containing lipoproteins, a major initiating factor in atherogenesis (Khalil et al., 2004).

Among factors other than LDL-C that are associated with dyslipidemia, a low level of HDL-cholesterol (HDL-C) is now most recognized (Gotto and Brinton, 2004). Several prospective epidemiological studies, including the Framingham Heart Study, US Physicians' Health Study, Prospective Cardiovascular Münster (PROCAM) Study, and Atherosclerosis Risk in Communities (ARIC) Study, have found that low serum HDL-C concentrations (defined as  $<40$  mg/dl in both sexes or as  $<40$  mg/dl in men and  $<50$  mg/dl in women) (Chapman et al., 2004) constitute an independent risk factor for coronary heart disease (CHD) in both nondiabetic and diabetic subjects (Maron, 2000; Sharrett et al., 2001; Gotto and Brinton, 2004). Moreover, low HDL-C is characteristic of atherogenic dyslipidemia and increased CV risk in patients with metabolic diseases such as type 2 diabetes and metabolic syndrome (MetS). In this context, it is of special relevance that the World Health Organization has estimated that the population of individuals with type 2 diabetes will have increased worldwide to 250 millions or more by 2025 (World Health Organization, 2004).

Prospective studies have revealed that CHD risk is elevated by 3% in women and 2% in men for each decrement of 1 mg/dl in HDL-C (Wilson, 1990). Conversely, a decreased risk of CV events is frequently observed in subjects with elevated HDL-C levels (Maron, 2000; Doggen et al., 2004; Gotto and Brinton, 2004); in addition, high concentrations of HDL-C ( $>60$  mg/dl) are typically associated with longevity (Barzilai et al., 2003; Barter, 2004). The prevalence of low HDL-C levels can vary from 20% in a general population to up to 60% in patients with established CHD (Franceschini, 2001). Not only are low HDL-C levels associated with an increased incidence of CHD but also with a greater risk for carotid atherosclerosis and ischemic stroke mortality and with a more aggressive progression of angiographically defined coronary artery disease (CAD) (Maron, 2000; Gotto and

Brinton, 2004). Finally, it is noteworthy that in the recent Myocardial Ischemia Reduction with Aggressive Cholesterol Lowering (MIRACL) trial in patients with acute coronary syndromes treated with atorvastatin, baseline HDL-C levels, rather than those of LDL-C, predicted the occurrence of CV events (Olsson et al., 2005).

#### A. Inflammation and Oxidative Stress in the Progression of Atherosclerosis

The imbalance between circulating levels of cholesterol transported in HDL relative to that in apoB-containing particles is intimately associated with induction of both endothelial dysfunction and oxidative stress in the arterial wall, which are in turn closely related to inflammation (Chisolm and Steinberg, 2000; Lusis, 2000); as a result, dyslipidemia, oxidative stress, and inflammation are closely interrelated in the development of atherosclerosis.

Oxidative stress is defined as an imbalance between prooxidant and antioxidant factors in favor of prooxidants and is central to the pathophysiology of atherosclerosis and CV disease (Fig. 2). Analysis of plaque composition has revealed products of protein and lipid oxidation, such as oxidized, chlorinated, and nitrated amino acids, lipid hydroperoxides (LOOH), short-chain aldehydes, oxidized phospholipids (PL), F $2\alpha$ -isoprostanes, and oxysterols, thereby suggesting the presence of local oxidative stress (Heinecke, 1998). The preferential retention of LDL in the arterial wall makes this lipoprotein a major substrate for oxidation by prooxidants produced by arterial wall cells. Various oxidative systems potentially contribute to LDL oxidation in vivo, and these include NAD(P)H oxidases, xanthine oxidase, myeloperoxidase, uncoupled nitric oxide synthase (NOS), lipoxygenases, and the mitochondrial electron transport chain (Madamanchi et al., 2005; Mueller et al., 2005). Accordingly, reactive oxygen, chlorine and nitrogen species, and lipid-derived free radicals are major prooxidants involved in the formation of oxidized LDL

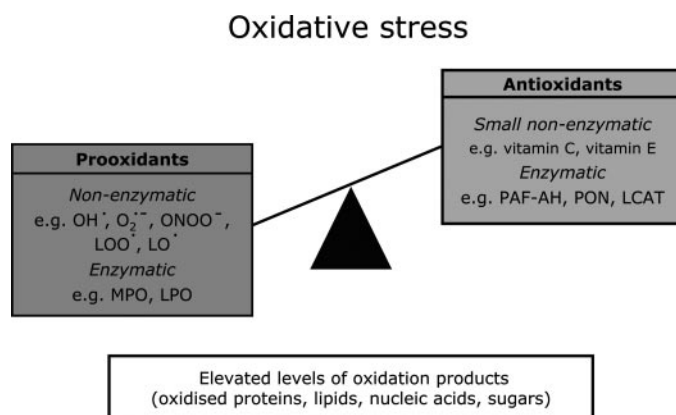


FIG. 2. Oxidative stress as an imbalance between prooxidant and antioxidant factors in favor of prooxidants. LPO, lipoxygenase; MPO, myeloperoxidase.

(oxLDL) in vivo. Significantly, production of both chlorine- and nitrogen-containing prooxidants is increased at sites of inflammation (Heinecke, 1998), suggesting that focal inflammation significantly contributes to the initiation of LDL oxidation at early stages of plaque formation. Consistent with the role of oxidative stress and oxidative modification of LDL in atherosclerosis, both urinary levels of F<sub>2</sub> $\alpha$ -isoprostanes, currently the most robust and integrative marker of oxidative stress in vivo in humans (Morrow, 2005) and plasma levels of oxLDL constitute strong and independent risk factors for CHD (Schwedhelm et al., 2004; Meisinger et al., 2005).

Inflammation is a systemic body response aimed to decrease the toxicity of harmful agents and repair damaged tissue. Chronic inflammation, which may be measured as circulating levels of an acute-phase protein, such as C-reactive protein (CRP), represents a major CV risk factor (Ridker et al., 2004b; Willerson and Ridker, 2004; Verma et al., 2005). A key feature of the inflammatory response involves activation of phagocytic cells involved in host defense, which produce an oxidative burst of reactive oxygen, chlorine, and nitrogen species, with subsequent creation of a highly prooxidative environment to combat invading pathogens. Local and systemic infections, arterial wall injury, and excessive retention of LDL may all potentiate activation of macrophages in the arterial wall, thereby triggering excessive production of prooxidant species (Hansson, 2005). As a result, oxidation of proteoglycan-bound LDL may occur in the extracellular space of the arterial intima (Memon et al., 2000).

OxLDL particles exhibit multiple atherogenic properties, which include uptake and accumulation in macrophages, as well as proinflammatory, immunogenic, apoptotic, and cytotoxic activities (Chisolm and Steinberg, 2000). In contrast to unmodified LDL, oxLDL is taken up through macrophage scavenger receptor pathways that are not down-regulated by excess ligand and lead to the formation of cholesterol-loaded foam cells, characteristic components of atherosclerotic plaques. The proinflammatory activities of oxLDL include chemoattractant action on circulating monocytes, induction of the expression of adhesion molecules on endothelial cells, promotion of monocyte differentiation into macrophages, induction of the production and release of proinflammatory cytokines and chemokines from macrophages, and inhibition of macrophage motility (Chisolm and Steinberg, 2000; Lusis, 2000). Most of the proinflammatory properties of oxLDL arise from products of LDL lipid peroxidation, such as 1-palmitoyl-2(5-oxovaleryl)-sn-glycero-3-phosphorylcholine, 1-palmitoyl-2-glutaryl-sn-glycero-3-phosphorylcholine, cholesteryl linoleate hydroperoxide, 7 $\beta$ -hydroperoxycholesterol, hydroxyoctadecadienoic acid, and 4-hydroxynonenal (Chisolm and Steinberg, 2000; Lusis, 2000; Van Lenten et al., 2001a). As a result, LDL oxidation further propagates the inflammatory process in the arterial wall, thereby accelerating

atherogenesis (Lusis, 2000). Atherosclerosis can therefore be regarded as a chronic inflammatory disease of the arterial wall mediated by oxLDL in concert with a spectrum of additional proinflammatory agents.

HDL particles are distinguished from atherogenic apoB-containing lipoproteins by their capacity to exert a wide spectrum of antiatherogenic biological activities, including 1) their capacity to mediate cellular cholesterol efflux by acting as primary acceptors, thereby facilitating reverse cholesterol transport (RCT) from the arterial wall and peripheral tissues to the liver, 2) the protection of LDL against oxidative stress, 3) anti-inflammatory actions on arterial wall cells, and 4) anti-apoptotic, 5) vasodilatory, 6) antithrombotic, and 7) anti-infectious activities. In this review, we will consider recent evidence for the heterogeneity of the atheroprotective properties of HDL particle subpopulations with emphasis on their ability both to protect against accumulation of lipids and to attenuate oxidative stress and inflammation in the arterial wall. Furthermore, new findings on functionally defective HDL will be discussed in the context of metabolic diseases associated with elevated CV risk; these data indicate that the potent antiatherogenic activities of small, dense HDL particles are impaired in the dyslipidemic and inflammatory state associated with type 2 diabetes and MetS. Finally, we will critically appraise innovative therapeutic strategies to normalize defective functionality of small, dense HDL particles; these exciting developments open new horizons for the treatment of atherogenic dyslipidemia in metabolic disease.

## II. Functional High-Density Lipoprotein

### A. Structure, Composition, and Heterogeneity

Functional plasma HDL are spherical or discoidal particles of high hydrated density (1.063–1.21 g/ml) due to elevated protein content (>30% by weight) compared with other lipoproteins (Fig. 3) (Asztalos and Schaefer, 2003; Barter et al., 2003b). Discoidal HDL are small particles consisting primarily of apoA-I embedded in a lipid monolayer constituted of PL and free cholesterol (Segrest et al., 1999, 2000). Spherical HDL are larger and additionally contain a hydrophobic core formed by cholesteryl esters (CE) and small amounts of TG. ApoA-I (molecular mass 28 kDa) is the major structural HDL apolipoprotein and accounts for ~70% of total HDL protein, whereas the second major HDL apolipoprotein, apoA-II, represents ~20%. Minor HDL protein components (typically <10% of the HDL protein moiety) include apoE, apoA-IV, apoA-V, apoJ, apoC-I, apoC-II, and apoC-III (Asztalos and Schaefer, 2003; Barter et al., 2003b; Karlsson et al., 2005). In small discoidal HDL, two molecules of apoA-I adopt a “double belt” orientation with their helices oriented parallel to the plane of the disc and perpendicular to the lipid acyl chains in such a way that they wrap around the lipid bilayer disc forming

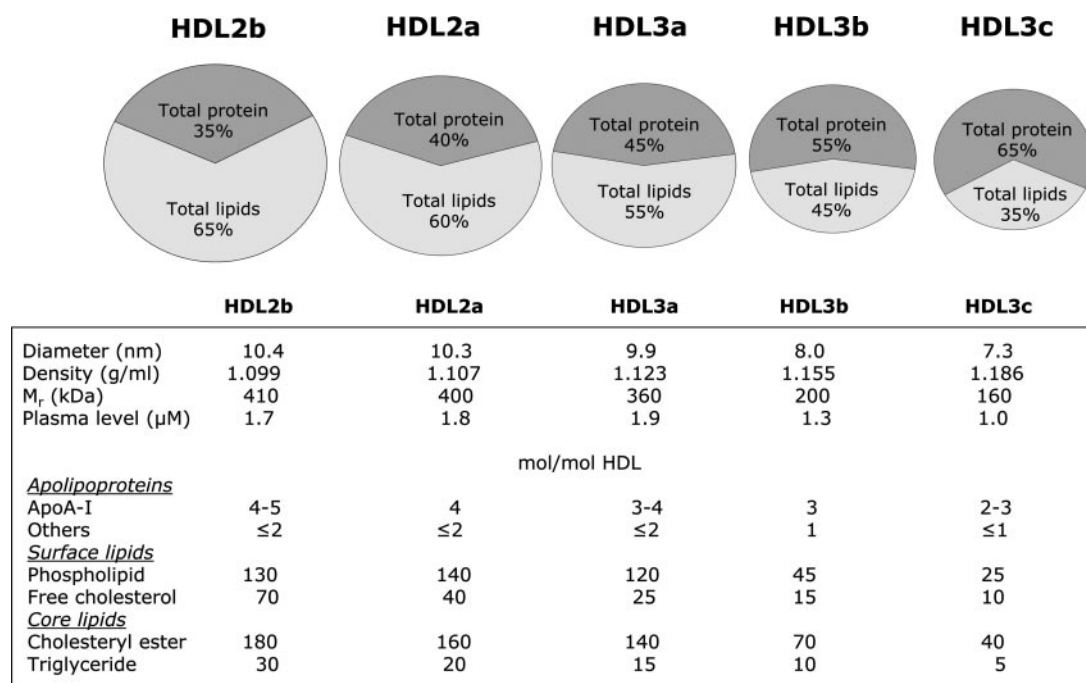


FIG. 3. Heterogeneity in the physicochemical properties of normal functional HDL in healthy normolipidemic subjects (Blanche et al., 1981; Barter et al., 1999; Kontush et al., 2003).

two stacked rings in an antiparallel orientation (Segrest et al., 1999, 2000; Silva et al., 2005); furthermore, apoA-I molecules appear to slide in relation to each other between two stable conformations (Silva et al., 2005). Plasma HDL particles also carry enzymes involved in lipid metabolism, including lecithin/cholesterol acyltransferase (LCAT), enzymes with plausible antioxidative activities, such as platelet-activating factor-acetyl hydrolase (PAF-AH, also called lipoprotein-associated phospholipase A<sub>2</sub>), paraoxonase 1 (PON1) and glutathione selenoperoxidase (GSPx) (Navab et al., 2004b), and other proteins and peptides, such as serum amyloid A (SAA), a major positive acute-phase reactant (Uthlar and Whitehead, 1999),  $\alpha$ -1-antitrypsin, a potent inhibitor of serine proteinases (Karlsson et al., 2005), or amyloid- $\beta$ , the principal constituent of senile plaques in Alzheimer's disease (Kontush, 2004).

Plasma HDL particles are highly heterogeneous in their physicochemical properties, metabolism, and biological activity (Fig. 3) (Asztalos and Schaefer, 2003; Barter et al., 2003b). Such heterogeneity results from differences in relative contents of apolipoproteins and lipids in HDL and is intimately related to the amphipathic helical structure of human apoA-I (Reschly et al., 2002; Maiorano et al., 2004); these helices possess a hinge domain that allows apoA-I to switch between two conformations corresponding to HDL particles of different size. When fractionated by ultracentrifugation, human HDL is typically separated into two major subfractions, HDL2 ( $d$  1.063–1.125 g/ml) and HDL3 ( $d$  1.125–1.21 g/ml) (Chapman et al., 1981). Nondenaturing polyacrylamide gradient gel electrophoresis has been

used to separate HDL into five distinct subpopulations of decreasing size, HDL2b, 2a, 3a, 3b, and 3c (Anderson et al., 1977); equivalent subpopulations can be quantitatively isolated using isopycnic density gradient ultracentrifugation (Fig. 3) (Tall et al., 1982; Goulinet and Chapman, 1997; Guerin et al., 2000a). Other separation methods, such as two-dimensional electrophoresis, allow identification of more than 10 HDL subspecies in which spherical  $\alpha$ -HDL predominate (Asztalos et al., 1993; Asztalos and Schaefer, 2003); each subspecies may, however, be heterogeneous in physicochemical properties, as in the case of ultracentrifugally isolated subfractions. HDL can also be immunoseparated on the basis of apolipoprotein composition into particles containing only apoA-I (LpA-I) and both apoA-I and apoA-II (LpA-I/A-II) (Duriez and Fruchart, 1999). In most human subjects, apoA-I is distributed approximately equally between LpA-I and LpA-I/A-II, whereas virtually all apoA-II is in LpA-I/A-II. Finally, ultrafiltration (Atmeh, 1990; Atmeh and Abd Elrazeq, 2005) and size-exclusion chromatography (Nanjee and Brinton, 2000) allow isolation of small, protein-rich HDL particles of low molecular mass (40–70 kDa). Given the complexity of HDL particle heterogeneity, small, dense HDL will be defined for present purposes as lipid-poor and protein-rich discoidal and spherical HDL particles of small size ( $\leq 9$  nm), low molecular mass ( $\leq 200$  kDa), and high density (1.125–1.24 g/ml). Depending on the fractionation method, small, dense HDL may include HDL3a, 3b, and 3c and very high-density lipoprotein separated by ultracentrifugation and pre- $\beta$ -HDL separated by gradient gel electrophoresis.

On a particle basis, HDL are the most numerous per unit volume of plasma and are present at the highest (micromolar) levels compared with other lipoproteins. Concentrations of major HDL2 and HDL3 subfractions typically are in the range of 2 to 6  $\mu\text{M}$  corresponding to 50 to 200 mg total mass/dl (Kontush et al., 2003, 2004, 2005; Hansel et al., 2004; Nobecourt et al., 2005).

The clinical relevance of circulating levels of individual HDL subfractions to atherosclerosis and CV disease is, however, unclear. Concentrations of HDL2-C and HDL3-C as estimates for plasma levels of the two major HDL subfractions were measured in several studies, which differed in separation methods (polyanion precipitation versus ultracentrifugation). Conflicting results were obtained, with evidence that either HDL2-C or HDL3-C constitutes a strong predictor of CHD or CV risk factors (Johansson et al., 1991; Drexel et al., 1992, 1994, 1996; Skinner, 1994; Robins et al., 2001; Alagona et al., 2002; Barter et al., 2003a; Yu et al., 2003; Desai et al., 2005). Furthermore, plasma levels of either large (Rosenson et al., 2002) or small (Mackey et al., 2002) HDL were reported to be associated with the progression of coronary atherosclerosis. Similarly controversial is the clinical significance of pre- $\alpha$ -HDL, pre- $\beta$ -HDL, and LpA-I/A-II levels. By contrast, plasma levels of  $\alpha$ 1-HDL and LpA-I are typically associated with protection from atherosclerosis (Duriez and Fruchart, 1999; Asztalos et al., 2003; Asztalos and Schaefer, 2003).

Discordance in these data reflects complex relationships between HDL subfractions separated by different methods. For example, immunisolated LpA-I/A-II is found predominantly in the HDL3 density range, whereas LpA-I is a prominent component of both HDL2 and HDL3 (Duriez and Fruchart, 1999). On the other hand,  $\alpha$ -migrating HDL predominate in both HDL2 and HDL3 subfractions, whereas pre- $\beta$ -HDL coisolates with small, dense HDL particles (Asztalos and Schaefer, 2003). Another important example concerns ultracentrifugally isolated small, dense HDL3c, which does not precisely correspond to small HDL subpopulations as determined by other methods. Human HDL3c represents a minor subfraction accounting for approximately 6% of total HDL mass and 10% of apoA-I (Kontush et al., 2003, 2004, 2005; Hansel et al., 2004; Nobecourt et al., 2005). In two-dimensional electrophoresis, HDL3c reveals further heterogeneity and produces multiple signals corresponding to small  $\alpha$ 3-, pre $\beta$ 3- and pre $\beta$ 1-HDL (S. Chantepie, A. Kontush, and M. J. Chapman, unpublished data). By contrast, small HDL  $\alpha$ 3, pre $\beta$ 3, and pre $\beta$ 1 subfractions measured by two-dimensional electrophoresis in whole plasma account for approximately 37, 4, and 12% of apoA-I, respectively (Asztalos et al., 2004a); moreover,  $\alpha$ 3 together with  $\alpha$ 2 represent two major HDL subfractions in normolipidemic subjects. It is essential to emphasize that routine clinical measurement of plasma HDL-C primarily reflects levels of

large, cholesterol-rich HDL particles and frequently lacks sensitivity to detect small cholesterol-poor HDL.

### B. Metabolism

Spherical plasma HDL are mature particles generated by intravascular processes from lipid-free apoA-I or lipid-poor pre- $\beta$ -HDL (Fig. 4) (Rye and Barter, 2004). These small HDL precursors are produced as nascent HDL by the liver or intestine, are also released as surface fragments from lipolysed TG-rich lipoproteins (VLDL and chylomicrons), and finally may be generated during the interconversion of HDL3 and HDL2 (von Eckardstein et al., 2001). Small nascent HDL are unstable and readily acquire lipids (Atmeh and Abd Elrazeq, 2005); their initial lipidation occurs at cellular membranes via the ATP-binding cassette transporter (ABC) A1-mediated efflux of cholesterol and PL from cells (Fig. 4) (Oram, 2002). ABCA1 is a major player in HDL metabolism; indeed, genetic defects in ABCA1 as occur in Tangier disease may result in low HDL-C levels, with cholesterol accumulation in peripheral tissues and premature atherosclerosis (Oram, 2002).

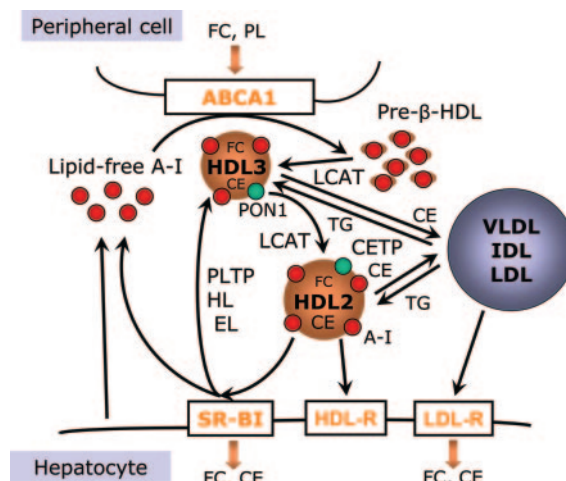


FIG. 4. Intravascular HDL particle remodeling and metabolism in normolipidemia. Spherical plasma HDL are generated from lipid-free apoA-I or lipid-poor pre- $\beta$ -HDL, which are produced as nascent HDL by the liver or intestine but can also be released as surface fragments from lipolysed TG-rich lipoproteins and/or during the interconversion of HDL3 and HDL2. Initial lipidation of small nascent HDL occurs at cellular membranes via the ABCA1-mediated efflux of cholesterol and PL from cells. Subsequent LCAT-mediated cholesterol esterification generates large spherical HDL2 particles, which undergo further remodeling via particle fusion and surface remnant transfer mediated by PLTP. Large HDL2 can be converted in turn to small HDL3 upon CETP-mediated transfer of CE from HDL to apoB-containing lipoproteins, upon SR-BI-mediated selective uptake of CE by the liver and steroidogenic organs, and HL- and endothelial lipase-mediated hydrolysis of TG. When CETP-mediated transfer of CE occurs between HDL and TG-rich lipoproteins, TG-rich HDL are generated, which can be further hydrolyzed by HL to small, TG-rich HDL particles. The concerted action of CETP and HL promotes reduction in HDL size, formation of lipid-poor HDL particles, and shedding from HDL of lipid-free apoA-I, which can interact with ABCA1 in the next lipidation cycle. HDL lipids are catabolized either separately from HDL proteins by selective uptake or via CETP transfer or as holoparticles together with HDL proteins primarily in the liver via uptake through LDL receptors for apoE-containing HDL and through hitherto unidentified receptors for HDL holoparticles. EL, endothelial lipase; FC, free cholesterol; HDL-R, HDL holoparticle receptor; LDL-R, LDL receptor.

Subsequent LCAT-mediated esterification of cell-derived cholesterol generates large spherical HDL2 particles with a neutral lipid core of CE and TG (Jonas, 2000); such particles undergo further remodeling via particle fusion and surface remnant transfer mediated by phospholipid transfer protein (PLTP) (van Tol, 2002). Large HDL2 can be converted in turn to small HDL3 upon cholesteryl ester transfer protein (CETP)-mediated transfer of CE from HDL to apoB-containing lipoproteins, upon scavenger receptor type BI (SR-BI)-mediated selective uptake of CE by the liver and steroidogenic organs and hepatic lipase (HL) and upon endothelial lipase-mediated hydrolysis of core TG (Fig. 4) (von Eckardstein et al., 2001). When CETP-mediated transfer of CE occurs between HDL and TG-rich lipoproteins, TG-rich HDL are generated (Le Goff et al., 2004), which can be further hydrolyzed by HL to small, TG-rich HDL particles (Santamarina-Fojo et al., 2004). The concerted action of CETP and HL promotes reduction in HDL size, formation of lipid-poor HDL particles and shedding from HDL of lipid-free apoA-I, which can interact with ABCA1 in the next lipidation cycle (Clay et al., 1992). HDL lipids are catabolized either separately from HDL proteins by selective uptake or via CETP transfer or as holoparticles primarily in the liver, via uptake through LDL receptors for apoE-containing HDL and through hitherto unidentified receptors for HDL holoparticles.

### C. Biological Activities

HDL particles possess multiple antiatherogenic activities (Stein and Stein, 1999; Nofer et al., 2002; Assmann and Nofer, 2003; Assmann and Gotto, 2004; Navab et al., 2004b). The central role of HDL in cellular cholesterol efflux and RCT is considered to form a basis for the capacity of HDL to attenuate atherogenesis (von Eckardstein et al., 2001; Nissen et al., 2003). However, compelling evidence has emerged that additional dimensions of the antiatherogenic action of HDL may be of major physiological and pathological relevance (Nofer et al., 2002; Assmann and Nofer, 2003; Assmann and Gotto, 2004; Navab et al., 2004b).

**1. Cholesterol Efflux Capacity.** The cholesterol efflux capacity of HDL particles is related to their ability to remove cholesterol from membranes of peripheral cells and particularly macrophages and foam cells via interaction with the ABCA1 and ABCG1 transporters and/or SR-BI receptor. Lipid-free apoA-I, apoA-II, apoE, and other HDL apolipoproteins induce fast, saturable, unidirectional and LCAT-independent efflux of cellular cholesterol and PL (von Eckardstein et al., 2001; Lewis and Rader, 2005); as a result, HDL particles efficiently acquire cholesterol in the extravascular compartment (Nanjee et al., 2001). ApoA-I is thought to play a central role in cholesterol transport from macrophages to the liver, consistent with the demonstration of accelerated RCT in mice overexpressing human apoA-I (Zhang et al., 2003); apoA-II is also able to act as a primary acceptor

and to efficiently remove cholesterol from macrophages in vivo (Rotllan et al., 2005).

Apolipoprotein-mediated lipid efflux involves specific interactions with membrane proteins, desorption of membrane lipids from caveolae, lipidation of lipid-free apolipoproteins and production of small, lipid-poor HDL (Rothblat et al., 1999; von Eckardstein et al., 2001). Lipid-free apolipoproteins remove cholesterol and PL from macrophages, aortic smooth muscle cells, and normal human skin fibroblasts but not from fibroblasts of patients with Tangier disease (Brousseau et al., 2000; Oram, 2000). Defective ABCA1 transporter function in Tangier disease has provided clear evidence that ABCA1 has a central role in lipid efflux mediated by lipid-poor apolipoproteins. In support of this mechanism, apoA-I-mediated cholesterol efflux is severely decreased by inhibition of ABCA1 with either antisense oligonucleotides or pharmacological compounds but is increased by the overexpression of ABCA1 (Oram, 2002). Thus, ABCA1 is a pivotal regulator of cellular cholesterol efflux and of the lipidation of apoA-I, a key step in formation of mature, spherical HDL particles.

ABCA1 has two highly conserved cytoplasmic ATP binding cassettes and two transmembrane domains, each of which consists of six membrane-spanning segments (Langmann et al., 1999; Santamarina-Fojo et al., 2000). It has been suggested that ABCA1 forms a channel within the plasma membrane through which cholesterol and PL are transferred ("flopped") from the inner to the outer leaflet of the plasma bilayer membrane (Hamon et al., 1997, 2000). There the lipids may be picked up by lipid-free apolipoproteins or lipid-poor particles, which bind to ABCA1 (Oram et al., 2000; Wang et al., 2000).

In addition to ABCA1, there are several other sterol-regulated ABC transporters, including ABCG1 and ABCG4, which are involved in cholesterol efflux from macrophages to mature HDL2 and HDL3 particles (Nakamura et al., 2004; Wang et al., 2004; Kennedy et al., 2005). Within the plasma membrane, ABCG1 redistributes cell cholesterol to domains that interact preferentially with mature HDL particles but not with lipid-poor apolipoproteins (Vaughan and Oram, 2005). The relative quantitative importance of cholesterol efflux mediated by ABCA1 compared with ABCG1 in macrophages remains unclear.

In contrast to lipid-free apolipoproteins, lipid-containing HDL particles induce both specific and nonspecific forms of cholesterol efflux (von Eckardstein et al., 2001). Nonspecific cholesterol efflux can be also mediated by PL vesicles, synthetic cyclodextrins, albumin or partially proteolysed HDL; it is slow, unsaturable, and bidirectional and thus appears to occur by aqueous diffusion (Rothblat et al., 1999; von Eckardstein et al., 2001). It has been suggested that SR-BI mediates the bidirectional flux between mature HDL and plasma membranes through the binding of HDL particles and subse-

quent reorganization of lipids within cholesterol- and caveolae-rich domains in the plasma membrane (de la Llera-Moya et al., 1999; Yancey et al., 2004). The PL content of HDL is an important determinant of such SR-BI-mediated cholesterol efflux (Yancey et al., 2000).

Another mechanism implicated in HDL-mediated cholesterol efflux is retroendocytosis, i.e., the uptake of HDL into clathrin-coated endosomes followed by intracellular enrichment with lipids and resecretion (Heeren et al., 1999; Takahashi and Smith, 1999). Finally, HDL-mediated cholesterol efflux from macrophages may be facilitated by apoE secretion (Mazzone, 1996). Indeed, macrophage-derived apoE can associate with HDL and improve its cholesterol acceptor properties.

Distinct cholesterol efflux properties of lipid-free and lipid-containing HDL are indicative of functional heterogeneity of HDL particles. Indeed, a decrease in the lipid content of HDL is generally thought to increase its capacity to remove cellular cholesterol (Ohta et al., 1995; Sasahara et al., 1998); small, dense, lipid-poor, protein-rich HDL particles are therefore considered to represent more efficient cholesterol acceptors compared with their large, light, lipid-rich, protein-poor counterparts (Asztalos et al., 1997). For example, small, lipid-poor HDL predominate in rabbits expressing human apoA-I; in parallel, the cholesterol efflux capacity of rabbit serum increases (Duverger et al., 1996a,b).

Interestingly, pre- $\beta$ 1-HDL, the initial product of apoA-I lipidation, is not essential for cellular cholesterol efflux (Sviridov et al., 2002), thereby suggesting that lipid-free, rather than lipid-poor, apolipoproteins function as primary cholesterol acceptors (Asztalos et al., 1997). Lipid-free and/or lipid-poor HDL apolipoproteins induce cholesterol uptake via interaction with ABCA1; consistent with this observation, plasma levels of small pre- $\beta$ 1-HDL particles correlate with serum capacity to induce ABCA1-mediated cholesterol efflux from J774 macrophages (Asztalos et al., 2005). Conversely, large, lipid-rich HDL particles appear to represent a better ligand for cellular uptake of CE mediated by SR-BI compared with small, lipid-poor HDL (de Beer et al., 2001; Thuahnai et al., 2004), consistent with the role of these particles in RCT from peripheral cells to the liver (von Eckardstein et al., 2001; Asztalos et al., 2005).

**2. Antioxidative Activity.** HDL antioxidative activity is typically observed as inhibition of LDL oxidation by HDL; LDL is thought to represent the major physiological target of HDL antioxidative action in vivo (Van Lenten et al., 2001a; Navab et al., 2004b). HDL is also able to inhibit generation of reactive oxygen species (ROS) in vitro under conditions of cell culture (Robbesyn et al., 2003; Lee et al., 2005) and in vivo in a rabbit model of acute arterial inflammation (Nicholls et al., 2005b). In addition, inhibitory actions of HDL on LDL oxidation have been reported in vitro upon their coincubation (Parthasarathy et al., 1990) and in vivo upon HDL injection (Klimov et al., 1993). HDL potently protects both

lipid and protein moieties of LDL and inhibits accumulation of various oxidation products in LDL, including oxidized PL and short-chain aldehydes (Van Lenten et al., 2001a; Navab et al., 2004b).

The antioxidative activity of HDL is related to the presence of several apolipoproteins and enzymes with antioxidative properties in HDL particles. Apolipoproteins that possess antioxidative activity include apoA-I, apoE, apoJ, apoA-II, and apoA-IV. It appears that a major component of the antioxidative activity of HDL can be ascribed to apoA-I which can prevent and/or delay LDL oxidation by removing oxidized PL, including 1-palmitoyl-2(5-oxovaleroyl)-*sn*-glycero-3-phosphorylcholine and 1-palmitoyl-2-glutaroyl-*sn*-glycero-3-phosphorylcholine, from LDL and from arterial wall cells (Navab et al., 2000a,b). The capacity of apoA-I to remove oxidized lipids is not specific for arterial wall cells, because similar effects have been reported for erythrocytes and astrocytes (Klimov et al., 2001; Ferretti et al., 2003, 2004). Circulating HDL accumulate LOOH and have been proposed to function as a "sink" for oxidized lipids (Bowry et al., 1992), ensuring their efficient elimination from the circulation through the liver.

ApoE possesses established antiatherosclerotic activity, which is normally ascribed to its lipid transport properties (Davignon, 2005). However, the action of apoE goes beyond such activity. Indeed, apoE possesses distinct antioxidative properties (Miyata and Smith, 1996) and can promote regression of atherosclerosis independently of lowering plasma cholesterol levels (Thorngate et al., 2000; Tangirala et al., 2001; Raffai et al., 2005). HDL-associated apoJ can inhibit oxidation of LDL by artery wall cells (Navab et al., 1997); in addition, apoJ is cytoprotective at low physiological levels (Trogakos et al., 2005). The beneficial actions of apoJ may be related to its ability to maintain integrity of membrane and lipoprotein lipids via its hydrophobic-binding domains (Jordan-Starck et al., 1992). Antioxidative properties have also been reported for apoA-II (Boisfer et al., 2002) and apoA-IV (Ostos et al., 2001). The capacity of apoA-II to protect LDL from oxidation is, however, questionable, given the fact that overexpression of human apoA-II in dyslipidemic mice accelerates atherosclerosis, increases aortic accumulation of oxLDL, and reduces antioxidative activity of HDL (Ribas et al., 2004; Rotllan et al., 2005). Such proatherogenic actions of apoA-II may be related to the displacement of antiatherogenic apoA-I and PON1 by apoA-II from HDL particles (Ribas et al., 2004). Finally, HDL is able to function as a preventive antioxidant through its capacity to bind transition metal ions (Kunitake et al., 1992), which in free form are potent catalyzers of LDL oxidation. Intriguingly, plasma HDL carry amyloid- $\beta$  peptide, a major component of senile neuritic plaques and a strong chelator of transition metals (Kontush, 2004).

Major HDL enzymes possessing antioxidative activity are PON1, PAF-AH, LCAT, and GSPx (Van Lenten et



al., 2001a; Navab et al., 2004b). PON1 is a component of HDL that is thought to hydrolyze LDL-derived short-chain oxidized PL once they are formed (Aviram et al., 1998). PON1 is anchored to lipids via its hydrophobic N terminus (Josse et al., 2002; Harel et al., 2004); the association of PON1 with HDL is a prerequisite for maintaining normal serum activity of the enzyme. HDL provides the optimal physiological acceptor complex for PON1, in terms of both stimulating enzyme secretion and stabilizing the secreted peptide (James and Deakin, 2004); PON1 interaction with apoA-I is critical for enzyme stability (Gaidukov and Tawfik, 2005). HDL and, less efficiently, VLDL but not LDL promote PON1 secretion from cells; the differences between these lipoproteins are related to differences in their lipid composition (Deakin et al., 2005).

PAF-AH and LCAT can also hydrolyze LDL-derived short-chain oxidized PL; the relationship between the hydrolyzing activities of PON1, PAF-AH, and LCAT toward oxidized PL remains unclear. Recent data question the ability of PON1 to hydrolyze oxidized PL and suggest that PAF-AH, rather than PON-1, is the oxidized PL hydrolase in HDL (Marathe et al., 2003; Connelly et al., 2005). Consistent with this conclusion, HDL-associated PAF-AH is thought to play an antiatherogenic role, in contrast to the LDL-associated enzyme (Quarek et al., 2001; Tsimihodimos et al., 2003; Zaleski and Macphee, 2005). Indeed, local arterial expression of PAF-AH reduces accumulation of oxLDL and inhibits inflammation, shear stress-induced thrombosis, and neointima formation in balloon-injured carotid arteries of nonhyperlipidemic rabbits (Arakawa et al., 2005).

The antioxidative activity of PON1 purified from human serum has recently been ascribed to the presence of detergents or some other unidentified proteins (Teiber et al., 2004). Interestingly, PON1 has been reported to catalyze the hydrolysis of a variety of lactones, including homocysteine thiolactone, suggesting that its native activity is as a lactonase (Jakubowski, 2000; Draganov et al., 2005; Khersonsky and Tawfik, 2005). Plasma levels of homocysteine are a strong CV risk factor (Duell and Malinow, 1997); by detoxifying homocysteine thiolactone, PON1 could protect against homocysteinylation, a post-translational modification of proteins associated with attenuated biological activity and a potential contributing factor to atherosclerosis.

In addition, HDL-associated PON1 enhances cholesterol efflux from macrophages via increased HDL binding mediated by ABCA1 (Rosenblat et al., 2005). PON1-induced cellular accumulation of lysophosphatidylcholine, which stimulates cholesterol efflux via the ABCA1 pathway, may account for this effect (Hara et al., 1997). One can hypothesize that both lactonase activity and an RCT-related mechanism may contribute to the antiatherosclerotic effects of PON1 observed in vivo (Shih et al., 1998; Tward et al., 2002).

Another HDL enzyme, GSPx, can reduce LOOH to corresponding hydroxides and thereby detoxify them (Maddipati and Marnett, 1987; Arthur, 2000; Chen et al., 2000). LOOH-reducing activity mediated by Met residues of apoA-I and apoA-II has also been reported (Sattler et al., 1994; Garner et al., 1998). Finally, upon HDL oxidation with peroxyxynitrite, apoA-I increases generation of PL core aldehydes that are subsequently hydrolyzed by HDL-associated enzymes, such as PAF-AH and/or PON1, with formation of lysophospholipids (Ahmed et al., 2001). Such a PAF-AH/PON1-coupled protective function of apoA-I can effectively divert proatherogenic LOOH to less harmful products (Van Lenten et al., 2001a; Tselepis and Chapman, 2002; Navab et al., 2004b).

Apolipoproteins and enzymes with antioxidative activities are nonuniformly distributed across HDL subfractions. In vivo PON1 is preferentially associated with large HDL but can be displaced to small, dense particles upon ultracentrifugation (Cabana et al., 2003; Kontush et al., 2003; Bergmeier et al., 2004). The size and shape of HDL seem to be critical for PON1 binding (Josse et al., 2002). By contrast, apoJ is associated with a subset of small HDL, which also contains PON1 (Kelso et al., 1994). Similarly, LCAT activity (Kontush et al., 2003), PAF-AH activity (Kontush et al., 2003), and apoA-IV (Bisgaier et al., 1985) are enriched in small, dense HDL isolated by ultracentrifugation. As a consequence, HDL particles are heterogeneous in their antioxidative activity. Under mild oxidative stress induced by an azo initiator 2,2'-azobis-(2-amidinopropane) hydrochloride or  $\text{Cu}^{2+}$ , the antioxidative activity of HDL subfractions isolated by density gradient ultracentrifugation against LDL oxidation increases with increment in density in the order: HDL2b < HDL2a < HDL3a < HDL3b < HDL3c, thereby establishing that small, dense HDL act as potent protectors of LDL from oxidative stress (Kontush et al., 2003). Similarly, HDL3 is a more potent protector of LDL from in vitro oxidation compared with HDL2 (Yoshikawa et al., 1997; Huang et al., 1998). The antioxidative activity of small, dense HDL is related to the inactivation of proatherogenic products of LDL lipid peroxidation, primarily LOOH (Kontush et al., 2003). Mechanistically, this activity may arise from synergy in inactivation of oxidized lipids by enzymatic (hydrolysis) and nonenzymatic (physical removal) mechanisms, in part reflecting distinct intrinsic physicochemical properties of the small, dense HDL3c subfraction (Kontush et al., 2003).

The relative importance of HDL antioxidative activity in the overall cardioprotective effect of HDL compared with other biological actions remains indeterminate. A recent study proposed that the antioxidative activity of HDL is less important than cholesterol efflux capacity, as suggested by the absence of antioxidative effects of human apoA-I expression in apoE<sup>-/-</sup> mice accompanied by delayed atherosclerosis (Choudhury et al., 2004). The

increase in non-HDL-C levels observed in this animal model, however, renders interpretation of these results complex. By contrast, both cholesterol efflux capacity and antioxidative activity of HDL were impaired in parallel to a similar extent in apoA-I<sup>-/-</sup> mice in another recent study (Moore et al., 2005).

**3. Anti-Inflammatory Activity.** The anti-inflammatory activity of HDL is illustrated by the ability of HDL to decrease cytokine-induced expression of adhesion molecules on endothelial cells and to inhibit monocyte adhesion to these cells. HDL efficiently inhibit expression of the vascular cell adhesion molecule-1, intercellular adhesion molecule-1, and E-selectin in vitro induced by tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin (IL)-1, or endotoxin (Cockerill et al., 1995; Calabresi et al., 1997; Baker et al., 1999). Moreover, this potent anti-inflammatory activity observed in vitro can be translated into inhibition of adhesion molecule expression and a decrease in neutrophil infiltration in the arterial wall by reconstituted HDL (rHDL) in a rabbit model of acute arterial inflammation (Nicholls et al., 2005b). The ability of HDL to inhibit adhesion molecule expression may be related to the presence of apoA-I, apoA-II, apoA-IV, and/or distinct molecular species of PL, including sphingosine-1-phosphate (S1P) and sphingosylphosphorylcholine (Baker et al., 1999; Recalde et al., 2004; Nofer and Assmann, 2005). The anti-inflammatory action of HDL involves inhibition of TNF- $\alpha$ -stimulated activation of sphingosine kinase and production of S1P, which induces adhesion molecule expression in endothelial cells (Xia et al., 1999); transforming growth factor  $\beta$  may function as an important mediator of the anti-inflammatory activity (Norata et al., 2005). In addition, HDL attenuate IL-6 production in endothelial cells exposed to proinflammatory stimuli, such as TNF- $\alpha$  or endotoxin (Gomasaschi et al., 2005).

The anti-inflammatory action of HDL also involves hydrolysis of oxidized lipids by HDL-associated enzymes (PAF-AH and PON1) and is mechanistically similar to the antioxidative activity of HDL (Van Lenten et al., 2001a; Navab et al., 2004b; Recalde et al., 2004). Oxidized PL possess potent proinflammatory activities and can trigger arterial inflammation (Furnkranz et al., 2005). Inactivation of oxidized lipids by HDL may be associated with decreased expression of adhesion molecules in and decreased macrophage adhesion to endothelial cells (Theilmeyer et al., 2000; Navab et al., 2004b).

Direct interaction of apoA-I with T lymphocytes, which can block subsequent activation of monocytes by lymphocytes, represents another plausible mechanism of HDL anti-inflammatory action (Burger and Dayer, 2002). In addition, apoA-I has been reported to diminish neutrophil activation in vitro (Liao et al., 2005). The anti-inflammatory activity of HDL in vivo is consistent with elevated levels of CRP in subjects with hypoalphalipoproteinemia (Sampietro et al., 2002), with negative correlation between plasma levels of CRP and HDL-C

(Pirro et al., 2003) but also between plasma levels of intercellular adhesion molecule-1 and HDL-C and particularly small, dense HDL3-C (Kent et al., 2004).

The potential heterogeneity of HDL anti-inflammatory activity remains poorly characterized. HDL3 has been reported to be superior to HDL2 in terms of its capacity to inhibit vascular cell adhesion molecule-1 expression in endothelial cells (Ashby et al., 1998), a finding that is consistent with the potent antioxidative activity of small, dense HDL3 particles (Yoshikawa et al., 1997; Huang et al., 1998; Kontush et al., 2003).

**4. Antiapoptotic, Vasodilatory, Antithrombotic, and Anti-Infectious Activities.** Other antiatherogenic activities of HDL include antiapoptotic and vasodilatory actions, mitogenic activity in endothelial cells, attenuated platelet activation, and anticoagulant and anti-infectious activities (Calabresi et al., 2003).

HDL potently inhibit apoptosis in endothelial cells induced by oxLDL (Suc et al., 1997; Robbesyn et al., 2003) or TNF- $\alpha$  (Sugano et al., 2000); this effect is paralleled by decreased intracellular generation of ROS and diminished levels of apoptotic markers, suggesting that it can be related to the intracellular antioxidative actions of HDL or HDL components (Suc et al., 1997; Sugano et al., 2000; Robbesyn et al., 2003). Indeed, HDL contain bioactive lysophospholipids, including S1P (Nofer and Assmann, 2005; Zhang et al., 2005), a potent antiapoptotic agent, which may mediate the antiapoptotic effect of HDL via increased NO production (Kwon et al., 2001).

Similarly, HDL vasodilatory activity may be related to the stimulation of NO release by endothelial cells mediated by intracellular Ca<sup>2+</sup> mobilization and phosphorylation of NOS upon association with apoA-I (Drew et al., 2004; Nofer et al., 2004). Such activation of NO production involves HDL binding to SR-BI with a subsequent increase in intracellular ceramide levels (Yuhanna et al., 2001; Li et al., 2002). Furthermore, HDL can stimulate production of prostacyclin, which possesses potent vasorelaxing activity (Beitz and Forster, 1980; Norata et al., 2004). Again, the vasoactive effects of HDL can be mediated by S1P acting via the lysophospholipid receptor S1P3 (Nofer et al., 2004). S1P may be equally important for mitogenic effects of HDL in endothelial cells and for the inhibitory action of HDL on the migration of vascular smooth muscle cells (Kimura et al., 2003; Nofer and Assmann, 2005; Tamama et al., 2005).

Similarly, increased production of NO may form a basis for the inhibitory action of HDL on platelet aggregation (Chen and Mehta, 1994). The antithrombotic activity of HDL is observed as inhibitory actions on factors that promote blood coagulation, including tissue factor, factors X, Va, and VIIIa (Nofer et al., 2002; Calabresi et al., 2003). Mechanistically, this effect may be related to the presence of cardiolipin and phosphatidylethanolamine, two minor anionic PL with potent anticoagulant properties that are enriched in the HDL fraction (Degu-

chi et al., 2000). In addition, HDL acts via its protein moiety to enhance the anticoagulant activity of protein S and activated protein C (Griffin et al., 1999).

Finally, HDL play a major role in the binding and clearance of circulating endotoxin to the bile and thereby inhibit endotoxin-induced cellular activation, resulting in potent anti-infectious activity (Pajkrt et al., 1996; Levels et al., 2001; Stoll et al., 2004). The inactivation of endotoxin by HDL is mediated by direct interaction with apoA-I (Ma et al., 2004) and involves reduced CD14 expression on monocytes as a key step (Pajkrt et al., 1996). In addition, human HDL possess specific trypanosome-lytic activity, which selectively protects humans from *Trypanosoma brucei brucei* (Hajduk et al., 1989).

The potential heterogeneity of these antiatherogenic activities among HDL particles is indeterminate. The anticoagulant activity of tissue factor pathway inhibitor in human plasma has been reported to be preferentially associated with dense subspecies of HDL and LDL (Lesnik et al., 1993). Similarly, the trypanosome-lytic activity is associated with a minor large and dense HDL subfraction with a molecular mass of 490 kDa (Hajduk et al., 1989). Finally, our recent data suggest that small, dense HDL potently inhibit apoptosis induced in endothelial cells by oxLDL (Suc et al., 1997; Robbesyn et al., 2003; J. de Souza, M. J. Chapman, and A. Kontush, unpublished data).

### III. Functionally Defective High-Density Lipoprotein in Dyslipidemic and Inflammatory States

HDL is known to undergo dramatic modification in structure and composition as a result of the concerted actions of the acute-phase response and inflammation (Khovidhunkit et al., 2004b; Esteve et al., 2005). The close association between inflammation, oxidative stress, dyslipidemia, and atherosclerosis suggests that such HDL alterations play a significant role in disease progression. As a result, HDL particles progressively lose normal biological activities and acquire altered properties. Such altered HDL particles have been termed "dysfunctional HDL" (Navab et al., 2001b), and HDL has been proposed to possess "chameleon-like properties" (Navab et al., 1996; Van Lenten et al., 2001a). It is essential to emphasize that the degree of loss of normal HDL function compared with the absence of this function depends on the assay used to characterize HDL functionality. Indeed, HDL can be dysfunctional (with total loss of function) in cell-based or cell-free assays aimed at measuring anti-inflammatory activity (Navab et al., 2001b; Ansell et al., 2003), whereas measurements of antioxidative activity (Kontush et al., 2003, 2004, 2005; Hansel et al., 2004; Nobecourt et al., 2005) or cholesterol efflux capacity (Banka et al., 1995; Cavalero et al., 1995; Brites et al., 2000; Khovidhunkit et al.,

2001) reveal a deficiency in normal HDL function rather than a complete dysfunction.

#### A. Altered High-Density Lipoprotein Composition and Enzymatic Activities in Dyslipidemic and Inflammatory States

**1. Apolipoproteins.** Both the plasma levels and apolipoprotein content of HDL can be significantly altered during the acute phase as well as during acute and chronic inflammation. Levels of apoA-I and apoA-II decrease, whereas those of apoA-IV, apoA-V, apoJ, and apoE increase (Khovidhunkit et al., 2004a,b). The decrease in HDL apoA-I levels in inflammatory states is related to both decreased apoA-I synthesis in the liver and apoA-I replacement in HDL particles by SAA (Fig. 5) (Khovidhunkit et al., 2004b; Esteve et al., 2005). SAA is a 12-kDa acute-phase protein whose circulating levels can be induced up to 1000-fold (Malle et al., 1993). HDL is a major carrier of SAA in human, rabbit, and murine plasma (Hoffman and Benditt, 1982a; Marhaug et al., 1982; Cabana et al., 1996). In the circulation, SAA does not exist in a free form and associates with non-HDL lipoproteins in the absence of HDL (Cabana et al., 2004). In the presence of HDL, SAA is specifically associated with small, dense HDL3 subspecies (Benditt and Erikssen, 1977; Hoffman and Benditt, 1982a; Coetzee et al., 1986; Cabana et al., 1996) via its N-terminal domain (Liang et al., 1996), but it is also present in large and intermediate HDL (Coetzee et al., 1986; Cabana et al., 1996).

SAA is able to replace apoA-I in small, dense HDL upon induction of the acute phase (Parks and Rudel, 1985; Coetzee et al., 1986); as a result, plasma levels of apoA-I decrease (Cabana et al., 1996). In dense HDL, SAA can account for up to 80% of total protein; such enrichment can further increase HDL protein content and density (Cabana et al., 1989). Elevated plasma levels of SAA are accompanied by elevated levels of lipid-free apoA-I, probably due to the dissociation of apoA-I from HDL (Cabana et al., 1996). In rabbits and mice, SAA can completely replace apoA-I in a subset of small, dense HDL particles, thereby functioning as a structural apolipoprotein (Cabana et al., 1996, 1999). In such SAA-only HDL, 20 molecules of SAA have been estimated to replace all 3 molecules of apoA-I in each HDL particle. SAA is mainly produced by the liver but also by arterial wall cells and adipocytes (Hoffman and Benditt, 1982b; Malle et al., 1993). Primary murine hepatocytes secrete SAA as a monomer, which subsequently associates with small, lipid-poor, apoA-I-containing HDL secreted via a separate pathway (Hoffman and Benditt, 1982b).

Similar to CRP, elevated plasma levels of SAA have been reported to represent a CV risk factor. In a prospective case-control study, plasma level of hs-CRP was the strongest univariate predictor of CV risk in apparently healthy postmenopausal women; the relative risk of events for women in the highest compared with the

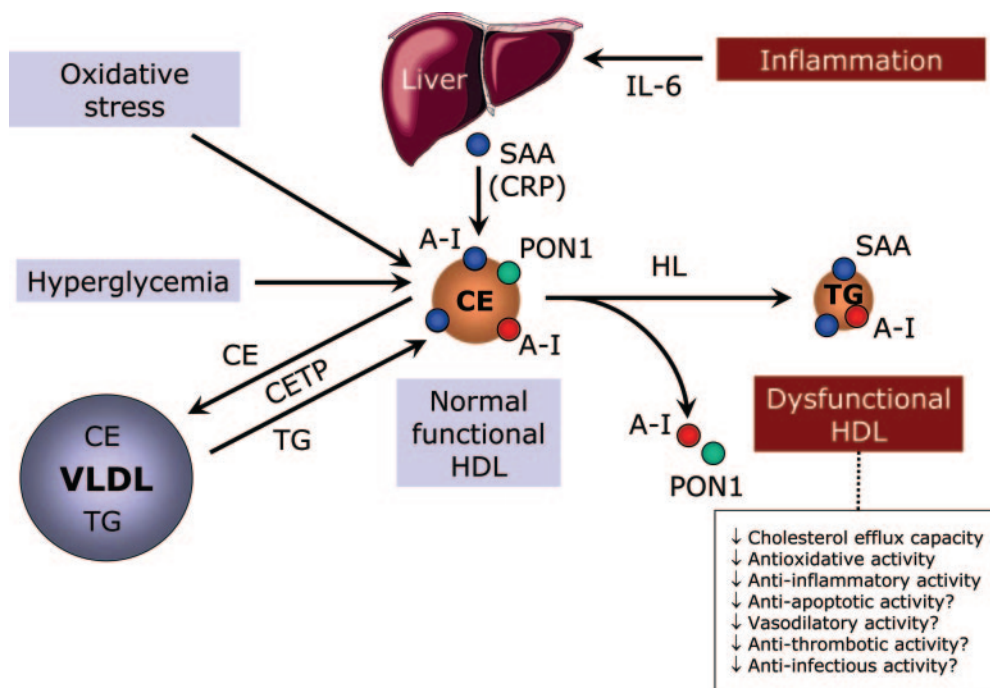


FIG. 5. Abnormal metabolism and deficient biological activities of HDL in atherogenic dyslipidemias of metabolic disease. Chronic inflammation characteristic of metabolic disease, such as MetS and type 2 diabetes, is associated with elevated plasma levels of IL-6. As a result, the liver produces SAA, which replaces apoA-I and PON1 in HDL. Oxidative stress, hyperglycemia, and elevated activity of CETP are other important modulators of HDL function. Oxidative stress modifies specific amino acids in apoA-I, whereas hyperglycemia results in apoA-I glycation. CETP exchanges CE and TG between HDL and TG-rich lipoproteins, such as VLDL; as a result, HDL become enriched in TG. Such enrichment in TG induces conformational changes in apoA-I, which becomes less accessible for the interaction with other lipoproteins, including LDL, and cannot eliminate oxidized lipids from LDL. Subsequent HDL hydrolysis by HL produces small, dense HDL that are enriched in TG and in SAA and contain apoA-I in an incorrect conformation; such HDL possess deficient functionality compared with normal HDL particles.

lowest quartile was 4.4 (95% confidence interval, 2.2–8.9) (Ridker et al., 2000). However, SAA levels also revealed a significant, albeit weaker, association with CV events (relative risk for the highest versus lowest quartile 3.0). Remarkably, the levels of hs-CRP and SAA were significant predictors of CV risk even in the subgroup of women with low LDL-C levels (Ridker et al., 2000).

In addition, baseline SAA levels were independently associated with angiographic CAD (Liuzzo et al., 1994) and were highly predictive of 3-year CV events in women referred for coronary angiography for suspected myocardial ischemia (Liuzzo et al., 1994) as well as of the progression of carotid atherosclerosis in patients undergoing ultrasound investigations (Schillinger et al., 2005). By comparison, hs-CRP was not associated with angiographic CAD but, like SAA, was strongly and independently predictive of adverse CV outcome. Elevation of hs-CRP and SAA levels at the time of hospital admission predicted poor outcome in patients with unstable angina (Liuzzo et al., 1994). Elevated SAA levels were associated with increased mortality in transplant patients with CAD (Fyfe et al., 1997). Furthermore, circulating SAA is elevated in CHD patients (Fyfe et al., 1997; Delanghe et al., 2002), subjects with CAD (Fyfe et al., 1997; Winkler et al., 2005), and patients with type 2 diabetes (Choudhury and Leyva, 1999) compared with healthy control subjects. Local concentrations of SAA

are elevated in the coronary artery at sites of plaque rupture compared with concentrations in the aorta in patients with acute myocardial infarction (Maier et al., 2005). Finally, plasma levels of SAA correlate with the development of atherosclerosis in mouse models of this disease (Herrington and Parks, 2004; Lewis et al., 2004).

The proatherogenic properties of SAA are intimately related to its biological activities. SAA is present in human and murine atherosclerotic lesions and colocalises with apoA-I, apoB, and the proteoglycan perlecan (Yamada et al., 1996; O'Brien et al., 2005). HDL enrichment in SAA enhances *in vitro* HDL binding to proteoglycans due to the presence of a proteoglycan-binding domain in the SAA molecule (Lewis et al., 2004; O'Brien et al., 2005); thus, SAA might immobilize HDL particles in the arterial wall, which would otherwise transport cholesterol from the plaque to the liver. In addition, SAA-enriched HDL are rapidly cleared from the circulation (Hoffman and Benditt, 1983); SAA may then play a role in the lipoprotein redistribution to the arterial wall. Finally, enrichment in SAA may impair the normal atheroprotective activities of HDL (see below).

Human LDL contain low levels of SAA; LDL-associated SAA has been recently proposed to represent a risk factor for cardiac events in stable CAD (Ogasawara et al., 2004). SAA-carrying LDL represent 5 to 6% of total LDL, contain increased levels of products of lipid and protein oxidation and may represent circulating oxLDL

(Ogasawara et al., 2004). The increased SAA content of such LDL may enhance its retention in atherosclerotic lesions. Such SAA-mediated accumulation of LDL lipids in the lesions may lead to plaque instability and result in plaque rupture (Johnson et al., 2004).

Apart from its replacement by SAA, apoA-I can undergo other modifications in the circulation. Amino acid residues in apoA-I, such as methionine, cysteine, tyrosine, and lysine residues, can be selectively modified under the action of prooxidants secreted by arterial wall cells (Bergt et al., 2004; Zheng et al., 2004; Nicholls et al., 2005d) and nonenzymatically glycosylated in the presence of high levels of glucose (Fievet et al., 1995). In human atherosclerotic lesions, proteins oxidized by HOCl or acrolein colocalise with extracellular apoA-I (Bergt et al., 2004; Shao et al., 2005c). Oxidized amino acid residues, including chlorotyrosines, nitrotyrosines and oxidized lysine and methionine residues, are present in apoA-I isolated from plasma and from human atherosclerotic lesions (Bergt et al., 2004; Zheng et al., 2004; Panzenbock and Stocker, 2005; Shao et al., 2005c); furthermore, the apoA-I content of chloro- and nitrotyrosines is increased in plasma of patients with CV disease. Myeloperoxidase, a major source of chlorinated ROS in the arterial wall, binds to apoA-I in vitro and in vivo and produces similar patterns of oxidized amino acids. Residue Tyr-192 represents the major target for myeloperoxidase-catalyzed oxidation in the apoA-I molecule both in vitro and in vivo, whereas three other tyrosine residues at positions 29, 166, and 236 are modified to a lesser extent (Zheng et al., 2005). Thus, apoA-I represents a selective target for chlorination and nitration in human atheromatous tissue catalyzed by myeloperoxidase. In vivo oxidation of apoA-I is equally consistent with the observation that HDL from hypercholesterolemic chickens contain higher amounts of oligomeric apoA-I and are more susceptible to in vitro oxidation than HDL from control animals (Artola et al., 1997).

**2. Enzymes with Antioxidative and Anti-Inflammatory Properties.** HDL-associated enzymes, including PAF-AH, PON1, and LCAT, can become dysfunctional and/or depleted under inflammatory conditions (Navab et al., 1997; Van Lenten et al., 2001b), in metabolic diseases involving low HDL levels (type 2 diabetes, MetS) (Hansel et al., 2004; Nobecourt et al., 2005), and in premature CHD (Ansell et al., 2003). Induction of the acute-phase response is associated with decreased PON1 activity, probably due to the replacement of PON1 by SAA (Fig. 5) (Navab et al., 1997; Van Lenten et al., 2001b). Furthermore, decreased PON1 activity may be caused by enzyme inactivation as a result of oxidation (Jaouad et al., 2003) and/or glycation (Hedrick et al., 2000; Ferretti et al., 2001). Consistent with these observations, serum concentrations of PON1 are decreased in subjects with MetS (Blatter Garin et al., 2005) and in patients with type 1 and type 2 diabetes (Boemi et al., 2001; Costa et al., 2005), who feature elevated levels of inflammation

and oxidative stress (Ridker et al., 2004a). Serum PON1 activity decreases with age (Costa et al., 2005) and is lower in subjects with MetS (Blatter Garin et al., 2005) and low HDL-C (Brites et al., 2004) and patients with type 2 diabetes (Boemi et al., 2001; Costa et al., 2005) and familial hypercholesterolemia (FH) (Mackness et al., 1991) compared with age-matched healthy control subjects. Moreover, low PON1 activity toward paraoxon has been reported to represent an independent risk factor for coronary events in men at high CV risk (Mackness et al., 2003).

HDL-associated PAF-AH activity, expressed as a percentage of total serum PAF-AH activity, is lower in FH patients than in control subjects (Karabina et al., 1997; Tsimihodimos et al., 2002). By contrast, LDL-associated PAF-AH activity is elevated in homozygous FH subjects (Tsimihodimos et al., 2002), indicating a major redistribution of PAF-AH activity in plasma of FH individuals from apoA-I- to apoB-containing lipoproteins (Tsimihodimos et al., 2002). Finally, LCAT activity is diminished under inflammatory conditions (Jonas, 2000).

**3. Lipid Components.** Although apolipoproteins and enzymes are major determinants of altered HDL function, it is considerably influenced by changes in lipid content. HDL core enrichment in TG with CE depletion is the most frequent abnormality of HDL lipid composition (Fig. 5) and occurs in hypertriglyceridemic states associated with decreased activity of LPL, decreased activity of HL, and/or decreased activity of LCAT; all these metabolic alterations are frequently observed in the acute phase and during inflammation (Cabana et al., 1996). In addition, HDL-TG content can be raised as a consequence of elevated CETP-mediated TG transfer from VLDL to HDL (de Grooth et al., 2004a; Le Goff et al., 2004).

Under such conditions, TG typically replace CE in the HDL core, resulting in a low CE/TG ratio and in a decrease in plasma HDL-C levels, another feature of the acute phase response (Khovidhunkit et al., 2004b). Interestingly, a similar elevation in HDL-TG, decrease in HDL-C, and increase in inflammatory markers are observed in the postprandial phase (Schaefer et al., 2005). Human acute-phase HDL obtained from patients undergoing bypass surgery are enriched in TG and depleted of CE (Pruzanski et al., 2000). Acute-phase HDL also contain elevated levels of nonesterified fatty acids (NEFA), lysophosphatidylcholines and isoprostanes compared with normal HDL; in addition, CE levels are decreased (Pruzanski et al., 2000). Similarly, HDL3 from subjects with myocardial infarction are enriched in TG and depleted of PL (Clifton et al., 1985). Induction of the acute-phase response in monkeys increases plasma TG levels and TG content in total HDL and in all HDL subfractions and also decreases HDL-C and HDL-CE; these effects are paralleled by cytokine-induced decreases in the activities of HL, LPL, and LCAT and increases in the activity of CETP (Auerbach and Parks, 1989; Ettinger et

al., 1990; Cabana et al., 1996). In addition, acute-phase HDL obtained from hamsters display reduced CE content and elevated PL and FC contents compared with normal HDL (Khovidhunkit et al., 2001). As a consequence of decreased LCAT activity, increased HDL concentrations of free cholesterol are frequently observed in inflammatory states; in addition, HDL free cholesterol is elevated in genetic LCAT deficiency (Jonas, 2000). By contrast, reduced PL content of HDL is a less consistent finding (Khovidhunkit et al., 2004b). The reduction of HDL-PL reported in some studies (Clifton et al., 1985; Cabana et al., 1989) may reflect elevated activity of secretory phospholipase A2 frequently observed in the acute phase (Crowl et al., 1991; Pruzanski et al., 1993; Tietge et al., 2002).

Equally, HDL composition can be abnormal in other forms of dyslipidemia. In FH, HDL levels of TG and the TG/CE ratio are increased (Bagdade et al., 1991; Frenais et al., 1999). HDL enrichment in TG is associated with accelerated CE transfer from LDL and HDL to TG-rich lipoproteins in plasma of FH patients, an observation that has been linked to abnormal properties of plasma VLDL1 (Bagdade et al., 1991; Guerin et al., 1995a, 2000a). By contrast, decreased CE transfer from LDL and HDL to TG-rich lipoproteins, such as those observed in plasma of subjects with heterozygous CETP deficiency, results in reduced HDL content of TG, elevated content of CE, and increased HDL size (Koizumi et al., 1991).

Finally, HDL lipids can be oxidized *in vivo* with formation of biologically active compounds. For instance, HDL oxidation by HOCl produces 2-chlorohexadecanal, a chlorinated fatty aldehyde formed upon oxidative cleavage of plasmalogen, which exerts inhibitory actions on endothelial NOS (Marsche et al., 2004).

### *B. Abnormal High-Density Lipoprotein Metabolism in Dyslipidemic and Inflammatory States*

HDL metabolism is substantially altered in dyslipidemic states, including hypertriglyceridemia, hypercholesterolemia, mixed dyslipidemia and hypo- and hyperalphalipoproteinemia and also during infection and inflammation. As discussed above, hypertriglyceridemia is characterized by decreased levels of HDL-C and increased HDL-TG content due to the action of CETP. Such low HDL-C dyslipidemias associated with hypertriglyceridemia are characteristic of metabolic diseases associated with elevated CV risk, such as type 2 diabetes and MetS. Mechanisms leading to reduced plasma HDL-C levels and HDL particle numbers in hypertriglyceridemic states are as follows: 1) small HDL particles, which result from the intravascular lipolysis of TG-enriched HDL, are cleared more rapidly from the circulation; 2) TG-enriched HDL are intrinsically more unstable in the circulation, with apoA-I loosely bound; 3) lipolysis of TG-enriched HDL lower HDL particle numbers by causing apoA-I to be shed from HDL particles

and cleared from the circulation; 4) dysfunctional LPL or reduced LPL activity contributes to the lowering of HDL levels by reducing the availability of surface constituents of TG-rich lipoproteins that sequester to the plasma pool of nascent HDL particles (Lamarche et al., 1999).

The CE/TG ratio therefore represents a critical factor in determining HDL particle stability and plasma residence time; HDL possessing decreased CE/TG ratios are less stable than normal particles (Sparks et al., 1995; Rashid et al., 2002; Borggreve et al., 2003; Rashid et al., 2003). Importantly, a decrease in circulating HDL-C levels and an increase in TG levels are typical components of the acute-phase reaction (Khovidhunkit et al., 2004b; Esteve et al., 2005). In humans, influenza infection is associated with decreased levels of HDL-C (Marchesi et al., 2005); in rodents, endotoxin injection increases plasma TG levels through increased hepatic secretion and/or delayed clearance (Feingold et al., 1992).

HDL metabolism critically depends on the activity of CETP. In metabolic diseases such as type 2 diabetes and MetS, elevated CETP activity results in increased CE transfer from HDL to TG-rich lipoproteins and in reciprocal TG transfer, producing TG-enriched HDL and decreasing HDL-C levels (Fig. 5) (Le Goff et al., 2004). Conversely, CETP deficiency reduces the exchange of TG and CE between HDL and TG-rich lipoproteins and elevates HDL-C due to CE retention. As a consequence, increased CETP activity is thought to be proatherogenic in humans (Barter et al., 2003b). Consistent with this hypothesis, the low-active CETP *TaqIB* variant B2B2 is associated with higher HDL-C plasma levels and a lower risk of CAD than the high-active variant B1B1 (Boekholdt et al., 2005). In addition, baseline CETP levels positively correlated with carotid intima-media thickness (IMT) in 2 years in FH patients treated with statins (de Grooth et al., 2004b). Finally, baseline CETP levels were associated with future CAD in a subset of hypertriglyceridemic subjects from an apparently healthy population (Boekholdt et al., 2004).

Elevated activity of CETP may therefore form a basis for low HDL-C phenotypes; alternatively, they may result from a deficiency of apoA-I (Ng et al., 1995), elevated activities of HL (Tato et al., 1995), reduced activities of LCAT (Kuivenhoven et al., 1997; Hovingh et al., 2005) or LPL (Blades et al., 1993), or a combination of these. Furthermore, HDL metabolism is altered in hyperalphalipoproteinemia, which can arise from a genetic deficiency of CETP and/or HL, as well as from increased production of apoA-I (Yamashita et al., 2000). Familial CETP deficiency is associated with accumulation of large CE-rich HDL2 particles (Yamashita et al., 2000); in addition, large HDL predominate in familial HL deficiency (Cohen et al., 1999).

In hypercholesterolemia, abnormalities of HDL metabolism include moderate decreases in plasma apoA-I and HDL-C levels (Schaefer et al., 1992; Frenais et al.,

1999). Subnormal plasma levels of apoA-I in FH are probably related to an increased fractional catabolic rate of apoA-I observed both in homozygous (Schaefer et al., 1992) and heterozygous (Frenais et al., 1999) forms of FH. In homozygous FH, the deleterious influence of altered apoA-I metabolism on HDL-C levels is further aggravated by decreased rates of apoA-I production (Schaefer et al., 1992). Moreover, elevated CETP activity due to the increased number of apoB-containing lipoproteins—mainly LDL—also contributes to depletion of CE from the plasma HDL pool in FH (Guerin et al., 1995, 2000b).

HDL heterogeneity and particle profile largely reflect abnormalities in HDL metabolism. In the atherogenic dyslipidemias of MetS and type 2 diabetes, circulating levels of large, cholesterol-rich HDL decrease in parallel with decrease in HDL-C (Syvanne et al., 1995; Blatter Garin et al., 2005). By contrast, levels of small, dense, cholesterol-poor HDL particles and their content of apoA-I are rarely reduced in low HDL-C dyslipidemia (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005). Consistent with this observation, the relative apoA-I contents of HDL2b and HDL2a decrease, whereas those of pre- $\beta$ 1, pre- $\beta$ 2, HDL3a, HDL3b, and HDL3c increase in hypercholesterolemia, hypertriglyceridemia, and mixed hyperlipidemia (Ishida et al., 1987; Xu and Fu, 2003; Yang et al., 2005). Furthermore, plasma concentrations of small pre- $\beta$ -HDL are increased in hypercholesterolemia, hypertriglyceridemia, LCAT deficiency, and CHD but not in CETP deficiency (Ishida et al., 1987; Miida et al., 1997). Finally, plasma levels of small 70-kDa HDL are elevated in mixed hyperlipidemia (Atmeh and Robenek, 1996). Familial low HDL-C dyslipidemia appears to represent the only low HDL-C phenotype that is characterized by the presence of reduced concentrations of small pre- $\beta$ -HDL particles (Soderlund et al., 2005).

In obesity and insulin resistance, frequent features of both MetS and type 2 diabetes, plasma levels of large HDL decrease in parallel with those of HDL-C, whereas levels of small HDL do not (Garvey et al., 2003; Festa et al., 2005; Goff et al., 2005; Okazaki et al., 2005). Levels of  $\alpha$ 1-HDL are lower and levels of  $\alpha$ 2-,  $\alpha$ 3-, and pre- $\beta$ 1-HDL are higher in obese subjects compared with lean control subjects (Sasahara et al., 1997). As a result, MetS, type 2 diabetes, obesity, and insulin resistance are all characterized by the prevalence of small, dense HDL in the HDL particle profile, indicating either impaired conversion from small to large HDL or accelerated turnover and remodeling of large HDL2-like particles.

Small, dense HDL also prevail in CHD patients. In male participants in the Framingham Offspring Study, subjects with CHD displayed higher levels of small pre- $\beta$ 1- and  $\alpha$ 3-particles and lower levels of large  $\alpha$ 1-, pre- $\alpha$ 1-, and pre- $\alpha$ 3-particles than subjects without CHD (Asztalos et al., 2004a). Similarly, subjects with new CV

events possessed higher levels of small pre- $\beta$ 1- and  $\alpha$ 3-HDL and lower levels of large  $\alpha$ 1-,  $\alpha$ 2-, pre- $\alpha$ 1-, and pre- $\alpha$ 2-HDL than subjects without such events in the Veterans Affairs HDL Intervention Trial (VA-HIT) study (Asztalos et al., 2005). CAD patients also display elevated levels of lipid-poor apoA-I (Suzuki et al., 2005). The increase in small HDL and decrease in HDL of intermediate size as measured by nuclear magnetic resonance are associated with CAD severity in men admitted for diagnostic coronary arteriography (Freedman et al., 1998). Small HDL also prevail in peripheral arterial disease (Mowat et al., 1997). By contrast, CETP deficiency elevates plasma levels of large HDL particles, particularly buoyant, apoE-containing HDL1 but in also the bulk of HDL2; levels of small particles are affected to a minor degree (Asztalos et al., 2004b). Similarly, large HDL prevail in patients with type 1 diabetes (Colhoun et al., 2002).

Abnormalities in HDL subfraction distribution in hypercholesterolemia include reduced levels of apoA-I in large HDL2b and HDL2a and elevated levels of apoA-I in small pre- $\beta$ 1, pre- $\beta$ 2, HDL3b, and HDL3c subfractions (Ishida et al., 1987; Xu and Fu, 2003). FH HDL are further characterized by elevated content of apoE in association with increased plasma levels of apoE-enriched large HDL1, which represents only a minor fraction of total HDL in normolipidemic individuals (Keidar et al., 1990).

### *C. Impaired High-Density Lipoprotein Biological Activities in Dyslipidemic and Inflammatory States*

**1. Cholesterol Efflux Capacity.** Alterations in HDL composition and metabolism as occur in dyslipidemia and inflammation are intimately associated with impaired biological activities (Fig. 5). However, data on HDL cholesterol efflux capacity in atherogenic dyslipidemia are conflicting. In primary hypertriglyceridemia associated with low HDL-C levels, TG-enriched HDL particles, whose intrinsic cholesterol efflux capacity from hepatoma Fu5AH cells is impaired, accumulate; the cholesterol efflux capacity of serum is also reduced (Brites et al., 2000). Similarly, LpA-I from hypertriglyceridemic patients with well-controlled type 2 diabetes exhibits decreased capacity to induce cholesterol efflux from adipose cells (Cavallero et al., 1995). TG-enriched HDL produced in vitro by coinubation of normal HDL with CETP are weak activators of cholesterol esterification by LCAT and poor donors of CE to HepG2 cells (Skeggs and Morton, 2002). Consistent with these findings, HDL capacity to deliver CE to hepatic cells through interaction with SR-BI diminishes as a result of HDL enrichment in TG (Greene et al., 2001). Furthermore, the presence of TG-rich lipoproteins may have deleterious consequences for HDL-mediated cellular cholesterol efflux as demonstrated by preincubation of lipid-loaded macrophages with TG-rich lipoproteins (Palmer et al., 2004).

By contrast, others have reported normal cholesterol efflux capacity of serum from hypertriglyceridemic subjects in Fu5AH cells, an observation that can be related to normal contents of HDL-PL, a key determinant of HDL-mediated efflux (Fournier et al., 2001). Furthermore, HDL from hypertriglyceridemic CAD patients with low HDL-C levels possess a normal capacity to extract cholesterol from smooth muscle cells (Uint et al., 2003). Consistent with these results, TG-enriched HDL are not deficient in cholesterol efflux properties from cholesterol-loaded J774 macrophages (Skeggs and Morton, 2002).

The intrinsic cholesterol efflux capacity of HDL is considerably impaired during inflammation. Cellular cholesterol efflux is largely mediated by apoA-I-containing HDL particles (Ohta et al., 1992); apoA-I replacement by SAA can therefore have a significant impact on efflux. Enrichment of HDL with SAA (up to high SAA contents of 86% of total HDL protein) results in increased HDL binding to, decreased cholesterol efflux capacity from, and increased selective CE uptake by macrophages (Banka et al., 1995; Artl et al., 2000). Importantly, SAA selectively impairs cholesterol efflux properties of small, dense HDL3 particles. Less pronounced enrichment of HDL with SAA in vivo (up to 27% of total HDL protein) does not influence cholesterol efflux but enhances HDL binding to macrophages (Banka et al., 1995).

The presence of SAA increases both HDL affinity to and selective CE uptake by macrophages but reduces affinity to and CE uptake by hepatocytes (Kisilevsky and Subrahmanyam, 1992; Artl et al., 2002; Cai et al., 2005); furthermore, SAA efficiently promotes cholesterol efflux from hepatoma cells (van der Westhuyzen et al., 2005). During inflammation, the number of binding sites for HDL-bound SAA increases on macrophages and decreases on hepatocytes; in addition, macrophage expression of ABCA1 is diminished (Baranova et al., 2002). Decreased PL contents in inflammatory HDL constitute another factor that contributes to deficient HDL cholesterol efflux properties as suggested by studies in patients with periodontitis (Pussinen et al., 2004). Together, these changes lead to a significant shift in the HDL-mediated cholesterol transport from hepatocytes toward macrophages under acute-phase conditions (Kisilevsky and Subrahmanyam, 1992). Biologically, such alterations may serve to redirect cholesterol to immune cells and to sites of injury and inflammation.

Similarly, acute-phase HDL from hamsters display diminished cholesterol efflux capacity from J774 macrophages, elevated cholesterol influx capacity, and decreased LCAT activity (Khovidhunkit et al., 2001). In vitro inactivation of LCAT in control HDL results in similar effects on cholesterol transport, identifying LCAT as another key player in the abnormal cholesterol transport properties of HDL particles in inflammatory states (Khovidhunkit et al., 2001).

Abnormal lipid composition may also impair cholesterol efflux properties of HDL particles, as demonstrated by the diminished capacity of large, CE-enriched HDL2 isolated from subjects with homozygous CETP deficiency to accept cholesterol from lipid-loaded mouse peritoneal macrophages (Ishigami et al., 1994). Normalization of the lipid composition of such HDL, as a result of the transfer of excess CE to SR-BI-overexpressing cells, improves HDL cholesterol efflux capacity (Kinoshita et al., 2004).

Oxidative modification represents another factor involved in the impairment of HDL cholesterol efflux capacity. In vitro oxidation of apoA-I by myeloperoxidase results in selective inhibition of ABCA1-dependent cholesterol efflux from macrophages (Bergt et al., 2004; Zheng et al., 2004); oxidation of Tyr-192 and Tyr-166 residues appears to specifically account for this effect (Shao et al., 2005a; Zheng et al., 2005). In parallel, the lipid-binding capacity of apoA-I is progressively impaired (Zheng et al., 2005). Similarly, both in vitro HDL oxidation by  $\text{Cu}^{2+}$  and HDL modification by acrolein decrease HDL-mediated cholesterol efflux from cultured cells (Rifici and Khachadurian, 1996; Shao et al., 2005b). Oxidized forms of cholesterol, including 7-ketocholesterol, may account for impaired cholesterol efflux from macrophage-derived foam cells mediated by apoA-I, such oxysterols act through alterations in cell membrane properties (Gelissen et al., 1999; Gaus et al., 2001). Finally, the cholesterol efflux capacity of apoA-I may be impaired as a consequence of nonenzymatic glycosylation (Fievet et al., 1995; Ferretti et al., 2005).

The central role of apoA-I in HDL-mediated cholesterol efflux is consistent with the deleterious role of apoA-I mutations. ApoA-I Oslo carrying the R160L substitution and apoA-I mutant carrying the P165R substitution, two naturally occurring apoA-I variants associated with low HDL-C levels, are less effective in promoting cholesterol efflux from smooth muscle cells compared with normal HDL (Daum et al., 1999). Both mutants display a reduced ability to activate LCAT. Furthermore, cholesterol efflux from human fibroblasts and murine peritoneal macrophages mediated by apoA-I Nichinan, a naturally occurring human apoA-I variant with a deletion of glutamic acid at codon 235, is reduced relative to normal apoA-I (Han et al., 1999; Huang et al., 2000). In addition, familial low HDL-C deficiency is characterized by reduced HDL3- and apoA-I-mediated cellular cholesterol efflux in the absence of abnormalities in cellular HDL3 binding (Marcil et al., 1999).

However, not all mutations in apoA-I lead to decreased cholesterol efflux capacity. ApoA-I Milano, a molecular variant of apoA-I characterized by the Arg173Cys substitution, displays potent capacity for cholesterol efflux, a unique feature that is related to the formation of apoA-I Milano homodimers with prolonged plasma residence time (Franceschini et al., 1999; Chiesa and Sirtori, 2003). Carriers of apoA-I Milano exhibit



severe hypoalphalipoproteinemia but are not at increased risk for premature CHD (Sirtori et al., 2001) (see below).

Finally, the capacity of HDL particles to extract cholesterol from peripheral cells may be impaired as a result of alterations in cellular HDL receptors, primarily ABCA1. Thus, individuals from families with ABCA1 mutations display lower levels of HDL-C, higher IMT, and lower cholesterol efflux from fibroblasts compared with matched control subjects (van Dam et al., 2002).

**2. Antioxidative Activity.** Recent evidence indicates that HDL particles are deficient in antioxidative activity in atherogenic dyslipidemias involving low HDL-C levels (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005). Thus, the antioxidative activity of small, dense HDL subfractions against LDL oxidation induced by 2,2'-azobis-(2-amidinopropane) hydrochloride is significantly impaired in patients with MetS (up to -23%) (Hansel et al., 2004) and well-controlled type 2 diabetes (up to -47%) (Nobecourt et al., 2005). HDL antioxidative activity is deficient both on a unit particle mass and on a particle number basis. In another study, large, light HDL2 show decreased protection of LDL against oxidation mediated by THP1 macrophages in poorly controlled type 2 diabetes (Gowri et al., 1999). The impaired antioxidative activity of small, dense HDL in MetS and type 2 diabetes is intimately related to the concomitant presentation of hypertriglyceridemia, hyperinsulinemia, and insulin resistance, thereby suggesting that abnormalities in both lipid and glucose metabolism underlie the antioxidative deficiency of HDL particles (Hansel et al., 2004; Nobecourt et al., 2005). Furthermore, all HDL subfractions from subjects with a normotriglyceridemic, normocholesterolemic, normoglycemic low HDL-C phenotype display lower antioxidative activity (up to -43%) than their counterparts from normolipidemic control subjects (Kontush et al., 2005). Interestingly, the intrinsic antioxidative activity of HDL particles is equally reduced in subjects with hyperalphalipoproteinemia associated with low HL activity and high HDL-TG content (Kontush et al., 2004).

The antioxidative HDL deficiency in low HDL-C dyslipidemias of MetS and type 2 diabetes and in a normotriglyceridemic low HDL-C phenotype is paralleled by decreased enzymatic activities and altered physicochemical properties of HDL (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005), thereby suggesting that the intrinsic properties of HDL particles, rather than low HDL-C levels per se, are determinants of antioxidative deficiency of HDL3 subfractions. In each study population (MetS, type 2 diabetes, and normotriglyceridemic low HDL-C phenotype), HDL3 subfractions were enriched in TG and CE depleted (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005), potentially reflecting elevated CETP activity and/or reduced HL activity; these alterations correlated with the diminished antioxidative activity of HDL3 subfractions.

Mechanistically, the relationship between TG enrichment of HDL particles and impairment of antioxidative activity can be explained by the fact that the replacement of CE by TG in the HDL lipid core considerably alters the conformation of the central and C-terminal domains of apoA-I, which are critical for HDL to act as an acceptor of oxidized lipids (Sparks et al., 1995; Curtiss et al., 2000). Moreover, replacement of CE by TG in spherical rHDL decreases the conformational stability of apoA-I (Sparks et al., 1995), resulting in TG-containing particles, which are unstable and which lose apoA-I upon storage.

Replacement of apoA-I by acute-phase proteins, primarily SAA, in small, dense HDL particles under conditions of chronic inflammation (Van Lenten et al., 2001a) may represent another mechanism contributing to the impairment of HDL antioxidative activity. As in the case of the replacement of CE by TG, the replacement of apoA-I by SAA may cause deficient activity of HDL as an acceptor of oxidized PL, resulting in their elevated accumulation in LDL.

Altered enzymatic activities also contribute to the antioxidative deficiency of small, dense HDL. PAF-AH and PON1 activities are consistently lower in all HDL subfractions from patients with type 2 diabetes compared with matched normolipidemic control subjects (Nobecourt et al., 2005). Moreover, PAF-AH and PON1 activities positively correlate with HDL3 antioxidative activity (Nobecourt et al., 2005), suggesting that these enzymes are implicated in the deficiency of HDL antioxidative function. In type 1 diabetes, serum concentrations of PON1 are reduced to such an extent that diminished oxidative protection of LDL by HDL in vitro results (Boemi et al., 2001). Consistent with this mechanism, inactivation of HDL-associated enzymes, such as PON1 or LCAT, by oxidation and/or glycation leads to decreased capacity of HDL to protect LDL from oxidative stress (Hedrick et al., 2000; Ferretti et al., 2001; Jaouad et al., 2003; Ferretti et al., 2005). The role of enzymes in HDL antioxidative deficiency is also consistent with data obtained in obese leptin-deficient (*ob/ob*), LDL-R<sup>-/-</sup> mice that possess dysfunctional HDL displaying not only decreased PON1 and LCAT activities, but also elevated levels of antibodies against oxLDL (Mertens et al., 2003).

An antioxidative deficiency of HDL may also be observed when antioxidative activity is measured in total HDL, rather than in individual HDL subfractions. Total HDL from humans and rabbits lose the ability to protect LDL against oxidation by artery wall cells in coculture during induction of the acute phase, concomitant with decreases in PON1 and PAF-AH activities (Watson et al., 1995a,b). Total HDL from mice that are genetically predisposed to diet-induced atherosclerosis do not protect LDL against oxidation in cocultures of artery wall cells when the mice are fed an atherogenic diet, injected with LDL-derived oxidized PL, or infected with influ-

enza A virus (Shih et al., 1996; Navab et al., 1997; Van Lenten et al., 2001b). Such loss of antioxidative activity of murine HDL is accompanied by decrease in PON1 activity. In addition, antioxidative deficiency of total HDL is observed in both apoE<sup>-/-</sup> (Navab et al., 1997) and apoA-II transgenic (Warden et al., 1993; Castellani et al., 1997) mice.

Furthermore, total HDL-mediated protection of LDL from oxidation by Cu<sup>2+</sup> is compromised in postmenopausal compared with premenopausal women (Zago et al., 2004). This effect is paralleled by decreased plasma levels of HDL-C, elevated HDL levels of TG, and increased HDL oxidability in the postmenopausal group; the two latter parameters are significantly correlated (Zago et al., 2004). By contrast, serum PON1 activity did not differ between the groups, lending further support to our hypothesis that alterations of HDL core lipid composition are a key determinant of the antioxidative function of HDL particles. In addition, the antioxidative activity of total HDL toward LDL oxidation by  $\gamma$ -radiolysis of water is attenuated in elderly compared with young subjects (Jaouad et al., 2005). Finally, a diminished capacity of HDL to remove lipid hydroperoxides from erythrocyte membranes and attenuated HDL PON1 activity are features of poorly equilibrated type 1 diabetes (Ferretti et al., 2004).

In another study, no difference in HDL antioxidative activity, chemical composition, and serum PON activity was detected between type 2 diabetic patients with glycemic control and healthy control subjects (Sanguinetti et al., 2001). Similarly, patients with renal disease do not display antioxidatively deficient HDL despite low serum PON1 activity and low HDL levels of CE and  $\alpha$ -tocopherol (Hasselwander et al., 1999). The high albumin content of HDL in this study is, however, noteworthy. These negative results suggest that measurement of the antioxidative activity of small, dense HDL may provide a more sensitive estimation of HDL antioxidative activity in some patient populations compared with measurements performed on total HDL.

Despite the fact that HDL antioxidative deficiency has been extensively documented in atherogenic dyslipidemias using in vitro assays, direct evidence for its presence in vivo is still lacking. Indirect evidence includes an association between elevated levels of plasma HDL and reduced levels of lipid peroxidation products after a single intravenous injection of a large dose of human HDL3 (200 mg of protein) in hypercholesterolemic rabbits (Klimov et al., 1993), suggesting compromised antioxidative activity of autologous rabbit HDL under conditions of hypercholesterolemia.

**3. Anti-Inflammatory Activity.** HDL particles possessing antioxidative activity within the normal range can prevent formation of or inactivate proinflammatory oxidized PL produced during LDL oxidation and are therefore anti-inflammatory (Navab et al., 2001a,b).

Such potent anti-inflammatory activity becomes deficient and even transforms into in vitro pro-inflammatory action under conditions favoring development of atherosclerosis. In contrast with functional HDL, proinflammatory dysfunctional HDL is unable to protect LDL from oxidation by arterial wall cells and to prevent monocyte migration induced by oxLDL (Navab et al., 2001a,b). Total HDL from CHD patients with normal or elevated HDL-C levels are proinflammatory in both cell culture and cell-free fluorescent assays (Ansell et al., 2003). Similarly, HDL from mice that are genetically predisposed to diet-induced atherosclerosis become proinflammatory when the mice are fed an atherogenic diet, injected with LDL-derived oxidized PL or infected with influenza A virus (Shih et al., 1996; Navab et al., 1997; Van Lenten et al., 2001b). Proinflammatory HDL can also be detected in apoE<sup>-/-</sup> mice (Navab et al., 1997). In addition, transgenic mice overexpressing apoA-II possess proinflammatory HDL and develop atherosclerosis on a chow diet (Warden et al., 1993; Castellani et al., 1997). Finally, the anti-inflammatory activity of HDL is diminished in patients with obstructive sleep apnea subjected to repetitive cycles of hypoxia/reoxygenation (Tan et al., 2005); such patients frequently exhibit the MetS.

Formation of proinflammatory HDL correlates with decreases in the activities of various HDL-associated enzymes, such as PON1, PAF-AH, and LCAT, which are replaced by acute-phase proteins, such as SAA and ceruloplasmin; indeed, the content of these proteins in HDL increases during an inflammatory response (Navab et al., 2001a,b). Copper-containing ceruloplasmin may provide a source of transition metals for oxidative reactions, thereby accounting for the enhancement of LDL modification by acute-phase HDL (Navab et al., 2001a,b). Purified ceruloplasmin can induce LDL oxidation due to its content of a loosely bound copper atom (Ehrenwald et al., 1994; Ehrenwald and Fox, 1996); the in vitro enrichment of HDL with ceruloplasmin abrogates the ability of HDL to inhibit LDL modification in aortic wall cell cocultures (Van Lenten et al., 1995).

Interestingly, HDL PON1 activity is not decreased in CHD patients possessing proinflammatory HDL (Ansell et al., 2003), indicating that factors other than PON1 determine anti-inflammatory HDL dysfunction in vitro. In this study, plasma TG levels were markedly elevated (+76%) in CHD patients compared with control subjects (Ansell et al., 2003), suggesting that concomitant HDL enrichment in TG might have significantly contributed to the formation of dysfunctional HDL as proposed elsewhere (Hansel et al., 2004; Kontush et al., 2004, 2005; Nobecourt et al., 2005).

Intriguingly, all of the alterations in HDL composition that lead to attenuated anti-inflammatory and antioxidative activities (depletion in CE, apoA-I, PON1, and LCAT and increase in TG and SAA) are observed

during inflammation and in the acute-phase response (Khovidhunkit et al., 2004b; Esteve et al., 2005). The pro-inflammatory and prooxidative rearrangement of HDL particles and formation of LDL-derived oxidized PL have been hypothesized to form part of an evolutionary conserved mechanism of nonspecific innate immunity aimed to protect against infection (Navab et al., 2001a,b). Such an innate inflammatory response may include subnormal levels of HDL-C, increased HDL-TG content, and altered HDL apolipoprotein composition, all of which impair cholesterol efflux capacity as well as the antioxidant and anti-inflammatory activities of HDL particles. These modifications in HDL may be aimed to redirect cholesterol from the liver to immune cells, particularly macrophages, during infection (Kisilevsky and Subrahmanyam, 1992). Such a response to acute infection or injury can be advantageous in the short term but may become maladaptive in the long term. A sustained response that is not able to repair the injury, such as an emerging atherosclerotic plaque, which can be considered as a local inflammation (Libby, 2002), may lead to a chronic alteration in plasma lipid levels; such a response may become harmful, accelerating the formation of atherosclerotic lesions (Esteve et al., 2005). This mechanism is consistent with a recent hypothesis that accelerated development of atherosclerosis in old age is related to increased inflammation and concomitant endothelial dysfunction during early life (Finch and Crimmins, 2004; Charakida et al., 2005; Napoli et al., 2005). Within this concept, classic lipid changes associated with MetS (low HDL-C and elevated TG levels) are envisioned as a highly conserved evolutionary response aimed to repair tissue (Esteve et al., 2005).

It is indeterminate as to whether deficient anti-inflammatory activity of HDL is selectively associated with a subset of HDL particles as suggested by studies with rHDL (Nanjee et al., 1999; Nicholls et al., 2005b). The specific association of the deficient antioxidative activity with small, dense HDL (Hansel et al., 2004; Kontush et al., 2004, 2005; Nobecourt et al., 2005) together with the direct mechanistic link between anti-inflammatory and antioxidative activities (Navab et al., 2001a,b) suggests that small, dense HDL is a major subset of the total HDL particle population, which is responsible for both potent anti-inflammatory activity under normal conditions and for deficient activity under pro-atherogenic, pro-inflammatory conditions. This conclusion is consistent with the corrected anti-inflammatory properties of HDL from PLTP-deficient mice, which are characterized by the prevalence of small, lipid-poor HDL particles (Yan et al., 2004). Intriguingly, small, dense HDL<sub>3c</sub> represents the major HDL subfraction in newborns (Kherkeulidze et al., 1991), consistent with an elevated need for protection against infection in early life.

#### IV. Physiological Relevance of Defective High-Density Lipoprotein Function in Dyslipidemia and Metabolic Disease

The attenuated atheroprotective properties of HDL in metabolic disease raise the possibility of an indirect putative proatherogenic effect of these particles. Indeed, attenuated cholesterol efflux capacity of HDL can result in enhanced accumulation of cholesterol in the arterial wall and reduced RCT flux. Reduced efficiency of cholesterol flux through the RCT pathway is thought to account for the epidemiological link between subnormal HDL-C levels and increased incidence of CV disease (Nofer et al., 2002; Assmann and Nofer, 2003; Assmann and Gotto, 2004; Navab et al., 2004b). Impaired RCT has been shown to lead to accelerated atherosclerosis in subjects with Tangier disease (Oram, 2000) and in some cases of LCAT deficiency (Kuivenhoven et al., 1997). However, no data are available to our knowledge on the direct link between atherogenesis and the cholesterol efflux capacity of HDL particles, although infusion of apoA-I Milano/phospholipid complex has been shown to lead to a reduction in atheroma volume in patients with acute coronary syndromes, suggestive of plaque cholesterol efflux (Nissen et al., 2003) (see below).

A deficiency in the antioxidative and anti-inflammatory properties of HDL may also result in accelerated atherosclerosis. The oxidation hypothesis of atherosclerosis postulates that oxidation of lipoproteins, primarily LDL, in the arterial wall is a key element in atherogenesis (Steinberg et al., 1989). The validity of this statement has been confirmed in innumerable studies (Chisolm and Steinberg, 2000; Steinberg and Witztum, 2002). Its important corollary is that deficient LDL protection from oxidation may accelerate atherogenesis. Our recent data indicate clearly that impairment of the antioxidative activity of small, dense HDL in dyslipidemias involving low HDL-C levels is intimately associated with elevated oxidative stress, a newly recognized CV risk factor (Schwedhelm et al., 2004; Meisinger et al., 2005), and may therefore contribute to enhanced atherogenesis (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005). Indeed, dyslipidemic subjects presenting with atherogenic low HDL-C levels (MetS, type 2 diabetes, and a normotriglyceridemic low HDL-C phenotype) are characterized by both deficient antioxidative activity of small, dense HDL (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005) and elevated systemic oxidative stress assessed as plasma levels of 8-isoprostanes, products of nonenzymatic oxidation of arachidonic acid (Davi et al., 1999; Devaraj et al., 2001; Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005). Furthermore, HDL antioxidative activity and plasma 8-isoprostanes are negatively correlated (Hansel et al., 2004; Nobecourt et al., 2005). In addition, in subjects with controlled type 2 diabetes, plasma 8-isoprostanes negatively correlate with HDL-C levels (No-

becourt et al., 2005), whereas in subjects with a normotriglyceridemic low HDL-C phenotype, plasma 8-isoprostanes positively correlate with an elevated ratio of total cholesterol/HDL-C, thereby reflecting an excess of atherogenic nonHDL-C relative to antiatherogenic HDL-C levels (Kontush et al., 2005). The elevation of plasma 8-isoprostanes in subjects with low HDL-C dyslipidemias is consistent with the elevation of F<sub>2</sub> $\alpha$ -isoprostanes in apoA-I deficient mice, emphasizing the link between oxidative stress and HDL deficiency (Moore et al., 2003). Mechanistically, HDL enrichment in TG may play a role in both elevated oxidative stress and the deficiency in HDL antioxidative activity, as suggested by strong association between plasma levels of oxLDL and the TG/HDL-C molar ratio in elderly subjects (Holvoet et al., 2003).

The presence of antioxidatively deficient HDL can facilitate or even trigger accumulation of LDL-derived proinflammatory oxidized PL in vivo, resulting in compromised anti-inflammatory activity (Ansell et al., 2003). Functional small, dense HDL particles may in turn provide protection of LDL against oxidative stress in the subendothelial space of the arterial wall via removal of oxidized lipids from LDL, with inactivation and subsequent transfer to the liver mediated by SR-BI. This mechanism may account, at least in part, for the negative results of recent large-scale placebo-controlled trials that did not show any beneficial effect of low-molecular-weight antioxidants, primarily vitamin E, on the development of CV disease (Stocker and Keaney, 2004). The Nutrition Committee of the American Heart Association Council on Nutrition, Physical Activity and Metabolism has recently concluded that "the existing scientific database does not justify routine use of antioxidant supplements for the prevention and treatment of CV disease" (Kris-Etherton et al., 2004). Moreover, a meta-analysis of performed trials suggests that supplementation with vitamin E may even increase all-cause mortality (Miller et al., 2005). We interpret these data to indicate that low-molecular-weight antioxidants do not play a key role in the protection of LDL from oxidation in vivo; by contrast, small, dense HDL may constitute a central element of such protection.

The impaired antioxidative activity of small, dense HDL particles in atherogenic dyslipidemia is intimately linked to the presence of a constellation of CV risk factors, including hypertriglyceridemia, hyperglycemia, hyperinsulinemia, insulin resistance, and a disequilibrium between circulating levels of atherogenic apoB-containing lipoproteins and antiatherogenic HDL in favor of the former (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005). All of these factors are independently characterized by their significant association with elevated systemic oxidative stress (Morrow, 2005). Such correlational data (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005) strongly suggest then that small, dense HDL particles function as a biosensor of

oxidative stress, integrating a wide spectrum of prooxidant signals; the integration of such signals is in turn expressed as attenuated HDL antioxidative activity. Diagnostic detection of small, dense HDL possessing deficient antioxidative activity may therefore serve as a novel biomarker to assess elevated CV risk.

### V. Functionally Defective Small, Dense High-Density Lipoprotein as a Therapeutic Target

Subjects at elevated CV risk in primary prevention and patients in secondary prevention with symptomatic coronary atherosclerosis possess small HDL<sub>3</sub> particles whose antiatherogenic properties are impaired. Such defective functionality of small, dense HDL is frequently paralleled by decreased levels of HDL-C, which can principally be accounted for on the one hand by subnormal levels of large, cholesterol-rich HDL<sub>2</sub>-like particles and on the other by altered particle structure and composition. The association between low HDL-C levels and functional deficiency of small, dense HDL particles led us to propose that the deficient antioxidative activity of HDL can be corrected and concomitantly that elevated oxidative stress and attenuated HDL anti-inflammatory activity can be normalized by therapeutic approaches targeted to raise HDL-C and apoA-I levels and to normalize HDL structure and composition (Ashen and Blumenthal, 2005; Nicholls et al., 2005c).

Approaches to raise HDL-C levels may involve up-regulation of apoA-I synthesis in hepatocytes, increased lipidation of apoA-I, accelerated efflux of cholesterol and PL from peripheral cells mediated by ABCA1, decreased activity of CETP, which results in the diminished heteroexchange of CE and TG between HDL and TG-rich lipoproteins, and inhibition of HDL<sub>2</sub> holoparticle uptake by the liver mediated by hitherto unidentified receptor(s) for HDL holoparticles (Fig. 6). Such approaches are focused on small molecules whose pharmacological action results in marked raising of HDL-C, such as a CETP inhibitor or niacin, either alone or in combination with a statin. In this way, the LDL-C/HDL-C ratio may be reduced in dyslipidemic subjects, together with normalization of HDL metabolism, composition, and anti-atherogenic function. Such normalization can result from 1) a decrease in plasma TG levels and concomitant replacement of TG by CE in the HDL core and normalization of apoA-I conformation and function and/or 2) a decrease in the level of oxidative stress and inflammation potentially involving replacement of SAA by apoA-I with normalization of intravascular HDL particle remodeling. Therapeutic raising of HDL levels is intimately associated with slowed progression of atherosclerosis and reduced CV risk, as observed in large-scale clinical studies such as the Armed Forces Regression Study (AFREGS) (Personius et al., 1998), the Bezafibrate Infarction Prevention Trial (BIP) [The Bezafibrate

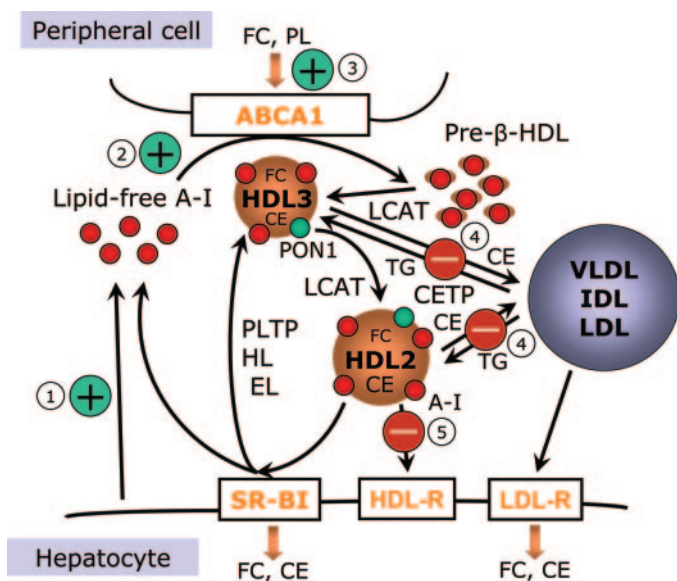


FIG. 6. Potential targets for therapeutic normalization of abnormal metabolism and deficient biological activities of HDL in atherosclerotic dyslipidemias of metabolic disease. ①, up-regulation of apoA-I synthesis in hepatocytes. ②, enhanced lipidation of apoA-I. ③, accelerated efflux of cholesterol and PL from peripheral cells mediated by ABCA1. ④, decreased activity of CETP, which results in the diminished heteroexchange of CE and TG between HDL and TG-rich lipoproteins. ⑤, inhibition of HDL2 holoparticle uptake by the liver mediated by hitherto unidentified receptor(s) for HDL holoparticles. EL, endothelial lipase; FC, free cholesterol; HDL-R, HDL holoparticle receptor; LDL-R, LDL receptor.

Infarction Prevention (BIP) Study, 2000], the VA-HIT (Robins et al., 2001), the HDL-Atherosclerosis Treatment Study (HATS) (Brown et al., 2001), and the Arterial Biology for the Investigation of the Treatment Effects of Reducing Cholesterol (ARBITER) 2 trial (Taylor et al., 2004).

#### A. Cholesteryl Ester Transfer Protein Inhibitors

CETP inhibitors are promising therapeutic agents that markedly decrease the activity of plasma CETP (Le Goff et al., 2004; van der Steeg et al., 2004). The therapeutic strategy to inhibit CETP and thereby raise HDL-C derives from the fact that genetic deficiency of CETP is associated with increased HDL-C and decreased LDL-C levels, a profile that is typically antiatherogenic (Le Goff et al., 2004). Human subjects with heterozygous CETP deficiency and HDL-C levels >60 mg/dl exhibit a reduced risk of CHD (Curb et al., 2004). Such subjects display elevated levels of  $\alpha$ 1- and reduced levels of  $\alpha$ 3- and pre $\beta$ -1-HDL subfractions, a profile consistent with corrected HDL antiatherogenicity (Asztalos et al., 2004b). Partial inhibition of CETP may therefore be atheroprotective; by contrast, the complete absence of CETP activity can create a potentially proatherogenic lipid profile (Brewer, 2004; Le Goff et al., 2004). The formation of large HDL particles with attenuated antiatherogenic activity and of heterogeneous LDL particles that are characteristic of lipoproteins in homozygous CETP-deficient patients can be avoided by partial inhibition of CETP. Studies in rabbits, a species with natu-

rally high levels of CETP, support the therapeutic potential of partial CETP inhibition as an approach to retarding or even reversing atherogenesis (Barter et al., 2003b; Gaofu et al., 2005).

Small-molecule inhibitors of CETP have now been tested in human subjects and shown to increase the concentration of HDL-C while decreasing those of LDL-C and apoB. In healthy young normolipidemic subjects, torcetrapib dose-dependently inhibits CETP activity by 12 to 80% and increases HDL-C by 28 to 91% at 30 and 240 mg/day, respectively (Clark et al., 2004). With respect to modulation of apolipoprotein levels, apoA-I and apoE were elevated by 27 and 66%, respectively, whereas apoB was reduced by 26% at a dose of 240 mg/day in these studies. Significantly, CETP inhibition led to a reduction in the TG content of HDL particles and an increase in the CE content (Clark et al., 2004). In subjects with low levels of HDL-C, treatment with 120 and 240 mg of torcetrapib daily raised plasma HDL-C concentrations by 46 and 106%, respectively (Brousseau et al., 2004).

Torcetrapib treatment led to elevations in HDL2-C (+87%) to a greater extent than HDL3-C (+29%), increased plasma apoA-I and apoA-II and reduced plasma TG levels. In addition, torcetrapib altered the distribution of cholesterol among HDL and LDL subclasses, resulting in an increase in the mean particle size of both HDL and LDL particles in each cohort. Finally, torcetrapib increased the amount of apoA-I in  $\alpha$ 1-HDL and apoA-I pool size and decreased apoA-I fractional catabolic rate, thereby prolonging the residence time of apoA-I in the circulation (Brousseau et al., 2005). It is plausible that such a beneficial modulation of HDL metabolism results from normalization of apoA-I lipidation and thus conformation subsequent to a normalized CE/TG core lipid ratio (Brousseau et al., 2005).

Another small-molecule CETP inhibitor, JTT-705, at a dose of 900 mg/day, inhibits CETP by 37% and increases HDL-C by 34%, apoA-I by 15%, HDL2-C by 59%, and HDL3-C by 19% in subjects with mild hyperlipidemia (de Grooth et al., 2002); comparable effects were observed in patients with type II dyslipidemia (Kuivenhoven et al., 2005) and with familial hypoalphalipoproteinemia (Bisoendial et al., 2005). In rabbits, JTT-705 given at a high dietary dose of 0.75% inhibits CETP activity, increases both HDL-C concentrations and the ratio of HDL2-C/HDL3-C, and decreases the fractional esterification rate of HDL-C, again indicating a preferential increase in large HDL particles (Zhang et al., 2004). Levels of apoE in HDL, serum PON activity, and HDL-associated PAF-AH activity also increase, whereas plasma lysophosphatidylcholine concentration decreases; enhanced apoE content in HDL particles may be of special relevance in the potentiation of their catabolism in the liver and peripheral tissues through the LDL receptor pathway. Similarly, JTT-705 at a high dose of 300 mg/kg daily in rabbits increased plasma total cho-

lesterol, HDL-C, HDL2-C, and HDL3-C and reduced HDL-TG and CETP activity but did not influence the cellular cholesterol efflux capacity of HDL (Kobayashi et al., 2002). Interestingly, torcetrapib at 120 mg increased HDL-C substantially more than JTT-705 at 900 mg (Brousseau et al., 2004).

Despite minor impact of CETP inhibitors on circulating levels of small HDL particles, HDL functionality may be considerably improved, as suggested by elevated HDL content of CE and decreased TG (Clark et al., 2004). This conclusion may be particularly relevant for HDL antioxidative activity, which strongly depends on the CE/TG ratio in HDL particles (Hansel et al., 2004; Kontush et al., 2005; Nobecourt et al., 2005). Consistent with this hypothesis, JTT-705 decreases circulating levels of oxLDL in familial hypoalphalipoproteinemia (Bisoendial et al., 2005), whereas CETP inhibition *in vitro* by a monoclonal antibody renders LDL more resistant to oxidation (Sugano et al., 2000), observations that could translate into improved HDL-mediated protection of LDL from oxidation *in vivo*. The critical role of TG metabolism in the mechanism of action of CETP inhibitors is also supported by recent data suggesting that CETP inhibition may be especially effective in reducing CV risk in patients with elevated TG levels (Wolfe and Rader, 2004).

### B. Niacin

Nicotinic acid (niacin), a vitamin of the B complex, has been used for almost 50 years as a lipid-modulating drug. The primary action of nicotinic acid is to suppress lipolysis of triacylglycerol in adipose tissue via inhibition of the hormone-sensitive TG lipase (Ganji et al., 2003; Rosenson, 2003; Karpe and Frayn, 2004; Meyers et al., 2004). The hormone-sensitive lipase is activated by re-

versible phosphorylation under the influence of protein kinase A. The antilipolytic action of nicotinic acid involves reduction of intracellular cyclic AMP levels in adipose tissue via a G-protein-coupled receptor that mediates inhibition of adenylyl cyclase. The recently discovered orphan G-protein-coupled receptor (HM74) in man has been identified as the nicotinic acid receptor in adipose tissue (Tunaru et al., 2003). HM74 appears to function as a low-affinity receptor for nicotinic acid, whereas the shorter homologous form (HM74A) represents a high-affinity receptor (Wise et al., 2003).

The key feature of the mechanism of action of nicotinic acid on lipid metabolism involves attenuated adipose tissue lipolysis, resulting in reduction of circulating levels of NEFA (Fig. 7). NEFA flux to the liver constitutes the main substrate for hepatic TG synthesis; this TG may either be integrated into nascent VLDL particles and secreted into the circulation or alternatively may be stored in the form of intracellular lipid droplets in the hepatocyte. Nicotinic acid is therefore distinguished as the sole pharmacological agent that markedly lowers NEFA and, as a direct consequence, plasma VLDL-TG levels.

TG levels are strongly inversely correlated with levels of HDL-C (Chapman et al., 2004). Thus, a nicotinic acid-mediated reduction in plasma TG levels predictably leads to marked raising of HDL-C. This effect is intimately linked to the action of CETP. By attenuating CETP-mediated depletion of HDL-CE in hypertriglyceridemic states such as MetS and type 2 diabetes, the TG-lowering action of nicotinic acid favors retention of CE in HDL with normalization of the HDL neutral lipid content, an increase in particle size, and a prolongation of plasma HDL-apoA-I residence time *in vivo*, thus resulting in effective raising of HDL-C and apoA-I levels.

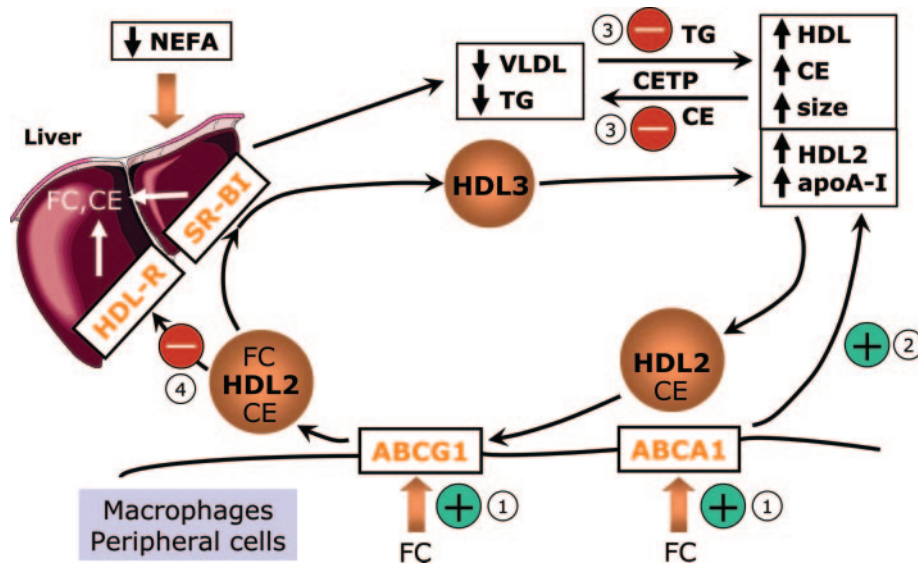


FIG. 7. Mechanisms involved in HDL raising by niacin. ① accelerated efflux of cholesterol from peripheral cells mediated by ABCA1 and ABCG1. ②, enhanced formation of mature HDL particles, primarily of HDL2. ③, diminished heteroexchange of CE and TG between HDL and TG-rich lipoproteins. ④, reduced uptake of HDL2 holoparticles by the liver mediated by receptor(s) for HDL holoparticles with maintained uptake of CE and free cholesterol from mature HDL by SR-BI. FC, free cholesterol; HDL-R, HDL holoparticle receptor.

A second mechanism that may contribute to nicotinic acid-induced raising of HDL-C involves the recent observation that this drug can stimulate cholesterol efflux from macrophages to primary HDL acceptors via the ABCA1 membrane transporter, thereby entering the RCT pathway (Fig. 7). Niacin activates ABCA1 via a nuclear peroxisome proliferator-activated receptor- $\gamma$ -dependent pathway (Rubic et al., 2004). In addition, nicotinic acid decreases HDL uptake by the liver (Sakai et al., 2001). Whether such actions lead to local, intraplaque depletion of cholesterol is conjectural, although this mechanism is consistent with the capacity of nicotinic acid to facilitate regression of coronary artery stenoses as observed in the HATS trial (Brown et al., 2001).

Clearly then, reduction in lipolysis in adipose tissue and in the NEFA supply to the liver are essential features of the pharmacological action of nicotinic acid in modifying the atherogenic lipid profile. As a result of these effects, nicotinic acid effectively decreases plasma levels not only of TG-rich lipoproteins, but also of small, dense LDL and lipoprotein(a), and also raises levels of HDL-C by a preferential decrease in CE flux to VLDL driven by CETP (Fig. 7) (Chapman et al., 2004; McKenney, 2004).

Importantly, niacin is presently the most effective commercially available agent for increasing HDL-C; the HDL-C-raising effect of niacin may reach 35% (Chapman et al., 2004). Meta-analysis of 30 randomized controlled trials reveals that niacin increases HDL-C on average by 16%; in parallel, niacin decreases plasma TG typically by 20% (Birjmohun et al., 2005). The action of niacin results in elevated plasma levels of both large and small HDL (McKenney et al., 2001; Morgan et al., 2003). In addition, niacin favorably modifies HDL composition, preferentially increasing apoA-I in the form of large HDL2-like, CE-rich particles (Sakai et al., 2001; Morgan et al., 2003). Such increases in HDL levels of apoA-I and CE at the expense of TG are consistent with the view that niacin, by virtue of its action in normalizing HDL structure and chemical composition but also in increasing HDL particle numbers and concentrations, may normalize deficient antiatherogenic functions of HDL particles in atherogenic dyslipidemia.

### C. Fibrates

Fibrates are peroxisome proliferator-activated receptor- $\alpha$  agonists that exert multiple effects on lipid and fatty acid metabolism and that also modulate the expression of genes of cellular cholesterol homeostasis, inflammation, and hemostasis (Steiner, 2005). Compared with CETP inhibitors and niacin, the average increase in HDL-C levels provided by fibrates is less pronounced and equaled +10% in 53 randomized, controlled trials (Birjmohun et al., 2005). The HDL-raising effect of fibrates is accompanied by a pronounced decrease in plasma levels of TG-rich lipoproteins of up to

48%. As a result, fibrates normalize HDL lipid composition, decreasing HDL-TG and increasing CE.

An alteration in the HDL subfraction profile is another central feature of fibrate therapy, which selectively increases circulating levels of small HDL particles, apoA-I and apoA-II. Fenofibrate (Sasaki et al., 2002; Ikewaki et al., 2004), bezafibrate (Miida et al., 2000; Kazama et al., 2003; Ikewaki et al., 2005), and gemfibrozil (Kahri et al., 1993) all selectively increase small and/or medium HDL particle numbers in patients with type 2 diabetes and in hypertriglyceridemic subjects. Remarkably, the increase in small HDL induced by fibrates may attain +168% in subjects with hypertriglyceridemia (Ikewaki et al., 2005). In addition, fenofibrate induces redistribution of PAF-AH from LDL to HDL in dyslipidemic patients, thereby lowering the proinflammatory potential of the enzyme (Tsimihodimos et al., 2003). Mechanistically, these effects can be accounted for by increased activities of both LPL, which provides release of surface fragments from TG-rich lipoproteins and their transfer to HDL during lipolysis, and of HL, which facilitates conversion of large to small HDL. Interestingly, plasma levels of small HDL3-C were a powerful predictor of CV risk in insulin-resistant subjects in the VA-HIT trial involving gemfibrozil treatment in insulin-resistant subjects (Robins et al., 2001). Fibrates may therefore be useful not only to induce an increase in circulating levels of HDL but also to enhance the functionality of small, dense HDL particles. The findings of the FIELD trial involving treatment of type 2 diabetes with fenofibrate and its effects on CV morbidity and mortality are eagerly awaited (FIELD Study Investigators, 2004).

### D. Statins

Statins are inhibitors of HMG-CoA reductase, whose major effect is to efficaciously decrease plasma levels of apoB-containing lipoproteins, primarily LDL, IDL, VLDL, and VLDL remnants. In addition, statins induce minor increases in HDL-C levels (by 5–10%) (Chong et al., 2002), consistent with a reduction in CETP activity (Guerin et al., 1995b, 2000b) and also with stimulation of apoA-I production (Schaefer et al., 1999). As a consequence, statins (atorvastatin and pravastatin) preferentially increase levels of HDL particles of large and medium size and  $\alpha$ -mobility (Otvos et al., 2002; Schaefer et al., 2002; Kazama et al., 2003; Soedamah-Muthu et al., 2003).

Importantly, statins exert a number of pleiotropic effects, which include anti-inflammatory and antioxidative activities. Antioxidative actions of statins involve increases in the activity of HDL-associated enzymes, such as that demonstrated for PON1 in type IIa hyperlipidemic patients treated with atorvastatin (Harangi et al., 2004). Beneficial effects of statins on HDL functionality therefore appear to be mediated by a decrease in CETP activity (Guerin et al., 1995b, 2000b), a reduction

in LDL-C levels (i.e., the numbers of LDL particles to be protected by HDL) (Chong et al., 2002), and a decrease in systemic oxidative stress (Harangi et al., 2004; Ceriello et al., 2005). Consistent with this mechanism, treatment of CHD patients with simvastatin at 40 mg/day for 4 weeks potently enhanced HDL functionality, rendering HDL anti-inflammatory (Ansell et al., 2003).

#### E. Reconstituted High-Density Lipoprotein

rHDL typically consist of apoA-I and PL but may also include apoE and other lipids. rHDL may provide an innovative approach to the management of CV disease by its ability to rapidly raise circulating HDL levels upon intravenous injection and to act as primary cholesterol acceptors at the arterial wall and in peripheral tissues, thereby facilitating RCT (Nanjee et al., 1999). Infusion of rHDL leads to inhibition of adhesion molecule expression, attenuation of endotoxin-induced release of proinflammatory cytokines, reduced ROS generation, enhanced NO bioavailability, restored impaired flow-mediated dilatation, and stabilized vulnerable plaque in dyslipidemic subjects (Spieker et al., 2002; Bisoendial et al., 2003) and/or in animal models (Cockerill et al., 2001a,b; Cuzzocrea et al., 2004; Nicholls et al., 2005a,b). In addition, rHDL favorably affects the distribution of antioxidative enzymes, particularly PAF-AH, between HDL and other lipoproteins (Kujiraoka et al., 2004).

Because of a potent cholesterol efflux capacity, rHDL containing apoA-I Milano offer an especially promising approach to treat CV disease (Sirtori et al., 1999), particularly under acute conditions at the diseased site, namely the vulnerable, unstable atherosclerotic plaque (Newton and Krause, 2002). Consistent with this notion, 5-weekly infusions of apoA-I Milano induced a 4.2% reduction in plaque volume in patients with acute coronary syndromes (Nissen et al., 2003). These promising findings indicate outperformance of typical reductions in plaque volume established after statin therapy, the most efficient approach at present to delay progression of atherosclerosis, e.g., 0.9% reduction after 18 months of intensive therapy with 80 mg of atorvastatin in the Reversal of Atherosclerosis with Aggressive Lipid Lowering (REVERSAL) study (Nissen et al., 2004; Birjmo-hun et al., 2005). In addition, apoA-I Milano-containing rHDL potently reduce atherosclerosis in atherosclerotic rabbits (Ameli et al., 1994; Soma et al., 1995) and mice (Shah et al., 1998, 2001). The regression of atherosclerosis induced by rHDL is most probably related to accelerated cholesterol efflux from the arterial wall with enhanced RCT to the liver (Nissen et al., 2003); the mechanistic relevance of other antiatherosclerotic activities of HDL to plaque regression remains unclear.

Importantly, injections of both apoA-I and apoA-I Milano result in the accumulation of small HDL particles (Nanjee et al., 1999), which also predominate in subjects with apoA-I Milano (Sirtori et al., 1999), thereby sug-

gesting that small apoA-I-containing HDL are particularly cardioprotective. Selective elevation in circulating concentrations of small HDL particles forms a basis for another approach to raise plasma HDL levels which involves reinfusion of HDL after selective delipidation (Sacks et al., 2004).

#### F. Apolipoprotein-Mimetic Peptides

Oral or intravenous administration of small amphipathic helical peptides that mimic HDL apolipoproteins represents another promising strategy to raise circulating HDL levels and to attenuate atherosclerosis (Navab et al., 2005b). Apolipoprotein-mimetic peptides typically include those derived from apoA-I (Navab et al., 2005b) but also from apoE (Gupta et al., 2005b) and apoJ (Navab et al., 2005c). As a result of their beneficial impact on HDL metabolism, apoA-I-mimetic peptides improve HDL-mediated cholesterol efflux, activate cholesterol efflux from macrophages, increase PON1 activity, convert HDL from proinflammatory to anti-inflammatory particles, increase endothelial production of nitric oxide, decrease endothelial production of superoxide, improve vasodilation, induce vascular heme oxygenase and superoxide dismutase, inhibit endotoxin-induced inflammatory responses, and reduce atherosclerosis in mice and monkeys (Garber et al., 2001; Navab et al., 2002; J. Ou et al., 2003; Z. Ou et al., 2003; Li et al., 2004; Navab et al., 2004a; Gupta et al., 2005a; Kruger et al., 2005).

Whereas typical apoA-I mimetics and apoA-I itself consist of L-amino acids, which are rapidly degraded in the digestive system and need to be supplemented parenterally, one of these peptides, D-4F, consists of D-amino acids, is not digested by mammalian enzymes, and can be administered orally (Navab et al., 2005b); the latter represents an important advantage in the development of apoA-I mimetics. Interestingly, covalent binding to an apoA-I mimetic of an apoE fragment critical for binding to the LDL receptor endows the resulting peptide with potent cholesterol-lowering capacity and further increases its antiatherogenic activity (Gupta et al., 2005b). Finally, orally administered small zwitterionic tetrapeptides, which are too small to form an amphipathic helix associate with HDL and display potent antiatherosclerotic, antioxidative, and anti-inflammatory activities in apoE<sup>-/-</sup> mice (Navab et al., 2005).

Significantly, apoA-I mimetics cause rapid formation of small, lipid-poor pre- $\beta$ -HDL, both in vivo when given to animal models and in vitro when added to human plasma (Navab et al., 2004a), consistent with a key antiatherosclerotic role for such particles. The anti-inflammatory properties of apoA-I mimetic peptides appear to depend on subtle differences in the configuration of the hydrophobic face of the peptides, which determines their ability to sequester pro-inflammatory oxidized lipids (Datta et al., 2004; Epanand et al., 2004).



### G. Combination Therapy

As distinct HDL-C raising agents function through complementary mechanisms, their effects may be additive; hence, association of such medications has been proposed. Another advantage of such combination therapy involves the potential for use of lower doses of each agent compared with their use in monotherapy to obtain additive elevations in HDL-C levels; such an approach may also lead to a reduction in adverse effects. Two trials evaluating niacin combined with either a statin or a bile acid sequestrant reported impressive HDL-C increases ranging from 25 to 41% with an unprecedented reduction in CV event rates ranging from 60 to 72% (Brown et al., 1995, 2001). Such effects can be accounted for by the additive benefit of concomitant reduction of LDL-C and raising of HDL-C. These impressive reduction rates also correspond well to the estimated reduction rates based on the increase in HDL-C obtained in these studies (Birjmohun et al., 2005) (i.e., 1% HDL-C increase being associated with a 1–3% reduction in CV events) (Robins et al., 2001). Furthermore, addition of extended-release niacin to statin therapy slows the progression of carotid atherosclerosis (measured as IMT), increases HDL-C (+21%), and decreases TG (–15%) and non-HDL-C (–7%) levels among individuals with established CHD and moderately low HDL-C (Taylor et al., 2004). Finally, one clinically important feature of the action of statins on HDL functionality involves their synergism with a CETP inhibitor (Brousseau et al., 2004, 2005) or apoA-I mimetic peptide (M. Navab et al., 2005a).

## VI. Conclusions

HDL particles possess potent biological activities, including cellular cholesterol efflux capacity, antioxidative, anti-inflammatory, antiapoptotic, antithrombotic, anti-infectious, and vasodilatory activities, which provide protection from atherosclerosis or may even favor plaque regression. Small, dense HDL afford potent protection of LDL against oxidative stress, possess pronounced anti-inflammatory properties, and display high cholesterol efflux capacity. The atheroprotective properties of HDL can, however, be compromised under conditions associated with accelerated development of atherosclerosis, such as in atherogenic low HDL-C dyslipidemias typical of metabolic diseases, including MetS and type 2 diabetes. Such functional HDL deficiency is intimately associated with alterations in HDL metabolism and structure. Formation of small, dense HDL particles with attenuated antioxidative activity is mechanistically related to HDL enrichment in TG and SAA, depletion of CE and apoA-I, and covalent modification of key HDL apolipoproteins. Deficiency of HDL function may result in accelerated atherosclerosis; therapeutic normalization of HDL function in terms of both the quantity and quality of HDL particles, using CETP in-

hibitors, niacin, rHDL or other agents, may therefore represent a novel therapeutic approach to attenuate atherosclerosis in dyslipidemic subjects with metabolic disease. Induction of selective increases in the circulating concentrations of HDL particles possessing normal antiatherogenic activity is especially promising; more specifically, recent studies suggest that small, dense HDL3 particles represent a new therapeutic target in atherogenic dyslipidemia, particularly in view of its intimate association with a proinflammatory state.

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## REFERENCES

- Ahmed Z, Ravandi A, Maguire GF, Emili A, Draganov D, La Du BN, Kuksis A, and Connelly PW (2001) Apolipoprotein A-I promotes the formation of phosphatidylcholine core aldehydes that are hydrolyzed by paraoxonase (PON-1) during high density lipoprotein oxidation with a peroxynitrite donor. *J Biol Chem* **276**:24473–24481.
- Alagona C, Soro A, Ylitalo K, Salonen R, Salonen JT, and Taskinen MR (2002) A low high density lipoprotein (HDL) level is associated with carotid artery intima-media thickness in asymptomatic members of low HDL families. *Atherosclerosis* **165**: 309–316.
- Ameli S, Hultgardh-Nilsson A, Cercek B, Shah PK, Forrester JS, Ageland H, and Nilsson J (1994) Recombinant apolipoprotein A-I Milano reduces intimal thickening after balloon injury in hypercholesterolemic rabbits. *Circulation* **90**:1935–1941.
- Anderson DW, Nichols AV, Forte TM, and Lindgren FT (1977) Particle distribution of human serum high density lipoproteins. *Biochim Biophys Acta* **493**:55–68.
- Ansell BJ, Navab M, Hama S, Kamranpour N, Fonarow G, Hough G, Rahmani S, Mottahedeh R, Dave R, Reddy ST, et al. (2003) Inflammatory/antiinflammatory properties of high-density lipoprotein distinguish patients from control subjects better than high-density lipoprotein cholesterol levels and are favorably affected by simvastatin treatment. *Circulation* **108**:2751–2756.
- Arakawa H, Qian J-Y, Baatar D, Karasawa K, Asada Y, Sasaguri Y, Miller ER, Witztum JL, and Ueno H (2005) Local expression of platelet-activating factor-acetylhydrolase reduces accumulation of oxidized lipoproteins and inhibits inflammation, shear stress-induced thrombosis, and neointima formation in balloon-injured carotid arteries in nonhyperlipidemic rabbits. *Circulation* **111**:3302–3309.
- Arthur JR (2000) The glutathione peroxidases. *Cell Mol Life Sci* **57**:1825–1835.
- Artl A, Marsche G, Lestavel S, Sattler W, and Malle E (2000) Role of serum amyloid A during metabolism of acute-phase HDL by macrophages. *Arterioscler Thromb Vasc Biol* **20**:763–772.
- Artl A, Marsche G, Pussinen P, Knipping G, Sattler W, and Malle E (2002) Impaired capacity of acute-phase high density lipoprotein particles to deliver cholesteryl ester to the human HUH-7 hepatoma cell line. *Int J Biochem Cell Biol* **34**:370–381.
- Artola RL, Conde CB, Bagatolli L, Pecora RP, Fidelio GD, and Kivatnitsch SC (1997) High-density lipoprotein from hypercholesterolemic animals has peroxidized lipids and oligomeric apolipoprotein A-I: its putative role in atherogenesis. *Biochem Biophys Res Commun* **239**:570–574.
- Ashby DT, Rye KA, Clay MA, Vadas MA, Gamble JR, and Barter PJ (1998) Factors influencing the ability of HDL to inhibit expression of vascular cell adhesion molecule-1 in endothelial cells. *Arterioscler Thromb Vasc Biol* **18**:1450–1455.
- Ashen MD and Blumenthal RS (2005) Clinical practice: low HDL cholesterol levels. *N Engl J Med* **353**:1252–1260.
- Assmann G and Gotto AM Jr (2004) HDL cholesterol and protective factors in atherosclerosis. *Circulation* **109**:III8–III14.
- Assmann G and Nofer JR (2003) Atheroprotective effects of high-density lipoproteins. *Annu Rev Med* **54**:321–341.
- Asztalos B, Zhang W, Roheim PS, and Wong L (1997) Role of free apolipoprotein A-I in cholesterol efflux: formation of pre- $\alpha$ -migrating high-density lipoprotein particles. *Arterioscler Thromb Vasc Biol* **17**:1630–1636.
- Asztalos BF, Batista M, Horvath KV, Cox CE, Dallal GE, Morse JS, Brown GB, and Schaefer EJ (2003) Change in  $\alpha 1$  HDL concentration predicts progression in coronary artery stenosis. *Arterioscler Thromb Vasc Biol* **23**:847–852.
- Asztalos BF, Collins D, Cupples LA, Demissie S, Horvath KV, Bloomfield HE, Robins SJ, and Schaefer EJ (2005) Value of high-density lipoprotein (HDL) subpopulations in predicting recurrent cardiovascular events in the Veterans Affairs HDL Intervention Trial. *Arterioscler Thromb Vasc Biol* **25**:2185–2191.
- Asztalos BF, Cupples LA, Demissie S, Horvath KV, Schaefer EJ, and Rothblat GH (2005) Differential effects of HDL subpopulations on cellular ABCA1- and SR-BI-mediated cholesterol efflux. *J Lipid Res* **46**:2246–2253.
- Asztalos BF, Horvath KV, Kajinami K, Nartsupha C, Cox CE, Batista M, Schaefer EJ, Inazu A, and Mabuchi H (2004b) Apolipoprotein composition of HDL in cholesteryl ester transfer protein deficiency. *J Lipid Res* **45**:448–455.
- Asztalos BF and Schaefer EJ (2003) HDL in atherosclerosis: actor or bystander? *Atheroscler Suppl* **4**:21–29.

- Asztalos BF, Sloop CH, Wong L, and Roheim PS (1993) Two-dimensional electrophoresis of plasma lipoproteins: recognition of new apo A-I-containing subpopulations. *Biochim Biophys Acta* **1169**:291–300.
- Atmeh RF (1990) Isolation and identification of HDL particles of low molecular weight. *J Lipid Res* **31**:1771–1780.
- Atmeh RF and Abd Elrazeq IO (2005) Small high density lipoprotein subclasses: some of their physico-chemical properties and stability in solution. *Acta Biochim Pol* **52**:515–525.
- Atmeh RG and Robenek H (1996) Measurement of small high density lipoprotein subclass by an improved immunoblotting technique. *J Lipid Res* **37**:2461–2469.
- Auerbach BJ and Parks JS (1989) Lipoprotein abnormalities associated with lipopolysaccharide-induced lecithin: cholesterol acyltransferase and lipase deficiency. *J Biol Chem* **264**:10264–10270.
- Aviram M, Rosenblat M, Bisgaier CL, Newton RS, Primo-Parmo SL, and La Du BN (1998) Paraoxonase inhibits high-density lipoprotein oxidation and preserves its functions: a possible peroxidative role for paraoxonase. *J Clin Invest* **101**:1581–1590.
- Bagdade JD, Ritter MC, and Subbiah PV (1991) Accelerated cholesteryl ester transfer in plasma of patients with hypercholesterolemia. *J Clin Invest* **87**:1259–1265.
- Baker PW, Rye KA, Gamble JR, Vadas MA, and Barter PJ (1999) Ability of reconstituted high density lipoproteins to inhibit cytokine-induced expression of vascular cell adhesion molecule-1 in human umbilical vein endothelial cells. *J Lipid Res* **40**:345–353.
- Banka CL, Yuan T, de Beer MC, Kindy M, Curtiss LK, and de Beer FC (1995) Serum amyloid A (SAA): influence on HDL-mediated cellular cholesterol efflux. *J Lipid Res* **36**:1058–1065.
- Baranova I, Vishnyakova T, Bocharov A, Chen Z, Remaley AT, Stonik J, Eggerman TL, and Patterson AP (2002) Lipopolysaccharide down regulates both scavenger receptor B1 and ATP binding cassette transporter A1 in RAW cells. *Infect Immun* **70**:2995–3003.
- Barter P (2004) HDL: a recipe for longevity. *Atheroscler Suppl* **5**:25–31.
- Barter P, Kastelein J, Nunn A, and Hobbs R (2003a) High density lipoproteins (HDLs) and atherosclerosis; the unanswered questions. *Atherosclerosis* **168**:195–211.
- Barter PJ, Brewer HB Jr, Chapman MJ, Hennekens CH, Rader DJ, and Tall AR (2003b) Cholesteryl ester transfer protein: a novel target for raising HDL and inhibiting atherosclerosis. *Arterioscler Thromb Vasc Biol* **23**:160–167.
- Barter PJ, Clay MA, and Rye KA (1999) High density lipoproteins: the anti-atherogenic fraction, in *Plasma Lipids and Their Role in Disease* (PJ Barter and KA Rye eds) vol 5, pp 85–107. Harwood Academic Publishers, Amsterdam.
- Barzilai N, Atzmon G, Schechter C, Schaefer EJ, Cupples AL, Lipton R, Cheng S, and Shuldiner AR (2003) Unique lipoprotein phenotype and genotype associated with exceptional longevity. *J Am Med Assoc* **290**:2030–2040.
- Beitz J and Forster W (1980) Influence of human low density and high density lipoprotein cholesterol on the in vitro prostaglandin I<sub>2</sub> synthetase activity. *Biochim Biophys Acta* **620**:352–355.
- Benditt EP and Eriksen N (1977) Amyloid protein SAA is associated with high density lipoprotein from human serum. *Proc Natl Acad Sci USA* **74**:4025–4028.
- Bergmeier C, Siekmeier R, and Gross W (2004) Distribution spectrum of paraoxonase activity in HDL fractions. *Clin Chem* **50**:2309–2315.
- Bergt C, Pennathur S, Fu X, Byun J, O'Brien K, McDonald TO, Singh P, Anantharamiah GM, Chait A, Brunzell J, et al. (2004) The myeloperoxidase product hypochlorous acid oxidizes HDL in the human artery wall and impairs ABCA1-dependent cholesterol transport. *Proc Natl Acad Sci USA* **101**:13032–13037.
- Birjmhoun RS, Hutten BA, Kastelein JJ, and Stroes ES (2005) Efficacy and safety of high-density lipoprotein cholesterol-increasing compounds: a meta-analysis of randomized controlled trials. *J Am Coll Cardiol* **45**:185–197.
- Bisgaier CL, Sachdev OP, Megna L, and Glickman RM (1985) Distribution of apolipoprotein A-IV in human plasma. *J Lipid Res* **26**:11–25.
- Bisoendial RJ, Hovingh GK, El Harchaoui K, Levels JHM, Tsimikas S, Pu K, Zwiderman AE, Kuivenhoven JA, Kastelein JJP, and Stroes ESG (2005) Consequences of cholesteryl ester transfer protein inhibition in patients with familial hypoalphalipoproteinemia. *Arterioscler Thromb Vasc Biol* **25**:e133–e134.
- Bisoendial RJ, Hovingh GK, Levels JH, Lerch PG, Andresen I, Hayden MR, Kastelein JJ, and Stroes ES (2003) Restoration of endothelial function by increasing high-density lipoprotein in subjects with isolated low high-density lipoprotein. *Circulation* **107**:2944–2948.
- Blades B, Vega GL, and Grundy SM (1993) Activities of lipoprotein lipase and hepatic triglyceride lipase in postheparin plasma of patients with low concentrations of HDL cholesterol. *Arterioscler Thromb* **13**:1227–1235.
- Blanche PJ, Gong EL, Forte TM, and Nichols AV (1981) Characterization of human high-density lipoproteins by gradient gel electrophoresis. *Biochim Biophys Acta* **665**:408–419.
- Blatter Garin MC, Kalix B, Morabia A, and James RW (2005) Small, dense lipoprotein particles and reduced paraoxonase-1 in patients with the metabolic syndrome. *J Clin Endocrinol Metab* **90**:2264–2269.
- Boekholdt SM, Kuivenhoven JA, Wareham NJ, Peters RJ, Jukema JW, Luben R, Bingham SA, Day NE, Kastelein JJ, and Khaw KT (2004) Plasma levels of cholesteryl ester transfer protein and the risk of future coronary artery disease in apparently healthy men and women: the prospective EPIC (European Prospective Investigation into Cancer and nutrition)-Norfolk population study. *Circulation* **110**:1418–1423.
- Boekholdt SM, Sacks FM, Jukema JW, Shepherd J, Freeman DJ, McMahon AD, Cambien F, Nicaud V, de Grooth GJ, Talmud PJ, et al. (2005) Cholesteryl ester transfer protein TaqIB variant, high-density lipoprotein cholesterol levels, cardiovascular risk, and efficacy of pravastatin treatment: individual patient meta-analysis of 13,677 subjects. *Circulation* **111**:278–287.
- Boemi M, Leviev I, Sirolla C, Pieri C, Marra M, and James RW (2001) Serum paraoxonase is reduced in type 1 diabetic patients compared to non-diabetic, first degree relatives; influence on the ability of HDL to protect LDL from oxidation. *Atherosclerosis* **155**:229–235.
- Boisfer E, Stengel D, Pastier D, Laplaud PM, Dousset N, Ninio E, and Kalopissis AD (2002) Antioxidant properties of HDL in transgenic mice overexpressing human apolipoprotein A-II. *J Lipid Res* **43**:732–741.
- Borggreve SE, De Vries R, and Dullaart RP (2003) Alterations in high-density lipoprotein metabolism and reverse cholesterol transport in insulin resistance and type 2 diabetes mellitus: role of lipolytic enzymes, lecithin:cholesterol acyltransferase and lipid transfer proteins. *Eur J Clin Invest* **33**:1051–1069.
- Bowry VW, Stanley KK, and Stocker R (1992) High density lipoprotein is the major carrier of lipid hydroperoxides in human blood plasma from fasting donors. *Proc Natl Acad Sci U.S.A.* **89**:10316–10320.
- Brewer HB Jr (2004) Increasing HDL cholesterol levels. *N Engl J Med* **350**:1491–1494.
- Brites FD, Bonavita CD, De Geitere C, Cloes M, Delfly B, Yael MJ, Fruchart J, Wikinski RW, and Castro GR (2000) Alterations in the main steps of reverse cholesterol transport in male patients with primary hypertriglyceridemia and low HDL-cholesterol levels. *Atherosclerosis* **152**:181–192.
- Brites FD, Verona J, Schreier LE, Fruchart JC, Castro GR, and Wikinski RL (2004) Paraoxonase 1 and platelet-activating factor acetylhydrolase activities in patients with low HDL-cholesterol levels with or without primary hypertriglyceridemia. *Arch Med Res* **35**:235–240.
- Brousseau ME, Diffenderfer MR, Millar JS, Nartsupha C, Asztalos BF, Welty FK, Wolfe ML, Rudling M, Bjorkhem I, Angelin B, et al. (2005) Effects of cholesteryl ester transfer protein inhibition on high-density lipoprotein subspecies: apolipoprotein A-I metabolism, and fecal sterol excretion. *Arterioscler Thromb Vasc Biol* **25**:1057–1064.
- Brousseau ME, Eberhart GP, Dupuis J, Asztalos BF, Goldkamp AL, Schaefer EJ, and Freeman MW (2000) Cellular cholesterol efflux in heterozygotes for Tangier disease is markedly reduced and correlates with high density lipoprotein cholesterol concentration and particle size. *J Lipid Res* **41**:1125–1135.
- Brousseau ME, Schaefer EJ, Wolfe ML, Bloedon LT, Digenio AG, Clark RW, Mancuso JP, and Rader DJ (2004) Effects of an inhibitor of cholesteryl ester transfer protein on HDL cholesterol. *N Engl J Med* **350**:1505–1515.
- Brown BG, Hillger L, Zhao XQ, Poulin D and Albers, JJ (1995) Types of change in coronary stenosis severity and their relative importance in overall progression and regression of coronary disease: observations from the FATS trial. Familial Atherosclerosis Treatment Study. *Ann NY Acad Sci* **748**:407–417; discussion 417–418.
- Brown BG, Zhao XQ, Chait A, Fisher LD, Cheung MC, Morse JS, Dowdy AA, Marino EK, Bolson EL, Alaupovic P, et al. (2001) Simvastatin and niacin, antioxidant vitamins, or the combination for the prevention of coronary disease. *N Engl J Med* **345**:1583–1592.
- Burger D and Dayer JM (2002) High-density lipoprotein-associated apolipoprotein A-I: the missing link between infection and chronic inflammation? *Autoimmun Rev* **1**:111–117.
- Cabana VG, Feng N, Reardon CA, Lukens J, Webb NR, de Beer FC, and Getz GS (2004) Influence of apoA-I and apoE on the formation of serum amyloid A-containing lipoproteins in vivo and in vitro. *J Lipid Res* **45**:317–325.
- Cabana VG, Lukens JR, Rice KS, Hawkins TJ, and Getz GS (1996) HDL content and composition in acute phase response in three species: triglyceride enrichment of HDL a factor in its decrease. *J Lipid Res* **37**:2662–2674.
- Cabana VG, Reardon CA, Feng N, Neath S, Lukens J, and Getz GS (2003) Serum paraoxonase: effect of the apolipoprotein composition of HDL and the acute phase response. *J Lipid Res* **44**:780–792.
- Cabana VG, Reardon CA, Wei B, Lukens JR, and Getz GS (1999) SAA-only HDL formed during the acute phase response in apoA-I<sup>+/+</sup> and apoA-I<sup>-/-</sup> mice. *J Lipid Res* **40**:1090–1103.
- Cabana VG, Siegel JN, and Sabesin SM (1989) Effects of the acute phase response on the concentration and density distribution of plasma lipids and apolipoproteins. *J Lipid Res* **30**:39–49.
- Cai L, de Beer MC, de Beer FC, and van der Westhuyzen DR (2005) Serum amyloid A is a ligand for scavenger receptor class B type I and inhibits high density lipoprotein binding and selective lipid uptake. *J Biol Chem* **280**:2954–2961.
- Calabresi L, Franceschini G, Sirtori CR, De Palma A, Saresella M, Ferrante P, and Taramelli D (1997) Inhibition of VCAM-1 expression in endothelial cells by reconstituted high density lipoproteins. *Biochem Biophys Res Commun* **238**:61–65.
- Calabresi L, Gomasrashi M, and Franceschini G (2003) Endothelial protection by high-density lipoproteins: from bench to bedside. *Arterioscler Thromb Vasc Biol* **23**:1724–1731.
- Castellani LW, Navab M, Van Lenten BJ, Hedrick CC, Hama SY, Goto AM, Fogelman AM, and Lusis AJ (1997) Overexpression of apolipoprotein AII in transgenic mice converts high density lipoproteins to proinflammatory particles. *J Clin Invest* **100**:464–474.
- Cavallero E, Brites F, Delfly B, Nicolaiew N, Decossin C, De Geitere C, Fruchart JC, Wikinski R, Jacotot B, and Castro G (1995) Abnormal reverse cholesterol transport in controlled type II diabetic patients: studies on fasting and postprandial LpA-I particles. *Arterioscler Thromb Vasc Biol* **15**:2130–2135.
- Ceriello A, Assaloni R, Da Ros R, Maier A, Piconi L, Quagliaro L, Esposito K, and Giugliano D (2005) Effect of atorvastatin and irbesartan, alone and in combination, on postprandial endothelial dysfunction, oxidative stress, and inflammation in type 2 diabetic patients. *Circulation* **111**:2518–2524.
- Chapman MJ, Assmann G, Fruchart JC, Shepherd J, and Sirtori C (2004) Raising high-density lipoprotein cholesterol with reduction of cardiovascular risk: the role of nicotinic acid—a position paper developed by the European Consensus Panel on HDL-C. *Curr Med Res Opin* **20**:1253–1268.
- Chapman MJ, Goldstein S, Lagrange D, and Laplaud PM (1981) A density gradient ultracentrifugal procedure for the isolation of the major lipoprotein classes from human serum. *J Lipid Res* **22**:339–358.
- Charakida M, Donald AE, Terese M, Leary S, Halcox JP, Ness A, Smith GD, Golding

- J, Friberg P, Klein NJ, et al. (2005) Endothelial dysfunction in childhood infection. *Circulation* **111**:1660–1665.
- Chen LY and Mehta JL (1994) Inhibitory effect of high-density lipoprotein on platelet function is mediated by increase in nitric oxide synthase activity in platelets. *Life Sci* **55**:1815–1821.
- Chen N, Liu Y, Greiner CD, and Holtzman JL (2000) Physiologic concentrations of homocysteine inhibit the human plasma GSH peroxidase that reduces organic hydroperoxides. *J Lab Clin Med* **136**:58–65.
- Chiesa G and Sirtori CR (2003) Apolipoprotein A-I (Milano): current perspectives. *Curr Opin Lipidol* **14**:159–163.
- Chisolm GM and Steinberg D (2000) The oxidative modification hypothesis of atherogenesis: an overview. *Free Radic Biol Med* **28**:1815–1826.
- Chong PH, Kezele R, and Franklin C (2002) High-density lipoprotein cholesterol and the role of statins. *Circ* **66**:1037–1044.
- Choudhury RP and Leyva F (1999) C-reactive protein, serum amyloid A protein, and coronary events. *Circulation* **100**:e65–e66.
- Choudhury RP, Rong JX, Trogan E, Elmalem VI, Dansky HM, Breslow JL, Witztum JL, Fallon JT and Fisher EA (2004) High-density lipoproteins retard the progression of atherosclerosis and favorably remodel lesions without suppressing indices of inflammation or oxidation. *Arterioscler Thromb Vasc Biol* **24**:1904–1909.
- Clark RW, Sutfin TA, Ruggeri RB, Willauer AT, Sugarman ED, Magnus-Aryitey G, Cosgrove PG, Sand TM, Wester RT, Williams JA, et al. (2004) Raising high-density lipoprotein in humans through inhibition of cholesteryl ester transfer protein: an initial multidosed study of torcetrapib. *Arterioscler Thromb Vasc Biol* **24**:490–497.
- Clay MA, Newnham HH, Forte TM, and Barter PJ (1992) Cholesteryl ester transfer protein and hepatic lipase activity promote shedding of apo A-I from HDL and subsequent formation of discoidal HDL. *Biochim Biophys Acta* **1124**:52–58.
- Clifton PM, Mackinnon AM, and Barter PJ (1985) Effects of serum amyloid A protein (SAA) on composition, size, and density of high density lipoproteins in subjects with myocardial infarction. *J Lipid Res* **26**:1389–1398.
- Cockerill GW, Huehns TY, Weerasinghe A, Stocker C, Lerch PG, Miller NE, and Haskard DO (2001a) Elevation of plasma high-density lipoprotein concentration reduces interleukin-1-induced expression of E-selectin in an in vivo model of acute inflammation. *Circulation* **103**:108–112.
- Cockerill GW, McDonald MC, Mota-Filipe H, Cuzzocrea S, Miller NE, and Thiemermann C (2001b) High density lipoproteins reduce organ injury and organ dysfunction in a rat model of hemorrhagic shock. *FASEB J* **15**:1941–1952.
- Cockerill GW, Rye KA, Gamble JR, Vadas MA, and Barter PJ (1995) High-density lipoproteins inhibit cytokine-induced expression of endothelial cell adhesion molecules. *Arterioscler Thromb Vasc Biol* **15**:1987–1994.
- Coetzee GA, Strachan AF, van der Westhuyzen DR, Hoppe HC, Jeenah MS, and de Beer FC (1986) Serum amyloid A-containing human high density lipoprotein 3: density, size, and apolipoprotein composition. *J Biol Chem* **261**:9644–9651.
- Cohen JC, Vega GL, and Grundy SM (1999) Hepatic lipase: new insights from genetic and metabolic studies. *Curr Opin Lipidol* **10**:259–267.
- Colhoun HM, Otvos JD, Rubens MB, Taskinen MR, Underwood SR, and Fuller JH (2002) Lipoprotein subclasses and particle sizes and their relationship with coronary artery calcification in men and women with and without type 1 diabetes. *Diabetes* **51**:1949–1956.
- Connelly PW, Draganov D, and Maguire GF (2005) Paraoxonase-1 does not reduce or modify oxidation of phospholipids by peroxy-nitrite. *Free Radic Biol Med* **38**:164–174.
- Costa LG, Vitalone A, Cole TB, and Furlong CE (2005) Modulation of paraoxonase (PON1) activity. *Biochem Pharmacol* **69**:541–550.
- Crowl RM, Stoller TJ, Conroy RR, and Stoner CR (1991) Induction of phospholipase A2 gene expression in human hepatoma cells by mediators of the acute phase response. *J Biol Chem* **266**:2647–2651.
- Curb JD, Abbott RD, Rodriguez BL, Masaki K, Chen R, Sharp DS, and Tall AR (2004) A prospective study of HDL-C and cholesteryl ester transfer protein gene mutations and the risk of coronary heart disease in the elderly. *J Lipid Res* **45**:948–953.
- Curtiss LK, Bonnet DJ, and Rye KA (2000) The conformation of apolipoprotein A-I in high-density lipoproteins is influenced by core lipid composition and particle size: a surface plasmon resonance study. *Biochemistry* **39**:5712–5721.
- Cuzzocrea S, Dugo L, Patel NS, Di Paola R, Cockerill GW, Genovese T, and Thiemermann C (2004) High-density lipoproteins reduce the intestinal damage associated with ischemia/reperfusion and colitis. *Shock* **21**:342–351.
- Datta G, Epanand RF, Epanand RM, Chaddha M, Kirksey MA, Garber DW, Lund-Katz S, Phillips MC, Hama S, Navab M, et al. (2004) Aromatic residue position on the nonpolar face of class A amphipathic helical peptides determines biological activity. *J Biol Chem* **279**:26509–26517.
- Daum U, Leren TP, Langer C, Chirazi A, Cullen P, Pritchard PH, Assmann G, and von Eckardstein A (1999) Multiple dysfunctions of two apolipoprotein A-I variants, apoA-I(R160L)Oslo and apoA-I(P165R), that are associated with hypoalphalipoproteinemia in heterozygous carriers. *J Lipid Res* **40**:486–494.
- Davi G, Ciabattoni G, Consoli A, Mezzetti A, Falco A, Santarone S, Pennese E, Vitacolonna E, Bucciarelli T, Costantini F, et al. (1999) In vivo formation of 8-iso-prostaglandin F<sub>2α</sub> and platelet activation in diabetes mellitus: effects of improved metabolic control and vitamin E supplementation. *Circulation* **99**:224–229.
- Davignon J (2005) Apolipoprotein E and atherosclerosis: beyond lipid effect. *Arterioscler Thromb Vasc Biol* **25**:267–269.
- de Beer MC, Durbin DM, Cai L, Jonas A, de Beer FC, and van der Westhuyzen DR (2001) Apolipoprotein A-I conformation markedly influences HDL interaction with scavenger receptor BI. *J Lipid Res* **42**:309–313.
- de Grooth GJ, Klerkx AH, Stroes ES, Stalenhoef AF, Kastelein JJ, and Kuivenhoven JA (2004a) A review of CETP and its relationship to atherosclerosis. *J Lipid Res* **45**:1967–1974.
- de Grooth GJ, Kuivenhoven JA, Stalenhoef AF, de Graaf J, Zwinderman AH, Posma JL, van Tol A, and Kastelein JJ (2002) Efficacy and safety of a novel cholesteryl ester transfer protein inhibitor, JTT-705, in humans: a randomized phase II dose-response study. *Circulation* **105**:2159–2165.
- de Grooth GJ, Smilde TJ, van Wissen S, Klerkx AHM, Zwinderman AH, Fruchart J-C, Kastelein JJP, Stalenhoef AFH, and Kuivenhoven JA (2004b) The relationship between cholesteryl ester transfer protein levels and risk factor profile in patients with familial hypercholesterolemia. *Atherosclerosis* **173**:261–267.
- de la Llera-Moya M, Rothblat GH, Connelly MA, Kellner-Weibel G, Sakr SW, Phillips MC, and Williams DL (1999) Scavenger receptor BI (SR-BI) mediates free cholesterol flux independently of HDL tethering to the cell surface. *J Lipid Res* **40**:575–580.
- Deakin S, Moren X, and James RW (2005) Very low density lipoproteins provide a vector for secretion of paraoxonase-1 from cells. *Atherosclerosis* **179**:17–25.
- Deguchi H, Fernandez JA, Hackeng TM, Banka CL, and Griffin JH (2000) Cardio-lipin is a normal component of human plasma lipoproteins. *Proc Natl Acad Sci USA* **97**:1743–1748.
- Delange JR, Langlois MR, De Bacquer D, Mak R, Capel P, Van Renterghem L, and De Backer G (2002) Discriminative value of serum amyloid A and other acute-phase proteins for coronary heart disease. *Atherosclerosis* **160**:471–476.
- Desai MY, Rodriguez A, Wasserman BA, Gerstenblith G, Agarwal S, Kennedy M, Blumenk DA, and Lima JAC (2005) Association of cholesterol subfractions and carotid lipid core measured by MRI. *Arterioscler Thromb Vasc Biol* **25**:e110–e111.
- Devaraj S, Hirany SV, Burk RF, and Jialal I (2001) Divergence between LDL oxidative susceptibility and urinary F<sub>2</sub>-isoprostanes as measures of oxidative stress in type 2 diabetes. *Clin Chem* **47**:1974–1979.
- Doggen CJ, Smith NL, Lemaitre RN, Heckbert SR, Rosendaal FR, and Psaty BM (2004) Serum lipid levels and the risk of venous thrombosis. *Arterioscler Thromb Vasc Biol* **24**:1970–1975.
- Draganov DI, Teiber JF, Speelman A, Osawa Y, Sunahara R, and La Du BN (2005) Human paraoxonases (PON1, PON2, and PON3) are lactonases with overlapping and distinct substrate specificities. *J Lipid Res* **46**:1239–1247.
- Drew BG, Fidge NH, Gallon-Beaumier G, Kemp BE, and Kingwell BA (2004) High-density lipoprotein and apolipoprotein AI increase endothelial NO synthase activity by protein association and multisite phosphorylation. *Proc Natl Acad Sci USA* **101**:6999–7004.
- Drexel H, Amann FW, Beran J, Rentsch K, Candinas R, Muntwyler J, Luethy A, Gasser T, and Follath F (1994) Plasma triglycerides and three lipoprotein cholesterol fractions are independent predictors of the extent of coronary atherosclerosis. *Circulation* **90**:2230–2235.
- Drexel H, Amann FW, Rentsch K, Neuenschwander C, Luethy A, Khan SI, and Follath F (1992) Relation of the level of high-density lipoprotein subfractions to the presence and extent of coronary artery disease. *Am J Cardiol* **70**:436–440.
- Drexel H, Steurer J, Muntwyler J, Meienberg S, Schmid HR, Schneider E, Grochenig E, and Amann FW (1996) Predictors of the presence and extent of peripheral arterial occlusive disease. *Circulation* **94**:II199–II205.
- Duell PB and Malinow MR (1997) Homocyst(e)ine: an important risk factor for atherosclerotic vascular disease. *Curr Opin Lipidol* **8**:28–34.
- Duriez P and Fruchart JC (1999) High-density lipoprotein subclasses and apolipoprotein A-I. *Clin Chim Acta* **286**:97–114.
- Duverger N, Kruth H, Emmanuel F, Caillaud JM, Vigiotta C, Castro G, Tailleux A, Fievet C, Fruchart JC, Houdebine LM, et al. (1996a) Inhibition of atherosclerosis development in cholesterol-fed human apolipoprotein A-I-transgenic rabbits. *Circulation* **94**:713–717.
- Duverger N, Vigiotta C, Berthou L, Emmanuel F, Tailleux A, Parmentier-Nihoul L, Laine B, Fievet C, Castro G, Fruchart JC, et al. (1996b) Transgenic rabbits expressing human apolipoprotein A-I in the liver. *Arterioscler Thromb Vasc Biol* **16**:1424–1429.
- Ehrenwald E, Chisolm GM, and Fox PL (1994) Intact human ceruloplasmin oxidatively modifies low density lipoprotein. *J Clin Invest* **93**:1493–1501.
- Ehrenwald E and Fox PL (1996) Role of endogenous ceruloplasmin in low density lipoprotein oxidation by human U937 monocytic cells. *J Clin Invest* **97**:884–890.
- Epanand RM, Epanand RF, Sayer BG, Datta G, Chaddha M, and Anantharamaiah GM (2004) Two homologous apolipoprotein AI mimetic peptides: relationship between membrane interactions and biological activity. *J Biol Chem* **279**:51404–51414.
- Esteve E, Ricart W, and Fernandez-Real JM (2005) Dyslipidemia and inflammation: an evolutionary conserved mechanism. *Clin Nutr* **24**:16–31.
- Ettinger WH, Miller LD, Albers JF, Smith TK, and Parks JS (1990) Lipopolysaccharide and tumor necrosis factor cause a fall in plasma concentration of lecithin: cholesterol acyltransferase in cynomolgus monkeys. *J Lipid Res* **31**:1099–1107.
- Feingold KR, Stapanian I, Memon RA, Moser AH, Shigenaga JK, Doerfler W, Dinarello CA, and Grunfeld C (1992) Endotoxin rapidly induces changes in lipid metabolism that produce hypertriglyceridemia: low doses stimulate hepatic triglyceride production while high doses inhibit clearance. *J Lipid Res* **33**:1765–1776.
- Ferretti G, Bacchetti T, Busni D, Rabini RA, and Curatola G (2004) Protective effect of paraoxonase activity in high-density lipoproteins against erythrocyte membranes peroxidation: a comparison between healthy subjects and type 1 diabetic patients. *J Clin Endocrinol Metab* **89**:2957–2962.
- Ferretti G, Bacchetti T, Marchionni C, Caldarelli L, and Curatola G (2001) Effect of glycation of high density lipoproteins on their physicochemical properties and on paraoxonase activity. *Acta Diabetol* **38**:163–169.
- Ferretti G, Bacchetti T, Moroni C, Vignini A, and Curatola G (2003) Copper-induced oxidative damage on astrocytes: protective effect exerted by human high density lipoproteins. *Biochim Biophys Acta* **1635**:48–54.
- Ferretti G, Bacchetti T, Negre-Salvayre A, Salvayre R, Dousset N, and Curatola G (2005) Structural modifications of HDL and functional consequences. *Atherosclerosis* **184**:1–7.
- Festa A, Williams K, Hanley AJG, Otvos JD, Goff DC, Wagenknecht LE, and Haffner SM (2005) Nuclear magnetic resonance lipoprotein abnormalities in prediabetic subjects in the Insulin Resistance Atherosclerosis Study. *Circulation* **111**:3465–3472.
- FIELD Study Investigators (2004) The need for a large-scale trial of fibrate therapy in diabetes: the rationale and design of the Fenofibrate Intervention and Event

- Lowering in Diabetes (FIELD) study. [ISRCTN64783481]. *Cardiovasc Diabetol* **3**:9.
- Fievat C, Igau B, Bresson R, Drouhin P, and Fruchart JC (1995) Non-enzymatic glycosylation of apolipoprotein A-I and its functional consequences. *Diabetes Metab* **21**:95–98.
- Finch CE and Crimmins EM (2004) Inflammatory exposure and historical changes in human life-spans. *Science (Wash DC)* **305**:1736–1739.
- Fournier N, Atger V, Cogny A, Védie B, Giral P, Simon A, Moatti N, and Paul JL (2001) Analysis of the relationship between triglyceridemia and HDL-phospholipid concentrations: consequences on the efflux capacity of serum in the Fu5AH system. *Atherosclerosis* **157**:315–323.
- Franceschini G (2001) Epidemiologic evidence for high-density lipoprotein cholesterol as a risk factor for coronary artery disease. *Am J Cardiol* **88**:9N–13N.
- Franceschini G, Calabresi L, Chiesa G, Parolini C, Sirtori CR, Canavesi M, and Bernini F (1999) Increased cholesterol efflux potential of sera from ApoA-IMilano carriers and transgenic mice. *Arterioscler Thromb Vasc Biol* **19**:1257–1262.
- Freedman DS, Otvos JD, Jeyarajah EJ, Barborkiak JJ, Anderson AJ, and Walker JA (1998) Relation of lipoprotein subclasses as measured by proton nuclear magnetic resonance spectroscopy to coronary artery disease. *Arterioscler Thromb Vasc Biol* **18**:1046–1053.
- Frenais R, Ougueram K, Maugeais C, Marchini JS, Benlian P, Bard JM, Magot T, and Krempf M (1999) Apolipoprotein A-I kinetics in heterozygous familial hypercholesterolemia: a stable isotope study. *J Lipid Res* **40**:1506–1511.
- Furnkranz A, Schober A, Bochkov VN, Bashtrykov P, Kronke G, Kadl A, Binder BR, Weber C, and Leitinger N (2005) Oxidized phospholipids trigger atherogenic inflammation in murine arteries. *Arterioscler Thromb Vasc Biol* **25**:633–638.
- Fyfe AI, Rothenberg LS, DeBeer FC, Cantor RM, Rotter JI, and Lusis AJ (1997) Association between serum amyloid A proteins and coronary artery disease: evidence from two distinct arteriosclerotic processes. *Circulation* **96**:2914–2919.
- Gaidukov L and Tawfik DS (2005) High affinity, stability, and lactonase activity of serum paraoxonase PON1 anchored on HDL with ApoA-I. *Biochemistry* **44**:11843–11854.
- Ganji SH, Kamanna VS, and Kashyap ML (2003) Niacin and cholesterol: role in cardiovascular disease (Review). *J Nutr Biochem* **14**:298–305.
- Gaofu Q, Jun L, Xiuyun Z, Wentao L, Jie W, and Jingjing L (2005) Antibody against cholesteryl ester transfer protein (CETP) elicited by a recombinant chimeric enzyme vaccine attenuated atherosclerosis in a rabbit model. *Life Sci* **77**:2690–2702.
- Garber DW, Datta G, Chaddha M, Palgunachari MN, Hama SY, Navab M, Fogelman AM, Segrest JP, and Anantharamaiah GM (2001) A new synthetic class A amphipathic peptide analogue protects mice from diet-induced atherosclerosis. *J Lipid Res* **42**:545–552.
- Garner B, Waldeck AR, Witting PK, Rye KA, and Stocker R (1998) Oxidation of high density lipoproteins. II. Evidence for direct reduction of lipid hydroperoxides by methionine residues of apolipoproteins AI and AII. *J Biol Chem* **273**:6088–6095.
- Garvey WT, Kwon S, Zheng D, Shaughnessy S, Wallace P, Hutto A, Pugh K, Jenkins AJ, Klein RL, and Liao Y (2003) Effects of insulin resistance and type 2 diabetes on lipoprotein subclass particle size and concentration determined by nuclear magnetic resonance. *Diabetes* **52**:453–462.
- Gaus K, Dean RT, Kritharides L, and Jessup W (2001) Inhibition of cholesterol efflux by 7-ketocholesterol: comparison between cells, plasma membrane vesicles, and liposomes as cholesterol donors. *Biochemistry* **40**:13002–13014.
- Gelissen IC, Rye KA, Brown AJ, Dean RT, and Jessup W (1999) Oxysterol efflux from macrophage foam cells: the essential role of acceptor phospholipid. *J Lipid Res* **40**:1636–1646.
- Goff DC Jr, D'Agostino RB Jr, Haffner SM, and Otvos JD (2005) Insulin resistance and adiposity influence lipoprotein size and subclass concentrations: results from the Insulin Resistance Atherosclerosis Study. *Metabolism* **54**:264–270.
- Gomaraschi M, Basilico N, Sisto F, Taramelli D, Eligini S, Colli S, Sirtori CR, Franceschini G, and Calabresi L (2005) High-density lipoproteins attenuate interleukin-6 production in endothelial cells exposed to pro-inflammatory stimuli. *Biochim Biophys Acta* **1736**:136–143.
- Gotto AM Jr and Brinton EA (2004) Assessing low levels of high-density lipoprotein cholesterol as a risk factor in coronary heart disease: a working group report and update. *J Am Coll Cardiol* **43**:717–724.
- Goulinet S and Chapman MJ (1997) Plasma LDL and HDL subspecies are heterogeneous in particle content of tocopherols and oxygenated and hydrocarbon carotenoids: relevance to oxidative resistance and atherogenesis. *Arterioscler Thromb Vasc Biol* **17**:786–796.
- Gowri MS, Van der Westhuizen DR, Bridges SR, and Anderson JW (1999) Decreased protection by HDL from poorly controlled type 2 diabetic subjects against LDL oxidation may be due to the abnormal composition of HDL. *Arterioscler Thromb Vasc Biol* **19**:2226–2233.
- Greene DJ, Skeggs JW, and Morton RE (2001) Elevated triglyceride content diminishes the capacity of high density lipoprotein to deliver cholesteryl esters via the scavenger receptor class B type I (SR-BI). *J Biol Chem* **276**:4804–4811.
- Griffin JH, Kojima K, Banka CL, Curtiss LK, and Fernandez JA (1999) High-density lipoprotein enhancement of anticoagulant activities of plasma protein S and activated protein C. *J Clin Invest* **103**:219–227.
- Guerin M, Dolphin PJ, Talusot C, Gardette J, Berthezene F, and Chapman MJ (1995) Pravastatin modulates cholesteryl ester transfer from HDL to apoB-containing lipoproteins and lipoprotein subspecies profile in familial hypercholesterolemia. *Arterioscler Thromb Vasc Biol* **15**:1359–1368.
- Guerin M, Lassel TS, Le Goff W, Farnier M, and Chapman MJ (2000a) Action of atorvastatin in combined hyperlipidemia: preferential reduction of cholesteryl ester transfer from HDL to VLDL1 particles. *Arterioscler Thromb Vasc Biol* **20**:189–197.
- Guerin M, Lassel TS, Le Goff W, Farnier M, and Chapman MJ (2000b) Action of atorvastatin in combined hyperlipidemia: preferential reduction of cholesteryl ester transfer from HDL to VLDL1 particles. *Arterioscler Thromb Vasc Biol* **20**:189–197.
- Gupta H, Dai L, Datta G, Garber DW, Grenett H, Li Y, Mishra V, Palgunachari MN, Handattu S, Gianturco SH, et al. (2005a) Inhibition of lipopolysaccharide-induced inflammatory responses by an apolipoprotein AI mimetic peptide. *Circ Res* **97**:236–243.
- Gupta H, White CR, Handattu S, Garber DW, Datta G, Chaddha M, Dai L, Gianturco SH, Bradley WA, and Anantharamaiah GM (2005b) Apolipoprotein E mimetic peptide dramatically lowers plasma cholesterol and restores endothelial function in Watanabe heritable hyperlipidemic rabbits. *Circulation* **111**:3112–3118.
- Hajduk SL, Moore DR, Vasudevacharya J, Siqueira H, Torri AF, Tytler EM, and Esko JD (1989) Lysis of *Trypanosoma brucei* by a toxic subspecies of human high density lipoprotein. *J Biol Chem* **264**:5210–5217.
- Hamon Y, Broccardo C, Chambenoit O, Luciani MF, Toti F, Chaslin S, Freyssinet JM, Devaux PF, McNeish J, Marguet D, et al. (2000) ABC1 promotes engulfment of apoptotic cells and transbilayer redistribution of phosphatidylserine. *Nat Cell Biol* **2**:399–406.
- Hamon Y, Luciani MF, Becq F, Verrier B, Rubartelli A, and Chimini G (1997) Interleukin-1 $\beta$  secretion is impaired by inhibitors of the ATP binding cassette transporter. *ABC1. Blood* **90**:2911–2915.
- Han H, Sasaki J, Matsunaga A, Hakamata H, Huang W, Ageta M, Taguchi T, Koga T, Kugi M, Horiuchi S, et al. (1999) A novel mutant, ApoA-I nichinan (Glu235 $\rightarrow$ O), is associated with low HDL cholesterol levels and decreased cholesterol efflux from cells. *Arterioscler Thromb Vasc Biol* **19**:1447–1455.
- Hansel B, Giral P, Nobecourt E, Chantepie S, Bruckert E, Chapman MJ, and Kontush A (2004) Metabolic syndrome is associated with elevated oxidative stress and dysfunctional dense high-density lipoprotein particles displaying impaired antioxidative activity. *J Clin Endocrinol Metab* **89**:4963–4971.
- Hansson GK (2005) Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* **352**:1685–1695.
- Hara S, Shike T, Takasu N, and Mizui T (1997) Lysophosphatidylcholine promotes cholesterol efflux from mouse macrophage foam cells. *Arterioscler Thromb Vasc Biol* **17**:1258–1266.
- Harangi M, Seres I, Varga Z, Emri G, Szilvassy Z, Paragh G, and Remenyik E (2004) Atorvastatin effect on high-density lipoprotein-associated paraoxonase activity and oxidative DNA damage. *Eur J Clin Pharmacol* **60**:685–691.
- Harel M, Aharoni A, Gaidukov L, Brumshtein B, Khersonsky O, Meged R, Dvir H, Ravelli RB, McCarthy A, Toker L, et al. (2004) Structure and evolution of the serum paraoxonase family of detoxifying and anti-atherosclerotic enzymes. *Nat Struct Mol Biol* **11**:412–419.
- Hasselwander O, McEneny J, McMaster D, Fogarty DG, Nicholls DP, Maxwell AP, and Young IS (1999) HDL composition and HDL antioxidant capacity in patients on regular haemodialysis. *Atherosclerosis* **143**:125–133.
- Hedrick CC, Thorpe SR, Fu MX, Harper CM, Yoo J, Kim SM, Wong H, and Peters AL (2000) Glycation impairs high-density lipoprotein function. *Diabetologia* **43**:312–320.
- Heeren J, Weber W, and Beisiegel U (1999) Intracellular processing of endocytosed triglyceride-rich lipoproteins comprises both recycling and degradation. *J Cell Sci* **112**:349–359.
- Heinecke JW (1998) Oxidants and antioxidants in the pathogenesis of atherosclerosis: implications for the oxidized low density lipoprotein hypothesis. *Atherosclerosis* **141**:1–15.
- Herrington DM and Parks JS (2004) Estrogen and HDL: all that glitters is not gold. *Arterioscler Thromb Vasc Biol* **24**:1741–1742.
- Hoffman JS and Benditt EP (1982a) Changes in high density lipoprotein content following endotoxin administration in the mouse: formation of serum amyloid protein-rich subfractions. *J Biol Chem* **257**:10510–10517.
- Hoffman JS and Benditt EP (1982b) Secretion of serum amyloid protein and assembly of serum amyloid protein-rich high density lipoprotein in primary mouse hepatocyte culture. *J Biol Chem* **257**:10518–10522.
- Hoffman JS and Benditt EP (1983) Plasma clearance kinetics of the amyloid-related high density lipoprotein apoprotein, serum amyloid protein (apoSAA), in the mouse: evidence for rapid apoSAA clearance. *J Clin Invest* **71**:926–934.
- Holvoet P, Harris TB, Tracy RP, Verhamme P, Newman AB, Rubin SM, Simonsick EM, Colbert LH and Kritchevsky SB (2003) Association of high coronary heart disease risk status with circulating oxidized LDL in the well-functioning elderly: findings from the Health, Aging, and Body Composition study. *Arterioscler Thromb Vasc Biol* **23**:1444–1448.
- Hovingh GK, Hutten BA, Holleboom AG, Petersen W, Rol P, Stalenhoef A, Zwinderman AH, de Groot E, Kastelein JJ, and Kuivenhoven JA (2005) Compromised LCAT function is associated with increased atherosclerosis. *Circulation* **112**: 879–884.
- Huang JM, Huang ZX, and Zhu W (1998) Mechanism of high-density lipoprotein subfractions inhibiting copper-catalyzed oxidation of low-density lipoprotein. *Clin Biochem* **31**:537–543.
- Huang W, Sasaki J, Matsunaga A, Han H, Li W, Koga T, Kugi M, Ando S, and Arakawa K (2000) A single amino acid deletion in the carboxy terminal of apolipoprotein A-I impairs lipid binding and cellular interaction. *Arterioscler Thromb Vasc Biol* **20**:210–216.
- Ikwaki K, Noma K, Tohyama J, Kido T, and Mochizuki S (2005) Effects of bezafibrate on lipoprotein subclasses and inflammatory markers in patients with hypertriglyceridemia - a nuclear magnetic resonance study. *Int J Cardiol* **101**:441–447.
- Ikwaki K, Tohyama J, Nakata Y, Wakikawa T, Kido T, and Mochizuki S (2004) Fenofibrate effectively reduces remnants, and small dense LDL, and increases HDL particle number in hypertriglyceridemic men—a nuclear magnetic resonance study. *J Arterioscler Thromb* **11**:278–285.
- Ishida BY, Frolich J, and Fielding CJ (1987) Pre $\beta$ -migrating high density lipoprotein: quantitation in normal and hyperlipidemic plasma by solid phase radioimmunoassay following electrophoretic transfer. *J Lipid Res* **28**:778–786.
- Ishigami M, Yamashita S, Sakai N, Arai T, Hirano K, Hiraoka H, Kameda-Takemura K, and Matsuzawa Y (1994) Large and cholesteryl ester-rich high-density lipoproteins in cholesteryl ester transfer protein (CETP) deficiency cannot

- protect macrophages from cholesterol accumulation induced by acetylated low-density lipoproteins. *J Biochem (Tokyo)* **116**:257–262.
- Jakubowski H (2000) Calcium-dependent human serum homocysteine thiolactone hydrolase: a protective mechanism against protein N-homocysteinylolation. *J Biol Chem* **275**:3957–3962.
- James RW and Deakin SP (2004) The importance of high-density lipoproteins for paraoxonase-1 secretion, stability, and activity. *Free Radic Biol Med* **37**:1986–1994.
- Jaouad L, de Guise C, Berrougui H, Cloutier M, Isabelle M, Fulop T, Payette H, and Khalil A (2005) Age-related decrease in high-density lipoproteins antioxidant activity is due to an alteration in the PON1's free sulfhydryl groups. *Atherosclerosis* **185**:195–200.
- Jaouad L, Milochevitch C, and Khalil A (2003) PON1 paraoxonase activity is reduced during HDL oxidation and is an indicator of HDL antioxidant capacity. *Free Radic Res* **37**:77–83.
- Johansson J, Carlsson LA, Landou C, and Hamsten A (1991) High density lipoproteins and coronary atherosclerosis: a strong inverse relation with the largest particles is confined to normotriglyceridemic patients. *Arterioscler Thromb* **11**:174–182.
- Johnson BD, Kip KE, Marroquin OC, Ridker PM, Kelsey SF, Shaw LJ, Pepine CJ, Sharaf B, Bairey Merz CN, Sopko G, et al. (2004) Serum amyloid A as a predictor of coronary artery disease and cardiovascular outcome in women: the National Heart, Lung, and Blood Institute-Sponsored Women's Ischemia Syndrome Evaluation (WISE). *Circulation* **109**:726–732.
- Jonas A (2000) Lecithin cholesterol acyltransferase. *Biochim Biophys Acta* **1529**:245–256.
- Jordan-Starck TC, Witte DP, Aronow BJ, and Harmony JA (1992) Apolipoprotein J: a membrane policeman? *Curr Opin Lipidol* **1992**:75–85, 1992.
- Josse D, Ebel C, Stroebel D, Fontaine A, Borges F, Echaliar A, Baud D, Renault F, Le Maire M, Chabrieres E, et al. (2002) Oligomeric states of the detergent-solubilized human serum paraoxonase (PON1). *J Biol Chem* **277**:33386–33397.
- Kahri J, Vuorinen-Markkola H, Tilly-Kiesi M, Lahdenpera S, and Taskinen MR (1993) Effect of gemfibrozil on high density lipoprotein subspecies in non-insulin dependent diabetes mellitus: relations to lipolytic enzymes and to the cholesteryl ester transfer protein activity. *Atherosclerosis* **102**:79–89.
- Karabina SA, Elisaf M, Bairaktari E, Tzallas C, Siamopoulos KC, and Tselepis AD (1997) Increased activity of platelet-activating factor acetylhydrolase in low-density lipoprotein subfractions induces enhanced lysophosphatidylcholine production during oxidation in patients with heterozygous familial hypercholesterolemia. *Eur J Clin Invest* **27**:595–602.
- Karlsson H, Leanderson P, Tagesson C, and Lindahl M (2005) Lipoproteomics II: mapping of proteins in high-density lipoprotein using two-dimensional gel electrophoresis and mass spectrometry. *Proteomics* **5**:1431–1445.
- Karpe F and Frayn KN (2004) The nicotinic acid receptor—a new mechanism for an old drug. *Lancet* **363**:1892–1894.
- Kazama H, Usui S, Okazaki M, Hosoi T, Ito H, and Orimo H (2003) Effects of bezafibrate and pravastatin on remnant-like lipoprotein particles and lipoprotein subclasses in type 2 diabetes. *Diabetes Res Clin Pract* **59**:181–189.
- Keidar S, Ostlund RE Jr, and Schonfeld G (1990) Apolipoprotein E-rich HDL in patients with homozygous familial hypercholesterolemia. *Atherosclerosis* **84**:155–163.
- Kelso GJ, Stuart WD, Richter RJ, Furlong CE, Jordan-Starck TC, and Harmony JA (1994) Apolipoprotein J is associated with paraoxonase in human plasma. *Biochemistry* **33**:832–839.
- Kennedy MA, Barrera GC, Nakamura K, Baldan A, Tarr P, Fishbein MC, Frank J, Francone OL, and Edwards PA (2005) ABCG1 has a critical role in mediating cholesterol efflux to HDL and preventing cellular lipid accumulation. *Cell Metab* **1**:121–131.
- Kent JW Jr, Comuzzie AG, Mahaney MC, Almsy L, Rainwater DL, VandeBerg JL, MacCluer JW, and Blangero J (2004) Intercellular adhesion molecule-1 concentration is genetically correlated with insulin resistance, obesity, and HDL concentration in Mexican Americans. *Diabetes* **53**:2691–2695.
- Khalil MF, Wagner WD, and Goldberg IJ (2004) Molecular interactions leading to lipoprotein retention and the initiation of atherosclerosis. *Arterioscler Thromb Vasc Biol* **24**:2211–2218.
- Kherkeulidze P, Johansson J, and Carlson LA (1991) High density lipoprotein particle size distribution in cord blood. *Acta Paediatr Scand* **80**:770–779.
- Khersonsky O and Tawfik DS (2005) Structure-reactivity studies of serum paraoxonase PON1 suggest that its native activity is lactonase. *Biochemistry* **44**:6371–6382.
- Khovidhunkit W, Duchateau PN, Medzhradszky KF, Moser AH, Naya-Vigne J, Shigenaga JK, Kane JP, Grunfeld C, and Feingold KR (2004a) Apolipoproteins A-IV and A-V are acute-phase proteins in mouse HDL. *Atherosclerosis* **176**:37–44.
- Khovidhunkit W, Kim MS, Memon RA, Shigenaga JK, Moser AH, Feingold KR, and Grunfeld C (2004b) Effects of infection and inflammation on lipid and lipoprotein metabolism: mechanisms and consequences to the host. *J Lipid Res* **45**:1169–1196.
- Khovidhunkit W, Shigenaga JK, Moser AH, Feingold KR, and Grunfeld C (2001) Cholesterol efflux by acute-phase high density lipoprotein: role of lecithin: cholesterol acyltransferase. *J Lipid Res* **42**:967–975.
- Kimura T, Sato K, Malchinkhuu E, Tomura H, Tamama K, Kuwabara A, Murakami M, and Okajima F (2003) High-density lipoprotein stimulates endothelial cell migration and survival through sphingosine 1-phosphate and its receptors. *Arterioscler Thromb Vasc Biol* **23**:1283–1288.
- Kinoshita M, Fujita M, Usui S, Maeda Y, Kudo M, Hirota D, Suda T, Taki M, Okazaki M, and Teramoto T (2004) Scavenger receptor type BI potentiates reverse cholesterol transport system by removing cholesterol ester from HDL. *Atherosclerosis* **173**:197–202.
- Kisilevsky R and Subrahmanyam L (1992) Serum amyloid A changes high density lipoprotein's cellular affinity: a clue to serum amyloid A's principal function. *Lab Invest* **66**:778–785.
- Klimov AN, Gurevich VS, Nikiforova AA, Shtatilina LV, Kuzmin AA, Plavinsky SL, and Teryukova NP (1993) Antioxidative activity of high density lipoproteins in vivo. *Atherosclerosis* **100**:13–18.
- Klimov AN, Kozhevnikova KA, Kuznetsov AS, and Belova EV (2001) On the ability of high density lipoproteins to remove phospholipid peroxidation products from erythrocyte membranes. *Biochemistry (Mosc)* **66**:300–304.
- Kobayashi J, Okamoto H, Otobe M, Bujo H, and Saito Y (2002) Effect of HDL, from Japanese white rabbit administered a new cholesteryl ester transfer protein inhibitor JTT-705, on cholesteryl ester accumulation induced by acetylated low density lipoprotein in J774 macrophage. *Atherosclerosis* **162**:131–135.
- Koizumi J, Inazu A, Yagi K, Koizumi I, Uno Y, Kajinami K, Miyamoto S, Moulin P, Tall AR, Mabuchi H, et al. (1991) Serum lipoprotein lipid concentration and composition in homozygous and heterozygous patients with cholesteryl ester transfer protein deficiency. *Atherosclerosis* **90**:189–196.
- Kontush A (2004) Apolipoprotein A $\beta$ : black sheep in a good family. *Brain Pathol* **14**:433–447.
- Kontush A, Chantepie S, and Chapman MJ (2003) Small, dense HDL particles exert potent protection of atherogenic LDL against oxidative stress. *Arterioscler Thromb Vasc Biol* **23**:1881–1888.
- Kontush A, de Faria EC, Chantepie S, and Chapman MJ (2004) Antioxidative activity of HDL particle subspecies is impaired in hyperalphalipoproteinemia: relevance of enzymatic and physicochemical properties. *Arterioscler Thromb Vasc Biol* **24**:526–533.
- Kontush A, de Faria EC, Chantepie S, and Chapman MJ (2005) A normotriglyceridemic, low HDL-cholesterol phenotype is characterised by elevated oxidative stress and HDL particles with attenuated antioxidative activity. *Atherosclerosis* **182**:277–285.
- Kris-Etherton PM, Lichtenstein AH, Howard BV, Steinberg D, Witztum JL, for the Nutrition Committee of the American Heart Association Council on Nutrition, Physical Activity, and Metabolism (2004) Antioxidant vitamin supplements and cardiovascular disease. *Circulation* **110**:637–641.
- Kruger AL, Peterson S, Turkseven S, Kaminski PM, Zhang FF, Quan S, Wolin MS, and Abraham NG (2005) D-4F induces heme oxygenase-1 and extracellular superoxide dismutase, decreases endothelial cell sloughing, and improves vascular reactivity in rat model of diabetes. *Circulation* **111**:3126–3134.
- Kuivenhoven JA, de Grooth GJ, Kawamura H, Klerkx AH, Wilhelm F, Trip MD, and Kastelein JJP (2005) Effectiveness of inhibition of cholesteryl ester transfer protein by JTT-705 in combination with pravastatin in type II dyslipidemia. *Am J Cardiol* **95**:1085–1088.
- Kuivenhoven JA, Pritchard H, Hill J, Frohlich J, Assmann G, and Kastelein J (1997) The molecular pathology of lecithin:cholesterol acyltransferase (LCAT) deficiency syndromes. *J Lipid Res* **38**:191–205.
- Kujiraoka T, Hattori H, Ito M, Nanjee MN, Ishihara M, Nagano M, Iwasaki T, Cooke CJ, Olszewski WL, and Stepanova IP (2004) Effects of intravenous apolipoprotein A-I-phosphatidylcholine discs on paraoxonase and platelet-activating factor acetylhydrolase in human plasma and tissue fluid. *Atherosclerosis* **176**:57–62.
- Kunitake ST, Jarvis MR, Hamilton RL, and Kane JP (1992) Binding of transition metals by apolipoprotein A-I-containing plasma lipoproteins: inhibition of oxidation of low density lipoproteins. *Proc Natl Acad Sci USA* **89**:6993–6997.
- Kwon YG, Min JK, Kim KM, Lee DJ, Billiar TR, and Kim YM (2001) Sphingosine 1-phosphate protects human umbilical vein endothelial cells from serum-deprived apoptosis by nitric oxide production. *J Biol Chem* **276**:10627–10633.
- Lamarche B, Rashid S, and Lewis GF (1999) HDL metabolism in hypertriglyceridemic states: an overview. *Clin Chim Acta* **286**:145–161, 1999.
- Langmann T, Klucken J, Reil M, Liebisch G, Luciani MF, Chimini G, Kaminski WE, and Schmitz G (1999) Molecular cloning of the human ATP-binding cassette transporter 1 (hABC1): evidence for sterol-dependent regulation in macrophages. *Biochem Biophys Res Commun* **257**:29–33.
- Le Goff W, Guerin M, and Chapman MJ (2004) Pharmacological modulation of cholesteryl ester transfer protein, a new therapeutic target in atherogenic dyslipidemia. *Pharmacol Ther* **101**:17–38.
- Lee CM, Chien CT, Chang PY, Hsieh MY, Jui HY, Liau CS, Hsu SM, and Lee YT (2005) High-density lipoprotein antagonizes oxidized low-density lipoprotein by suppressing oxygen free-radical formation and preserving nitric oxide bioactivity. *Atherosclerosis* **183**:251–258.
- Lesnik P, Vonica A, Guerin M, Moreau M, and Chapman MJ (1993) Anticoagulant activity of tissue factor pathway inhibitor in human plasma is preferentially associated with dense subspecies of LDL and HDL and with Lp(a). *Arterioscler Thromb* **13**:1066–1075.
- Levels JH, Abraham PR, van den Ende A, and van Deventer SJ (2001) Distribution and kinetics of lipoprotein-bound endotoxin. *Infect Immun* **69**:2821–2828.
- Lewis GF and Rader DJ (2005) New insights into the regulation of HDL metabolism and reverse cholesterol transport. *Circ Res* **96**:1221–1232.
- Lewis KE, Kirk EA, McDonald TO, Wang S, Wight TN, O'Brien KD, and Chait A (2004) Increase in serum amyloid A evoked by dietary cholesterol is associated with increased atherosclerosis in mice. *Circulation* **110**:540–545.
- Li X, Chyu K-Y, Neto JRF, Yano J, Nathwani N, Ferreira C, Dimayuga PC, Cercek B, Kaul S, and Shah PK (2004) Differential effects of apolipoprotein A-I-mimetic peptide on evolving and established atherosclerosis in apolipoprotein E-null mice. *Circulation* **110**:1701–1705.
- Li XA, Titlov WB, Jackson BA, Giltiay N, Nikolova-Karakashian M, Uittenbogaard A, and Smart EJ (2002) High density lipoprotein binding to scavenger receptor, class B, type 1 activates endothelial nitric-oxide synthase in a ceramide-dependent manner. *J Biol Chem* **277**:11058–11063.
- Liang JS, Schreiber BM, Salmons M, Phillip G, Gonnerman WA, de Beer FC, and Sipe JD (1996) Amino terminal region of acute phase, but not constitutive, serum amyloid A (apoSAA) specifically binds and transports cholesterol into aortic smooth muscle and HepG2 cells. *J Lipid Res* **37**:2109–2116.
- Liao XL, Lou B, Ma J, and Wu MP (2005) Neutrophils activation can be diminished by apolipoprotein A-I. *Life Sci* **77**:325–335.
- Libby P (2002) Inflammation in atherosclerosis. *Nature (Lond)* **420**:868–874.
- Liuzzo G, Biasucci LM, Gallimore JR, Grillo RL, Rebuzzi AG, Pepys MB, and Maseri

- A (1994) The prognostic value of C-reactive protein and serum amyloid A protein in severe unstable angina. *N Engl J Med* **331**:417–424.
- Lusis AJ (2000) Atherosclerosis. *Nature (Lond)* **407**:203–241, 2000.
- Ma J, Liao XL, Lou B, and Wu MP (2004) Role of apolipoprotein A-I in protecting against endotoxin toxicity. *Acta Biochim Biophys Sin (Shanghai)* **36**:419–424.
- Mackey RH, Kuller LH, Sutton-Tyrrell K, Evans RW, Holubkov R, and Matthews KA (2002) Lipoprotein subclasses and coronary artery calcium in postmenopausal women from the healthy women study. *Am J Cardiol* **90**:711–761.
- Mackness B, Durrington P, McElduff P, Yarnell J, Azam N, Watt M, and Mackness M (2003) Low paraoxonase activity predicts coronary events in the Caerphilly Prospective Study. *Circulation* **107**:2775–2779.
- Mackness MI, Harty D, Bhatnagar D, Winocour PH, Arrol S, Ishola M, and Durrington PN (1991) Serum paraoxonase activity in familial hypercholesterolaemia and insulin-dependent diabetes mellitus. *Atherosclerosis* **86**:193–199.
- Madamanchi NR, Vendrov A, and Runge MS (2005) Oxidative stress and vascular disease. *Arterioscler Thromb Vasc Biol* **25**:29–38.
- Maddipati KR and Marnett LJ (1987) Characterization of the major hydroperoxide-reducing activity of human plasma: purification and properties of a selenium-dependent glutathione peroxidase. *J Biol Chem* **262**:17398–17403.
- Maier W, Altwegg LA, Corti R, Gay S, Hersberger M, Maly FE, Sutsch G, Roffi M, Neidhart M, Eberli FR, et al. (2005) Inflammatory markers at the site of ruptured plaque in acute myocardial infarction: locally increased interleukin-6 and serum amyloid A but decreased C-reactive protein. *Circulation* **111**:1355–1361.
- Maiorano JN, Jandacek RJ, Horace EM, and Davidson WS (2004) Identification and structural ramifications of a hinge domain in apolipoprotein A-I discoidal high-density lipoproteins of different size. *Biochemistry* **43**:11717–11726.
- Malle E, Steinmetz A, and Raynes JG (1993) Serum amyloid A (SAA): an acute phase protein and apolipoprotein. *Atherosclerosis* **102**:131–146.
- Marathe GK, Zimmerman GA, and McIntyre TM (2003) Platelet-activating factor acetylhydrolase, and not paraoxonase-1, is the oxidized phospholipid hydrolase of high density lipoprotein particles. *J Biol Chem* **278**:3937–3947.
- Marchesi S, Lupattelli G, Lombardini R, Sensini A, Siepi D, Mannarino M, Vaudo G, and Mannarino E (2005) Acute inflammatory state during influenza infection and endothelial function. *Atherosclerosis* **178**:345–350.
- Marcil M, Yu L, Krimbou L, Boucher B, Oram JF, Cohn JS, and Genest J Jr (1999) Cellular cholesterol transport and efflux in fibroblasts are abnormal in subjects with familial HDL deficiency. *Arterioscler Thromb Vasc Biol* **19**:159–169.
- Marhaug G, Sletten K, and Husby G (1982) Characterization of amyloid related protein SAA complexed with serum lipoproteins (apoSAA). *Clin Exp Immunol* **50**:382–389.
- Maron DJ (2000) The epidemiology of low levels of high-density lipoprotein cholesterol in patients with and without coronary artery disease. *Am J Cardiol* **86**:11L–14L.
- Marsche G, Heller R, Fauler G, Kovacevic A, Nuszkowski A, Graier W, Sattler W, and Malle E (2004) 2-Chlorohexadecanal derived from hypochlorite-modified high-density lipoprotein-associated plasmalogen is a natural inhibitor of endothelial nitric oxide biosynthesis. *Arterioscler Thromb Vasc Biol* **24**:2302–2306.
- Mazzone T (1996) Apolipoprotein E secretion by macrophages: its potential physiological functions. *Curr Opin Lipidol* **7**:303–307.
- McKenney J (2004) New perspectives on the use of niacin in the treatment of lipid disorders. *Arch Intern Med* **164**:697–705.
- McKenney JM, McCormick LS, Schaefer EJ, Black DM, and Watkins ML (2001) Effect of niacin and atorvastatin on lipoprotein subclasses in patients with atherogenic dyslipidemia. *Am J Cardiol* **88**:270–274.
- Meisinger C, Baumert J, Khuseynova N, Loewel H, and Koenig W (2005) Plasma oxidized low-density lipoprotein, a strong predictor for acute coronary heart disease events in apparently healthy, middle-aged men from the general population. *Circulation* **112**:651–657.
- Memon RA, Staprans I, Noor M, Holleran WM, Uchida Y, Moser AH, Feingold KR, and Grunfeld C (2000) Infection and inflammation induce LDL oxidation in vivo. *Arterioscler Thromb Vasc Biol* **20**:1536–1542.
- Mertens A, Verhamme P, Bielicki JK, Phillips MC, Quarck R, Verreth W, Stengel D, Ninio E, Navab M, Mackness B, et al. (2003) Increased low-density lipoprotein oxidation and impaired high-density lipoprotein antioxidant defense are associated with increased macrophage homing and atherosclerosis in dyslipidemic obese mice: LCAT gene transfer decreases atherosclerosis. *Circulation* **107**:1640–1646.
- Meyers CD, Kamanna VS, and Kashyap ML (2004) Niacin therapy in atherosclerosis. *Curr Opin Lipidol* **15**:659–665.
- Miida T, Inano K, Yamaguchi T, Tsuda T, and Okada M (1997) LpA-I levels do not reflect pre $\beta$ 1-HDL levels in human plasma. *Atherosclerosis* **133**:221–226.
- Miida T, Sakai K, Ozaki K, Nakamura Y, Yamaguchi T, Tsuda T, Kashiwa T, Murakami T, Inano K, and Okada M (2000) Bezafibrate increases pre $\beta$ 1-HDL at the expense of HDL2b in hypertriglyceridemia. *Arterioscler Thromb Vasc Biol* **20**:2428–2433.
- Miller ER 3rd, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ and Guallar E (2005) Meta-analysis: high-dose vitamin E supplementation may increase all-cause mortality. *Ann Intern Med* **142**:37–46.
- Miyata M and Smith JD (1996) Apolipoprotein E allele-specific antioxidant activity and effects on cytotoxicity by oxidative insults and  $\beta$ -amyloid peptides. *Nat Genet* **14**:55–61.
- Moore RE, Kawashiri MA, Kitajima K, Secretò A, Millar JS, Pratico D, and Rader DJ (2003) Apolipoprotein A-I deficiency results in markedly increased atherosclerosis in mice lacking the LDL receptor. *Arterioscler Thromb Vasc Biol* **23**:1914–1920.
- Moore RE, Navab M, Millar JS, Zimetti F, Hama S, Rothblat GH, and Rader DJ (2005) Increased atherosclerosis in mice lacking apolipoprotein A-I attributable to both impaired reverse cholesterol transport and increased inflammation. *Circ Res* **97**:763–771.
- Morgan JM, Capuzzi DM, Baksh RI, Intenzo C, Carey CM, Reese D, and Walker K (2003) Effects of extended-release niacin on lipoprotein subclass distribution. *Am J Cardiol* **91**:1432–1436.
- Morrow JD (2005) Quantification of isoprostanes as indices of oxidant stress and the risk of atherosclerosis in humans. *Arterioscler Thromb Vasc Biol* **25**:279–286.
- Mowat BF, Skinner ER, Wilson HM, Leng GC, Fowkes FG, and Horrobin D (1997) Alterations in plasma lipids, lipoproteins and high density lipoprotein subfractions in peripheral arterial disease. *Atherosclerosis* **131**:161–166.
- Mueller CF, Laude K, McNally JS, and Harrison DG (2005) Redox mechanisms in blood vessels. *Arterioscler Thromb Vasc Biol* **25**:274–278.
- Nakamura K, Kennedy MA, Baldan A, Bojanic DD, Lyons K, and Edwards PA (2004) Expression and regulation of multiple murine ATP-binding cassette transporter G1 mRNAs/isoforms that stimulate cellular cholesterol efflux to high density lipoprotein. *J Biol Chem* **279**:45980–45989.
- Nanjee MN and Brinton EA (2000) Very small apolipoprotein A-I-containing particles from human plasma: isolation and quantification by high-performance size-exclusion chromatography. *Clin Chem* **46**:207–223.
- Nanjee MN, Cooke CJ, Wong JS, Hamilton RL, Olszewski WL, and Miller NE (2001) Composition and ultrastructure of size subclasses of normal human peripheral lymph lipoproteins: quantification of cholesterol uptake by HDL in tissue fluids. *J Lipid Res* **42**:639–648.
- Nanjee MN, Doran JE, Lerch PG, and Miller NE (1999) Acute effects of intravenous infusion of ApoA1/phosphatidylcholine discs on plasma lipoproteins in humans. *Arterioscler Thromb Vasc Biol* **19**:979–989.
- Napoli C, Pignalosa O, de Nigris F, and Sica V (2005) Childhood infection and endothelial dysfunction: a potential link in atherosclerosis? *Circulation* **111**:1568–1570.
- Navab M, Anantharamaiah GM, Hama S, Garber DW, Chaddha M, Hough G, Lallone R, and Fogelman AM (2002) Oral administration of an Apo A-I mimetic peptide synthesized from D-amino acids dramatically reduces atherosclerosis in mice independent of plasma cholesterol. *Circulation* **105**:290–292.
- Navab M, Anantharamaiah GM, Hama S, Hough G, Reddy ST, Frank JS, Garber DW, Handattu S, and Fogelman AM (2005a) D-4F and statins synergize to render HDL antiinflammatory in mice and monkeys and cause lesion regression in old apolipoprotein E-null mice. *Arterioscler Thromb Vasc Biol* **25**:1426–1432.
- Navab M, Anantharamaiah GM, Reddy ST, Hama S, Hough G, Frank JS, Grijalva VR, Ganesh VK, Mishra VK, Palgunachari MN, et al. (2005) Oral small peptides render HDL antiinflammatory in mice and monkeys and reduce atherosclerosis in ApoE null mice. *Circ Res* **97**:524–532.
- Navab M, Anantharamaiah GM, Reddy ST, Hama S, Hough G, Grijalva VR, Wagner AC, Frank JS, Datta G, Garber D, et al. (2004a) Oral D-4F causes formation of pre- $\beta$  high-density lipoprotein and improves high-density lipoprotein-mediated cholesterol efflux and reverse cholesterol transport from macrophages in apolipoprotein E-null mice. *Circulation* **109**:3215–3220.
- Navab M, Anantharamaiah GM, Reddy ST, Hama S, Hough G, Grijalva VR, Yu N, Ansell BJ, Datta G, Garber DW, et al. (2005b) Apolipoprotein A-I mimetic peptides. *Arterioscler Thromb Vasc Biol* **25**:1335–1341.
- Navab M, Anantharamaiah GM, Reddy ST, Van Lenten BJ, Wagner AC, Hama S, Hough G, Garber DW, Mishra VK, Palgunachari MN, et al. (2005c) An oral ApoJ peptide renders HDL antiinflammatory in mice and monkeys and dramatically reduces atherosclerosis in apolipoprotein E-null mice. *Arterioscler Thromb Vasc Biol* **25**:1932–1937.
- Navab M, Anantharamaiah GM, Reddy ST, Van Lenten BJ, Ansell BJ, Fonarow GC, Vahabzadeh K, Hama S, Hough G, et al. (2004b) The oxidation hypothesis of atherogenesis: the role of oxidized phospholipids and HDL. *J Lipid Res* **45**:993–1007.
- Navab M, Berliner JA, Subbanagounder G, Hama S, Lusis AJ, Castellani LW, Reddy S, Shih D, Shi W, Watson AD, et al. (2001a) HDL and the inflammatory response induced by LDL-derived oxidized phospholipids. *Arterioscler Thromb Vasc Biol* **21**:481–488.
- Navab M, Berliner JA, Watson AD, Hama SY, Territo MC, Lusis AJ, Shih DM, Van Lenten BJ, Frank JS, Demer LL, et al. (1996) The yin and yang of oxidation in the development of the fatty streak: A review based on the 1994 George Lyman Duff Memorial Lecture. *Arterioscler Thromb Vasc Biol* **16**:831–842.
- Navab M, Hama LS, Van LB, Fonarow GC, Cardinez CJ, Castellani LW, Brennan ML, Lusis AJ, and Fogelman AM (1997) Mildly oxidized LDL induces an increased apolipoprotein J/paraoxonase ratio. *J Clin Invest* **99**:2005–2019.
- Navab M, Hama SY, Anantharamaiah GM, Hassan K, Hough GP, Watson AD, Reddy ST, Sevanian A, Fonarow GC, and Fogelman AM (2000a) Normal high density lipoprotein inhibits three steps in the formation of mildly oxidized low density lipoprotein: steps 2 and 3. *J Lipid Res* **41**:1495–1508.
- Navab M, Hama SY, Cooke CJ, Anantharamaiah GM, Chaddha M, Jin L, Subbanagounder G, Faulk KF, Reddy ST, Miller NE, et al. (2000b) Normal high density lipoprotein inhibits three steps in the formation of mildly oxidized low density lipoprotein: step 1. *J Lipid Res* **41**:1481–1494.
- Navab M, Hama SY, Hough GP, Subbanagounder G, Reddy ST, and Fogelman AM (2001b) A cell-free assay for detecting HDL that is dysfunctional in preventing the formation of or inactivating oxidized phospholipids. *J Lipid Res* **42**:1308–1317.
- Newton RS and Krause BR (2002) HDL therapy for the acute treatment of atherosclerosis. *Atheroscler Suppl* **3**:31–38.
- Ng DS, Vezina C, Wolever TS, Kuksis A, Hegele RA, and Connelly PW (1995) Apolipoprotein A-I deficiency: biochemical and metabolic characteristics. *Arterioscler Thromb Vasc Biol* **15**:2157–2164.
- Nicholls SJ, Cutri B, Worthley SG, Kee P, Rye K-A, Bao S, and Barter PJ (2005a) Impact of short-term administration of high-density lipoproteins and atorvastatin on atherosclerosis in rabbits. *Arterioscler Thromb Vasc Biol* **25**:2416–2421.
- Nicholls SJ, Dusting GJ, Cutri B, Bao S, Drummond GR, Rye KA, and Barter PJ (2005b) Reconstituted high-density lipoproteins inhibit the acute pro-oxidant and proinflammatory vascular changes induced by a periarterial collar in normocholesterolemic rabbits. *Circulation* **111**:1543–1550.
- Nicholls SJ, Rye KA, and Barter PJ (2005c) High-density lipoproteins as therapeutic targets. *Curr Opin Lipidol* **16**:345–349.
- Nicholls SJ, Zheng L, and Hazen SL (2005d) Formation of dysfunctional high-density lipoprotein by myeloperoxidase. *Trends Cardiovasc Med* **15**:212–219.

- Nissen SE, Tsunoda T, Tuzcu EM, Schoenhagen P, Cooper CJ, Yasin M, Eaton GM, Lauer MA, Sheldon WS, Grines CL, et al. (2003) Effect of recombinant ApoA-I Milano on coronary atherosclerosis in patients with acute coronary syndromes: a randomized controlled trial. *J Am Med Assoc* **290**:2292–2300.
- Nissen SE, Tuzcu EM, Schoenhagen P, Brown BG, Ganz P, Vogel RA, Crowe T, Howard G, Cooper CJ, Brodie B, et al. (2004) Effect of intensive compared with moderate lipid-lowering therapy on progression of coronary atherosclerosis: a randomized controlled trial. *J Am Med Assoc* **291**:1071–1080.
- Nobecourt E, Jacqueminet S, Hansel B, Chantepie S, Grimaldi A, Chapman MJ, and Kontush A (2005) Defective antioxidative activity of small, dense HDL particles in type 2 diabetes: relationship to elevated oxidative stress and hyperglycemia. *Diabetologia* **48**:529–538.
- Nofer J-R and Assmann G (2005) Atheroprotective effects of high-density lipoprotein-associated lysosphingolipids. *Trends Cardiovasc Med* **15**:265–271.
- Nofer JR, Kehrel B, Fobker M, Levkau B, Assmann G, and von Eckardstein A (2002) HDL and arteriosclerosis: beyond reverse cholesterol transport. *Atherosclerosis* **161**:1–16.
- Nofer JR, van der Giet M, Tolle M, Wolinska I, von Wnuck Lipinski K, Baba HA, Tietge UJ, Godecke A, Ishii I, Kleuser B, et al. (2004) HDL induces NO-dependent vasorelaxation via the lysosphospholipid receptor S1P3. *J Clin Invest* **113**:569–581.
- Norata GD, Callegari E, Inoue H, and Catapano AL (2004) HDL3 induces cyclooxygenase-2 expression and prostacyclin release in human endothelial cells via a p38 MAPK/CRE-dependent pathway: effects on COX-2/PGL-synthase coupling. *Arterioscler Thromb Vasc Biol* **24**:871–877.
- Norata GD, Callegari E, Marchesi M, Chiesa G, Eriksson P, and Catapano AL (2005) High-density lipoproteins induce transforming growth factor- $\beta$ 2 expression in endothelial cells. *Circulation* **111**:2805–2811.
- O'Brien KD, McDonald TO, Kunjathoor V, Eng K, Knopp EA, Lewis K, Lopez R, Kirk EA, Chait A, Wight TN, et al. (2005) Serum amyloid A and lipoprotein retention in murine models of atherosclerosis. *Arterioscler Thromb Vasc Biol* **25**:785–790.
- Ogasawara K, Mashiba S, Wada Y, Sahara M, Uchida K, Aizawa T, and Kodama T (2004) A serum amyloid A and LDL complex as a new prognostic marker in stable coronary artery disease. *Atherosclerosis* **174**:349–356.
- Ohta T, Nakamura R, Ikeda Y, Shinohara M, Miyazaki A, Horiuchi S, and Matsuda I (1992) Differential effect of subspecies of lipoprotein containing apolipoprotein A-I on cholesterol efflux from cholesterol-loaded macrophages: functional correlation with lecithin: cholesterol acyltransferase. *Biochim Biophys Acta* **1165**:119–128.
- Ohta T, Saku K, Takata K, Nakamura R, Ikeda Y, and Matsuda I (1995) Different effects of subclasses of HDL containing apoA-I but not apoA-II (LpA-I) on cholesterol esterification in plasma and net cholesterol efflux from foam cells. *Arterioscler Thromb Vasc Biol* **15**:956–962.
- Okazaki M, Usui S, Ishigami M, Sakai N, Nakamura T, Matsuzawa Y, and Yamashita S (2005) Identification of unique lipoprotein subclasses for visceral obesity by component analysis of cholesterol profile in high-performance liquid chromatography. *Arterioscler Thromb Vasc Biol* **25**:578–584.
- Olsson AG, Schwartz GG, Szarek M, Sasiela WJ, Ezekowitz MD, Ganz P, Oliver MF, Waters D, and Zeiher, (2005) High-density lipoprotein, but not low-density lipoprotein cholesterol levels influence short-term prognosis after acute coronary syndrome: results from the MIRACL trial. *Eur Heart J* **26**:890–896.
- Oram JF (2000) Tangier disease and ABCA1. *Biochim Biophys Acta* **1529**:321–330.
- Oram JF (2002) The cholesterol mobilizing transporter ABCA1 as a new therapeutic target for cardiovascular disease. *Trends Cardiovasc Med* **12**:170–175.
- Oram JF, Lawn RM, Garvin MR, and Wade DP (2000) ABCA1 is the cAMP-inducible apolipoprotein receptor that mediates cholesterol secretion from macrophages. *J Biol Chem* **275**:34508–34511.
- Ostos MA, Conconi M, Vergnes L, Barouk N, Ribalta J, Girona J, Caillaud JM, Ochoa A, and Zakin MM (2001) Antioxidative and antiatherosclerotic effects of human apolipoprotein A-IV in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* **21**:1023–1028.
- Otvos JD, Shalaurova I, Freedman DS, and Rosenson RS (2002) Effects of pravastatin treatment on lipoprotein subclass profiles and particle size in the PLAC-I trial. *Atherosclerosis* **160**:41–48.
- Ou Z, Ou Z, Jones DW, Holzhauser S, Hatoum OA, Ackerman AW, Wehrauch DW, Gutterman DD, Guice K, Oldham KT, et al. (2003) L-4F, an apolipoprotein A-1 mimetic, dramatically improves vasodilation in hypercholesterolemia and sickle cell disease. *Circulation* **107**:2337–2341.
- Ou Z, Ou J, Ackerman AW, Oldham KT, and Pritchard KA Jr (2003) L-4F, an apolipoprotein A-1 mimetic, restores nitric oxide and superoxide anion balance in low-density lipoprotein-treated endothelial cells. *Circulation* **107**:1520–1524.
- Pajkrt D, Doran JE, Koster F, Lerch PG, Arnet B, van der Poll T, ten Cate JW, and van Deventer SJ (1996) Antiinflammatory effects of reconstituted high-density lipoprotein during human endotoxemia. *J Exp Med* **184**:1601–1608.
- Palmer AM, Murphy N, and Graham A (2004) Triglyceride-rich lipoproteins inhibit cholesterol efflux to apolipoprotein (apo) A1 from human macrophage foam cells. *Atherosclerosis* **173**:27–38.
- Panzenbock U and Stocker R (2005) Formation of methionine sulfoxide-containing specific forms of oxidized high-density lipoproteins. *Biochim Biophys Acta* **1703**:171–181.
- Parks JS and Rudel LL (1985) Alteration of high density lipoprotein subfraction distribution with induction of serum amyloid A protein (SAA) in the nonhuman primate. *J Lipid Res* **26**:82–91.
- Parthasarathy S, Barnett J, and Fong LG (1990) High-density lipoprotein inhibits the oxidative modification of low-density lipoprotein. *Biochim Biophys Acta* **1044**:275–283.
- Personius BE, Brown BG, and Gotto AM Jr (1998) Effects of increasing HDL-C, lowering triglyceride and lowering LDL-C on "fixed" atherosclerotic coronary artery disease: AFREGS (the Armed Forces Regression study). *Circulation* **98**:1450–1451.
- Pirro M, Siepi D, Lupattelli G, Roscini AR, Schillaci G, Gemelli F, Vaudo G, Marchesi S, Pasqualini L, and Mannarino E (2003) Plasma C-reactive protein in subjects with hypo/hyperalphalipoproteinemias. *Metabolism* **52**:432–436.
- Pruzanski W, Stefanski E, de Beer FC, de Beer MC, Ravandi A, and Kuksis A (2000) Comparative analysis of lipid composition of normal and acute-phase high density lipoproteins. *J Lipid Res* **41**:1035–1047.
- Pruzanski W, Vadas P, and Browning J (1993) Secretory non-pancreatic group II phospholipase A2: role in physiologic and inflammatory processes. *J Lipid Mediat* **8**:161–167.
- Pussinen PJ, Jauhainen M, Vilkkuna-Rautiainen T, Sundvall J, Vesanen M, Mattila K, Palosuo T, Alfthan G, and Asikainen S (2004) Periodontitis decreases the antiatherogenic potency of high density lipoprotein. *J Lipid Res* **45**:139–147.
- Quarck R, De Geest B, Stengel D, Mertens A, Lox M, Theilmeier G, Michiels C, Raes M, Bult H, Collen D, et al. (2001) Adenovirus-mediated gene transfer of human platelet-activating factor-acetylhydrolase prevents injury-induced neointima formation and reduces spontaneous atherosclerosis in apolipoprotein E-deficient mice. *Circulation* **103**:2495–2500.
- Raffai RL, Loeb SM, and Weisgraber KH (2005) Apolipoprotein E promotes the regression of atherosclerosis independently of lowering plasma cholesterol levels. *Arterioscler Thromb Vasc Biol* **25**:436–441.
- Rashid S, Barrett PH, Uffelman KD, Watanabe T, Adeli K, and Lewis GF (2002) Lipolytically modified triglyceride-enriched HDLs are rapidly cleared from the circulation. *Arterioscler Thromb Vasc Biol* **22**:483–487.
- Rashid S, Watanabe T, Sakaue T, and Lewis GF (2003) Mechanisms of HDL lowering in insulin resistant, hypertriglyceridemic states: the combined effect of HDL triglyceride enrichment and elevated hepatic lipase activity. *Clin Biochem* **36**:421–429.
- Recalde D, Ostos MA, Badell E, Garcia-Otin AL, Pidoux J, Castro G, Zakin MM, and Scott-Algara D (2004) Human apolipoprotein A-IV reduces secretion of proinflammatory cytokines and atherosclerotic effects of a chronic infection mimicked by lipopolysaccharide. *Arterioscler Thromb Vasc Biol* **24**:756–761.
- Reschly EJ, Sorci-Thomas MG, Davidson WS, Meredith SC, Reardon CA, and Getz GS (2002) Apolipoprotein A-I  $\alpha$ -helices 7 and 8 modulate high density lipoprotein subclass distribution. *J Biol Chem* **277**:9645–9654.
- Ribas V, Sanchez-Quesada JL, Anton R, Camacho M, Julve J, Escola-Gil JC, Vila L, Ordonez-Llanos J, and Blanco-Vaca F (2004) Human apolipoprotein A-II enrichment displaces paraoxonase from HDL and impairs its antioxidant properties: a new mechanism linking HDL protein composition and antiatherogenic potential. *Circ Res* **95**:789–797.
- Ridker PM, Brown NJ, Vaughan DE, Harrison DG, and Mehta JL (2004a) Established and emerging plasma biomarkers in the prediction of first atherothrombotic events. *Circulation* **109**:IV-6–IV-19.
- Ridker PM, Hennekens CH, Buring JE, and Rifai N (2000) C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N Engl J Med* **342**:836–843.
- Ridker PM, Wilson PWF, and Grundy SM (2004b) Should C-reactive protein be added to metabolic syndrome and to assessment of global cardiovascular risk? *Circulation* **109**:2818–2825.
- Rifici VA and Khachadurian AK (1996) Effects of dietary vitamin C and E supplementation on the copper mediated oxidation of HDL and on HDL mediated cholesterol efflux. *Atherosclerosis* **127**:19–26.
- Robbesyn F, Garcia V, Auge N, Vieira O, Frisach, MF, Salvayre R, and Negre-Salvayre A (2003) HDL counterbalance the proinflammatory effect of oxidized LDL by inhibiting intracellular reactive oxygen species rise, proteasome activation, and subsequent NF- $\kappa$ B activation in smooth muscle cells. *FASEB J* **17**:743–745.
- Robins SJ, Collins D, Wittes JT, Papademetriou V, Deedwania PC, Schaefer EJ, McNamara JR, Kashyap ML, Hershman JM, Wexler LF, et al. (2001) Relation of gemfibrozil treatment and lipid levels with major coronary events: VA-HIT: a randomized controlled trial. *J Am Med Assoc* **285**:1585–1591.
- Rosenblat M, Vaya J, Shih D, and Aviram M (2005) Paraoxonase 1 (PON1) enhances HDL-mediated macrophage cholesterol efflux via the ABCA1 transporter in association with increased HDL binding to the cells: a possible role for lysophosphatidylcholine. *Atherosclerosis* **179**:69–77.
- Rosenon RS (2003) Antiatherothrombotic effects of nicotinic acid. *Atherosclerosis* **171**:87–96.
- Rosenon RS, Otvos JD, and Freedman DS (2002) Relations of lipoprotein subclass levels and low-density lipoprotein size to progression of coronary artery disease in the Pravastatin Limitation of Atherosclerosis in the Coronary Arteries (PLAC-I) trial. *Am J Cardiol* **90**:89–94.
- Rothblat GH, de la Llera-Moya M, Atger V, Kellner-Weibel G, Williams DL, and Phillips MC (1999) Cell cholesterol efflux: integration of old and new observations provides new insights. *J Lipid Res* **40**:781–796.
- Rotllan N, Ribas V, Calpe-Berdiel L, Martin-Campos JM, Blanco-Vaca F, and Escola-Gil JC (2005) Overexpression of human apolipoprotein A-II in transgenic mice does not impair macrophage-specific reverse cholesterol transport in vivo. *Arterioscler Thromb Vasc Biol* **25**:e128–e132.
- Rubic T, Trottmann M, and Lorenz RL (2004) Stimulation of CD36 and the key effector of reverse cholesterol transport ATP-binding cassette A1 in monocyte cells by niacin. *Biochem Pharmacol* **67**:411–419.
- Rye KA and Barter PJ (2004) Formation and metabolism of prebeta-migrating, lipid-poor apolipoprotein A-I. *Arterioscler Thromb Vasc Biol* **24**:421–428.
- Sacks FM, Brewer HB, Alaupovic P, Kostner G, Asztalos B, Schaefer E, Rothblat GH, Akeefe H, Conner A, Perlman T, et al. (2004) Selective plasma HDL delipidation and reinfusion: a unique new approach for acute HDL therapy in the treatment of cardiovascular disease, in *Proceedings of the American Heart Association Scientific Sessions*; 2004 Nov 7–10; New Orleans, LA.
- Sakai T, Kamanna VS, and Kashyap ML (2001) Niacin, but not gemfibrozil, selectively increases LP-AI, a cardioprotective subfraction of HDL, in patients with low HDL cholesterol. *Arterioscler Thromb Vasc Biol* **21**:1783–1789.
- Sampietro T, Bigazzi F, Dal Pino B, Fusaro S, Greco F, Tuoni M, and Bionda A (2002)

- Increased plasma C-reactive protein in familial hypoalphalipoproteinemia: a proinflammatory condition? *Circulation* **105**:11–14.
- Sanguinetti SM, Brites FD, Fasulo V, Verona J, Elbert A, Wikinski RL, and Schreier LE (2001) HDL oxidizability and its protective effect against LDL oxidation in type 2 diabetic patients. *Diabetes Nutr Metab* **14**:27–36.
- Santamarina-Fojo S, Gonzalez-Navarro H, Freeman L, Wagner E, and Nong Z (2004) Hepatic lipase, lipoprotein metabolism, and atherogenesis. *Arterioscler Thromb Vasc Biol* **24**:1750–1754.
- Santamarina-Fojo S, Peterson K, Knapper C, Qiu Y, Freeman L, Cheng JF, Osorio J, Remaley A, Yang XP, Haudenschild C, et al. (2000) Complete genomic sequence of the human ABCA1 gene: analysis of the human and mouse ATP-binding cassette A promoter. *Proc Natl Acad Sci USA* **97**: 7987–92.
- Sasahara T, Nestel P, Fidge N, and Sviridov D (1998) Cholesterol transport between cells and high density lipoprotein subfractions from obese and lean subjects. *J Lipid Res* **39**:544–554.
- Sasahara T, Yamashita T, Sviridov D, Fidge N, and Nestel P (1997) Altered properties of high density lipoprotein subfractions in obese subjects. *J Lipid Res* **38**:600–611.
- Sasaki J, Yamamoto K, and Ageta M (2002) Effects of fenofibrate on high-density lipoprotein particle size in patients with hyperlipidemia: a randomized, double-blind, placebo-controlled, multicenter, crossover study. *Clin Ther* **24**:1614–1626.
- Sattler W, Maiorino M, and Stocker R (1994) Reduction of HDL- and LDL-associated cholesteryl ester and phospholipid hydroperoxides by phospholipid hydroperoxide glutathione peroxidase and Ebselen (PZ 51). *Arch Biochem Biophys* **309**:214–221.
- Schaefer EJ, McNamara JR, Asztalos BF, Taylor T, Daly JA, Gleason JL, Seman LJ, Ferrari A, and Rubenstein JJ (2005) Effects of atorvastatin versus other statins on fasting and postprandial C-reactive protein and lipoprotein-associated phospholipase A2 in patients with coronary heart disease versus control subjects. *Am J Cardiol* **95**:1025–1032.
- Schaefer EJ, McNamara JR, Taylor T, Daly JA, Gleason JA, Seman LJ, Ferrari A, and Rubenstein JJ (2002) Effects of atorvastatin on fasting and postprandial lipoprotein subclasses in coronary heart disease patients versus control subjects. *Am J Cardiol* **90**:689–696.
- Schaefer JR, Rader DJ, Ikwaki K, Fairwell T, Zech LA, Kindt MR, Davignon J, Gregg RE, and Brewer HB Jr (1992) In vivo metabolism of apolipoprotein A-I in a patient with homozygous familial hypercholesterolemia. *Arterioscler Thromb* **12**: 843–848.
- Schaefer JR, Schweer H, Ikwaki K, Stracke H, Seyberth HJ, Kaffarnik H, Maisch B, and Steinmetz A (1999) Metabolic basis of high density lipoproteins and apolipoprotein A-I increase by HMG-CoA reductase inhibition in healthy subjects and a patient with coronary artery disease. *Atherosclerosis* **144**:177–184.
- Schilling M, Exner M, Mlekusch W, Sabeti S, Amighi J, Nikowitsch R, Timmel E, Kicking B, Minar C, Pones M, et al. (2005) Inflammation and Carotid Artery-Risk for Atherosclerosis Study (ICARAS). *Circulation* **111**:2203–2209.
- Schwedhelm E, Bartling A, Lenzen H, Tsikas D, Maas R, Brummer J, Gutzki FM, Berger J, Frolich JC, and Boger RH (2004) Urinary 8-iso-prostaglandin F<sub>2α</sub> as a risk marker in patients with coronary heart disease: a matched case-control study. *Circulation* **109**:843–848.
- Segrest JP, Harvey SC, and Zannis V (2000) Detailed molecular model of apolipoprotein A-I on the surface of high-density lipoproteins and its functional implications. *Trends Cardiovasc Med* **10**:246–252.
- Segrest JP, Jones MK, Klon AE, Sheldahl CJ, Hellinger M, De Loof H, and Harvey SC (1999) A detailed molecular belt model for apolipoprotein A-I in discoidal high density lipoprotein. *J Biol Chem* **274**:31755–31758.
- Shah PK, Nilsson J, Kaul S, Fishbein MC, Ageland H, Hamsten A, Johansson J, Karpe F, and Cercek B (1998) Effects of recombinant apolipoprotein A-I (Milano) on aortic atherosclerosis in apolipoprotein E-deficient mice. *Circulation* **97**:780–785.
- Shah PK, Yano J, Reyes O, Chyu KY, Kaul S, Bisgaier CL, Drake S, and Cercek B (2001) High-dose recombinant apolipoprotein A-I (Milano) mobilizes tissue cholesterol and rapidly reduces plaque lipid and macrophage content in apolipoprotein E-deficient mice: potential implications for acute plaque stabilization. *Circulation* **103**:3047–3050.
- Shao B, Bergt C, Fu X, Green P, Voss JC, Oda MN, Oram JF, and Heinecke JW (2005a) Tyrosine 192 in apolipoprotein A-I is the major site of nitration and chlorination by myeloperoxidase, but only chlorination markedly impairs ABCA1-dependent cholesterol transport. *J Biol Chem* **280**:5983–5993.
- Shao B, Fu X, McDonald TO, Green PS, Uchida K, O'Brien K, D, Oram JF, and Heinecke JW (2005b) Acrolein impairs ABCA1-dependent cholesterol export from cells through site-specific modification of apolipoprotein A-I. *J Biol Chem* **280**: 38386–38396.
- Shao B, O'Brien K, D, McDonald TO, Fu X, Oram JF, Uchida K, and Heinecke JW (2005c) Acrolein modifies apolipoprotein A-I in the human artery wall. *Ann N Y Acad Sci* **1043**:396–403.
- Sharrett AR, Ballantyne CM, Coady SA, Heiss G, Sorlie PD, Catellier D, and Patsch W (2001) Coronary heart disease prediction from lipoprotein cholesterol levels, triglycerides, lipoprotein (a), apolipoproteins A-I and B, and HDL density subfractions: the Atherosclerosis Risk in Communities (ARIC) Study. *Circulation* **104**: 1108–1113.
- Shih DM, Gu L, Hama S, Xia YR, Navab M, Fogelman AM, and Lusis AJ (1996) Genetic-dietary regulation of serum paraoxonase expression and its role in atherosclerosis in a mouse model. *J Clin Invest* **97**:1630–1639.
- Shih DM, Gu L, Xia YR, Navab M, Li WF, Hama S, Castellani LW, Furlong CE, Costa LG, Fogelman AM, et al. (1998) Mice lacking serum paraoxonase are susceptible to organophosphate toxicity and atherosclerosis. *Nature (Lond)* **394**: 284–287.
- Silva RA, Hilliard GM, Li L, Segrest JP, and Davidson WS (2005) A mass spectrometric determination of the conformation of dimeric apolipoprotein A-I in discoidal high density lipoproteins. *Biochemistry* **44**:8600–8607.
- Sirtori CR, Calabresi L, and Franceschini G (1999) Recombinant apolipoproteins for the treatment of vascular diseases. *Atherosclerosis* **142**:29–40.
- Sirtori CR, Calabresi L, Franceschini G, Baldassarre D, Amato M, Johansson J, Salvetti M, Monteduro C, Zulli R, Muesan ML, et al. (2001) Cardiovascular status of carriers of the apolipoprotein A-I (Milano) mutant: the Limone sul Garda study. *Circulation* **103**:1949–1954.
- Skeggs JW and Morton RE (2002) LDL and HDL enriched in triglyceride promote abnormal cholesterol transport. *J Lipid Res* **43**:1264–1274.
- Skinner ER (1994) High-density lipoprotein subclasses. *Curr Opin Lipidol* **5**:241–247.
- Soderlund S, Soro-Paavonen A, Ehnholm C, Jauhiainen M, and Taskinen MR (2005) Hypertriglyceridemia is associated with pre-β-HDL concentrations in subjects with familial low HDL. *J Lipid Res* **46**:1643–1651.
- Soedamah-Muthu SS, Colhoun HM, Thomason MJ, Betteridge DJ, Durrington PN, Hitman GA, Fuller JH, Julier K, Mackness MI, and Neil HA (2003) The effect of atorvastatin on serum lipids, lipoproteins and NMR spectroscopy defined lipoprotein subclasses in type 2 diabetic patients with ischaemic heart disease. *Atherosclerosis* **167**:243–255.
- Soma MR, Donetti E, Parolini C, Sirtori CR, Fumagalli R, and Franceschini G (1995) Recombinant apolipoprotein A-I (Milano) dimer inhibits carotid intimal thickening induced by perivascular manipulation in rabbits. *Circ Res* **76**:405–411.
- Sparks DL, Davidson WS, Lund-Katz S, and Phillips MC (1995) Effects of the neutral lipid content of high density lipoprotein on apolipoprotein A-I structure and particle stability. *J Biol Chem* **270**:26910–26917.
- Spieker LE, Sudano I, Hurlimann D, Lerch PG, Lang MG, Binggeli C, Corti R, Ruschitzka F, Luscher TF, and Noll G (2002) High-density lipoprotein restores endothelial function in hypercholesterolemic men. *Circulation* **105**:1399–1402.
- Stein O and Stein Y (1999) Atheroprotective mechanisms of HDL. *Atherosclerosis* **144**:285–301.
- Steinberg D, Parthasarathy S, Carew TE, Khoo JC, and Witztum JL (1989) Beyond cholesterol. *N Engl J Med* **320**:915–924, 1989.
- Steinberg D and Witztum JL (2002) Is the oxidative modification hypothesis relevant to human atherosclerosis? Do the antioxidant trials conducted to date refute the hypothesis? *Circulation* **105**:2107–2111.
- Steiner G (2005) Fibrates and coronary risk reduction. *Atherosclerosis* **182**:199–207.
- Stocker R and Keane JF Jr (2004) Role of oxidative modifications in atherosclerosis. *Physiol. Rev.* **84**:1381–1478.
- Stoll LL, Denning GM, and Weintraub NL (2004) Potential role of endotoxin as a proinflammatory mediator of atherosclerosis. *Arterioscler Thromb Vasc Biol* **24**: 2227–2236.
- Suc I, Escargueil-Blanc I, Trolly M, Salvayre R, and Negre-Salvayre A (1997) HDL and ApoA prevent cell death of endothelial cells induced by oxidized LDL. *Arterioscler Thromb Vasc Biol* **17**:2158–2166.
- Sugano M, Sawada S, Tsuchida K, Makino N, and Kamada M (2000) Low density lipoproteins develop resistance to oxidative modification due to inhibition of cholesteryl ester transfer protein by a monoclonal antibody. *J Lipid Res* **41**:126–133.
- Sugano M, Tsuchida K, and Makino N (2000) High-density lipoproteins protect endothelial cells from tumor necrosis factor-α-induced apoptosis. *Biochem Biophys Res Commun* **272**:872–876.
- Suzuki M, Wada H, Maeda S, Saito K, Minatoguchi S, Saito K, and Seishima M (2005) Increased plasma lipid-poor apolipoprotein A-I in patients with coronary artery disease. *Clin Chem* **51**:132–137.
- Sviridov D, Miyazaki O, Theodore K, Hoang A, Fukamachi I, and Nestel P (2002) Delineation of the role of pre-β1-HDL in cholesterol efflux using isolated pre-β1-HDL. *Arterioscler Thromb Vasc Biol* **22**:1482–1488.
- Syvänne M, Ahola M, Lahdenpera S, Kahri J, Kuusi T, Virtanen KS, and Taskinen MR (1995) High density lipoprotein subfractions in non-insulin-dependent diabetes mellitus and coronary artery disease. *J Lipid Res* **36**:573–582.
- Takahashi Y and Smith JD (1999) Cholesterol efflux to apolipoprotein AI involves endocytosis and resecretion in a calcium-dependent pathway. *Proc Natl Acad Sci USA* **96**:11358–11363.
- Tall AR, Blum CB, Forester GP, and Nelson CA (1982) Changes in the distribution and composition of plasma high density lipoproteins after ingestion of fat. *J Biol Chem* **257**:198–207.
- Tamama K, Tomura H, Sato K, Malchinkhuu E, Damirin A, Kimura T, Kuwabara A, Murakami M, and Okajima F (2005) High-density lipoprotein inhibits migration of vascular smooth muscle cells through its sphingosine 1-phosphate component. *Atherosclerosis* **178**:19–23.
- Tan KCB, Chow W-S, Lam JCM, Lam B, Wong W-K, Tam S, and Ip MSM (2005) HDL dysfunction in obstructive sleep apnea. *Atherosclerosis* **184**:277–282.
- Tangirala RK, Pratico D, FitzGerald GA, Chun S, Tsukamoto K, Maugeais C, Usher DC, Pure E, and Rader DJ (2001) Reduction of isoprostanol and regression of advanced atherosclerosis by apolipoprotein E. *J Biol Chem* **276**:261–266.
- Tato F, Vega GL, and Grundy SM (1995) Bimodal distribution of cholesteryl ester transfer protein activities in normotriglyceridemic men with low HDL cholesterol concentrations. *Arterioscler Thromb Vasc Biol* **15**:446–451.
- Taylor, AJ, Sullenberger, LE, Lee HJ, Lee JK, and Grace KA (2004) Arterial biology for the investigation of the treatment effects of reducing cholesterol (ARBITER) 2: a double-blind, placebo-controlled study of extended-release niacin on atherosclerosis progression in secondary prevention patients treated with statins. *Circulation* **110**: 3512–3517.
- Teiber JF, Draganov DI, and La Du BN (2004) Purified human serum PON1 does not protect LDL against oxidation in the in vitro assays initiated with copper or AAPH. *J Lipid Res* **45**: 2260–2268.
- The Bezafibrate Infarction Prevention (BIP) Study (2000) Secondary prevention by raising HDL cholesterol and reducing triglycerides in patients with coronary artery disease: the Bezafibrate Infarction Prevention (BIP) study. *Circulation* **102**:21–27.
- Theilmeier G, De Geest B, Van Veldhoven PP, Stengel D, Michiels C, Lox M, Landeloos M, Chapman MJ, Ninio E, Collen D, et al. (2000) HDL-associated PAF-AH reduces endothelial adhesiveness in apoE<sup>-/-</sup> mice. *FASEB J* **14**:2032–2039.
- Thorngate FE, Rudel LL, Walzem RL, and Williams DL (2000) Low levels of extra-



- hepatic nonmacrophage ApoE inhibit atherosclerosis without correcting hypercholesterolemia in ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* **20**:1939–1945.
- Thuahnai ST, Lund-Katz S, Dhanasekaran P, de la Llera-Moya M, Connelly MA, Williams DL, Rothblat GH, and Phillips MC (2004) Scavenger receptor class B type I-mediated cholesteryl ester-selective uptake and efflux of unesterified cholesterol. Influence of high density lipoprotein size and structure. *J Biol Chem* **279**:12448–12455. Epub 2004 Jan 12.
- Tietge UJ, Maugeais C, Lund-Katz S, Grass D, deBeer FC, and Rader DJ (2002) Human secretory phospholipase A2 mediates decreased plasma levels of HDL cholesterol and apoA-I in response to inflammation in human apoA-I transgenic mice. *Arterioscler Thromb Vasc Biol* **22**:1213–1218.
- Trougakos IP, Lourda M, Agiostratidou G, Kletsas D, and Gonos ES (2005) Differential effects of clusterin/apolipoprotein J on cellular growth and survival. *Free Radic Biol Med* **38**:436–449.
- Tselepis AD and Chapman MJ (2002) Inflammation, bioactive lipids and atherosclerosis: potential roles of a lipoprotein-associated phospholipase A2, platelet activating factor-acetylhydrolase. *Atheroscler Suppl* **3**:57–68.
- Tsimihodimos V, Kakafika A, Tambaki AP, Bairaktari E, Chapman MJ, Elisaf M, and Tselepis AD (2003) Fenofibrate induces HDL-associated PAF-AH but attenuates enzyme activity associated with apoB-containing lipoproteins. *J Lipid Res* **44**:927–934.
- Tsimihodimos V, Karabina SA, Tambaki AP, Bairaktari E, Miltiadous G, Goudevanos JA, Cariolou MA, Chapman MJ, Tselepis AD, and Elisaf M (2002) Altered distribution of platelet-activating factor-acetylhydrolase activity between LDL and HDL as a function of the severity of hypercholesterolemia. *J Lipid Res* **43**:256–263.
- Tunaru S, Kero J, Schaub A, Wufka C, Blaukat A, Pfeffer K, and Offermanns S (2003) PUMA-G and HM74 are receptors for nicotinic acid and mediate its anti-lipolytic effect. *Nat Med* **9**:352–355.
- Tward A, Xia YR, Wang XP, Shi YS, Park C, Castellani LW, Lusis AJ, and Shih DM (2002) Decreased atherosclerotic lesion formation in human serum paraoxonase transgenic mice. *Circulation* **106**:484–490.
- Uhlir CM and Whitehead AS (1999) Serum amyloid A, the major vertebrate acute-phase reactant. *Eur J Biochem* **265**:501–523.
- Uint L, Sposito A, Brandizzi LI, Yoshida VM, Maranhao RC, and Luz PL (2003) Cellular cholesterol efflux mediated by HDL isolated from subjects with low HDL levels and coronary artery disease. *Arq Bras Cardiol* **81**:39–41, 35–38.
- van Dam MJ, de Groot E, Clee SM, Hovingh GK, Roelants R, Brooks-Wilson A, Zwinderman AH, Smit AJ, Smelt AHM, and Groen AK (2002) Association between increased arterial-wall thickness and impairment in ABCA1-driven cholesterol efflux: an observational study. *Lancet* **359**:37–42.
- van der Steeg WA, Kuivenhoven JA, Klerkx AH, Boekholdt SM, Hovingh GK, and Kastelein JJ (2004) Role of CETP inhibitors in the treatment of dyslipidemia. *Curr Opin Lipidol* **15**:631–636.
- van der Westhuyzen DR, Cai L, de Beer MC, and de Beer FC (2005) SAA promotes cholesterol efflux mediated by SR-BI. *J Biol Chem* **280**:35890–35895.
- Van Lenten BJ, Hama SY, de Beer FC, Stafforini DM, McIntyre TM, Prescott SM, La Du BN, Fogelman AM, and Navab M (1995) Anti-inflammatory HDL becomes pro-inflammatory during the acute phase response: loss of protective effect of HDL against LDL oxidation in aortic wall cell cocultures. *J Clin Invest* **96**:2758–2767.
- Van Lenten BJ, Navab M, Shih D, Fogelman AM, and Lusis AJ (2001a) The role of high-density lipoproteins in oxidation and inflammation. *Trends Cardiovasc Med* **11**:155–161.
- Van Lenten BJ, Wagner AC, Nayak DP, Hama S, Navab M, and Fogelman AM (2001b) High-density lipoprotein loses its anti-inflammatory properties during acute influenza infection. *Circulation* **103**:2283–2288.
- van Tol A (2002) Phospholipid transfer protein. *Curr Opin Lipidol* **13**:135–139.
- Vaughan AM and Oram JF (2005) ABCG1 redistributes cell cholesterol to domains removable by high density lipoprotein but not by lipid-depleted apolipoproteins. *J Biol Chem* **280**:30150–30157.
- Verma S, Szmitko PE, and Ridker PM (2005) C-reactive protein comes of age. *Nat Clin Pract Cardiovasc Med* **2**:29–36.
- von Eckardstein A, Nofer JR, and Assmann G (2001) High density lipoproteins and arteriosclerosis: role of cholesterol efflux and reverse cholesterol transport. *Arterioscler Thromb Vasc Biol* **21**:13–27.
- Wang N, Lan D, Chen W, Matsuura F, and Tall AR (2004) ATP-binding cassette transporters G1 and G4 mediate cellular cholesterol efflux to high-density lipoproteins. *Proc Natl Acad Sci USA* **101**:9774–9779.
- Wang N, Silver DL, Costet P, and Tall AR (2000) Specific binding of ApoA-I, enhanced cholesterol efflux, and altered plasma membrane morphology in cells expressing ABC1. *J Biol Chem* **275**:33053–33058.
- Warden CH, Hedrick CC, Qiao JH, Castellani LW, and Lusis AJ (1993) Atherosclerosis in transgenic mice overexpressing apolipoprotein A-II. *Science (Wash DC)* **261**:469–472.
- Watson AD, Berliner JA, Hama SY, La Du BN, Faull KF, Fogelman AM, and Navab M (1995a) Protective effect of high density lipoprotein associated paraoxonase: inhibition of the biological activity of minimally oxidized low density lipoprotein. *J Clin Invest* **96**:2882–2891.
- Watson AD, Navab M, Hama SY, Sevanian A, Prescott SM, Stafforini DM, McIntyre TM, Du BN, Fogelman AM, and Berliner JA (1995b) Effect of platelet activating factor-acetylhydrolase on the formation and action of minimally oxidized low density lipoprotein. *J Clin Invest* **95**:774–782.
- Willerson JT and Ridker PM (2004) Inflammation as a cardiovascular risk factor. *Circulation* **109**:II-2–II-10.
- Wilson PW (1990) High-density lipoprotein, low-density lipoprotein and coronary artery disease. *Am J Cardiol* **66**:7A–10A.
- Winkler K, Winkelmann BR, Scharnagl H, Hoffmann MM, Grawitz AB, Nauck M, Bohm BO, and Marz W (2005) Platelet-activating factor acetylhydrolase activity indicates angiographic coronary artery disease independently of systemic inflammation and other risk factors: the Ludwigshafen Risk and Cardiovascular Health Study. *Circulation* **111**:980–987.
- Wise A, Foord SM, Fraser NJ, Barnes AA, Elshourbagy N, Eilert M, Ignar DM, Murdock PR, Steplewski K, Green A, et al. (2003) Molecular identification of high and low affinity receptors for nicotinic acid. *J Biol Chem* **278**:9869–9874.
- Wolfe ML and Rader DJ (2004) Cholesteryl ester transfer protein and coronary artery disease: an observation with therapeutic implications. *Circulation* **110**:1338–1340.
- World Health Organization (2004) *The atlas of heart disease and stroke*, World Health Organization, Geneva, Switzerland
- Xia P, Vadas MA, Rye K-A, Barter PJ, and Gamble JR (1999) High density lipoproteins (HDL) interrupt the sphingosine kinase signaling pathway: a possible mechanism for protection against atherosclerosis by HDL. *J Biol Chem* **274**:33143–33147.
- Xu Y and Fu M (2003) Alterations of HDL subclasses in hyperlipidemia. *Clin Chim Acta* **332**:95–102.
- Yamada T, Kakiyama T, Kamishima T, Fukuda T, and Kawai T (1996) Both acute phase and constitutive serum amyloid A are present in atherosclerotic lesions. *Pathol Int* **46**:797–800.
- Yamashita S, Maruyama T, Hirano K, Sakai N, Nakajima N, and Matsuzawa Y (2000) Molecular mechanisms, lipoprotein abnormalities and atherogenicity of hyperalphalipoproteinemia. *Atherosclerosis* **152**:271–285.
- Yan D, Navab M, Bruce C, Fogelman AM, and Jiang XC (2004) PLTP deficiency improves the anti-inflammatory properties of HDL and reduces the ability of LDL to induce monocyte chemotactic activity. *J Lipid Res* **45**:1852–1858.
- Yancey PG, Asztalos BF, Stettler N, Piccoli D, Williams DL, Connelly MA, and Rothblat GH (2004) SR-BI- and ABCA1-mediated cholesterol efflux to serum from patients with Alagille syndrome. *J Lipid Res* **45**:1724–1732.
- Yancey PG, de la Llera-Moya M, Swarnakar S, Monzo P, Klein SM, Connelly MA, Johnson WJ, Williams DL, and Rothblat GH (2000) High density lipoprotein phospholipid composition is a major determinant of the bi-directional flux and net movement of cellular free cholesterol mediated by scavenger receptor BI. *J Biol Chem* **275**:36596–36604.
- Yang Y, Yan B, Fu M, Xu Y, and Tian Y (2005) Relationship between plasma lipid concentrations and HDL subclasses. *Clin Chim Acta* **354**:49–58.
- Yoshikawa M, Sakuma N, Hibino T, Sato T, and Fujinami T (1997) HDL3 exerts more powerful anti-oxidative, protective effects against copper-catalyzed LDL oxidation than HDL2. *Clin Biochem* **30**:221–225.
- Yu S, Yarnell JW, Sweetnam P, and Bolton CH (2003) High density lipoprotein subfractions and the risk of coronary heart disease: 9-years follow-up in the Caerphilly Study. *Atherosclerosis* **166**:331–338.
- Yuhanna IS, Zhu Y, Cox BE, Hahner LD, Osborne-Lawrence S, Lu P, Marcel YL, Anderson RG, Mendelsohn ME, Hobbs HH, et al. (2001) High-density lipoprotein binding to scavenger receptor-BI activates endothelial nitric oxide synthase. *Nat Med* **7**:853–857.
- Yusuf S, Hawken S, Ounpuu S, Dans T, Avezum A, Lanas F, McQueen M, Budaj A, Pais P, Varigos J, et al. (2004) Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. *Lancet* **364**:937–952.
- Zago V, Sanguinetti S, Brites F, Berg G, Verona J, Basilio F, Wikinski R, and Schreier L (2004) Impaired high density lipoprotein antioxidant activity in healthy postmenopausal women. *Atherosclerosis* **177**:203–210.
- Zalewski A and Macphee C (2005) Role of lipoprotein-associated phospholipase A2 in atherosclerosis: biology, epidemiology, and possible therapeutic target. *Arterioscler Thromb Vasc Biol* **25**:923–931.
- Zhang B, Fan P, Shimoji E, Xu H, Takeuchi K, Bian C, and Saku K (2004) Inhibition of cholesteryl ester transfer protein activity by JTT-705 increases apolipoprotein E-containing high-density lipoprotein and favorably affects the function and enzyme composition of high-density lipoprotein in rabbits. *Arterioscler Thromb Vasc Biol* **24**:1910–1915.
- Zhang B, Tomura H, Kuwabara A, Kimura T, Miura S-i, Noda K, Okajima F, and Saku K (2005) Correlation of high density lipoprotein (HDL)-associated sphingosine 1-phosphate with serum levels of HDL-cholesterol and apolipoproteins. *Atherosclerosis* **178**:199–205.
- Zhang Y, Zanotti I, Reilly MP, Glick JM, Rothblat GH, and Rader DJ (2003) Overexpression of apolipoprotein A-I promotes reverse transport of cholesterol from macrophages to feces in vivo. *Circulation* **108**:661–663.
- Zheng L, Nukuna B, Brennan ML, Sun M, Goormastic M, Settle M, Schmitt D, Fu X, Thomson L, Fox PL, et al. (2004) Apolipoprotein A-I is a selective target for myeloperoxidase-catalyzed oxidation and functional impairment in subjects with cardiovascular disease. *J Clin Invest* **114**:529–541.
- Zheng L, Settle M, Brubaker G, Schmitt D, Hazen SL, Smith JD, and Kinter M (2005) Localization of nitration and chlorination sites on apolipoprotein A-I catalyzed by myeloperoxidase in human atheroma and associated oxidative impairment in ABCA1-dependent cholesterol efflux from macrophages. *J Biol Chem* **280**:38–47.